

Glucocorticoids and foetal heart maturation; implications for prematurity and foetal programming

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Abstract

Glucocorticoids are steroid hormones, essential in mammals to prepare for life after birth. Blood levels of glucocorticoids (cortisol in most mammals including humans; corticosterone in rats and mice) rise dramatically shortly before birth. This is mimicked clinically in the routine administration of synthetic glucocorticoids to pregnant women threatened by a preterm birth or to preterm infants to improve neonatal survival. Whilst effects on lung are well documented and essential for postnatal survival, those on heart are less well known. In this study, we review recent evidence for a crucial role of glucocorticoids in late gestational heart maturation. Either insufficient or excessive glucocorticoid exposure before birth may alter the normal glucocorticoid-regulated trajectory of heart maturation with potential life-long consequences.

Key Words

- ▶ glucocorticoids
- ▶ cardiovascular
- ▶ development
- ▶ embryo/foetus
- ▶ adrenal hormones

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Introduction

Glucocorticoids are steroid hormones produced by the adrenal cortex. They regulate stress responses and exert potent immunomodulatory effects in adults (McEwen 1997) and are essential for the transition from foetal to neonatal life. Plasma levels of glucocorticoids rise markedly in late gestation, to mature foetal tissues and organs (Fowden *et al.* 1998) and this is mimicked clinically with the routine administration of synthetic glucocorticoids (betamethasone or dexamethasone) to pregnant women in whom preterm delivery is anticipated, to improve neonatal survival. Whilst there is no question that the enhancement of foetal lung maturity is life-saving, animal models and some studies on humans have raised concerns about long-term effects of synthetic glucocorticoid administration, particularly on the cardiovascular system. Although a number of studies have examined the mechanisms that underlie the effects of excess glucocorticoid exposure on the heart and vasculature, few have

addressed the maturational pathways normally regulated by glucocorticoids in the foetal heart and vasculature. In this study, we present a non-systematic review of the evidence for a critical role of glucocorticoids in late gestational heart maturation and discuss the implications of both inadequate and excess glucocorticoid action within the late gestation foetal heart.

Glucocorticoid receptors

Glucocorticoids bind to cytoplasmic receptors belonging to the nuclear receptor superfamily; the widely distributed glucocorticoid receptor (GR, or the type II receptor) and the more restricted mineralocorticoid receptor (MR, the type I GR). Both exert effects predominantly through binding to DNA, though rapid non-genomic signalling has also been reported for both (reviewed Funder (1997), Buckingham (2006), Dooley *et al.* (2012) and

Samarasinghe *et al.* (2012)). GR is the major GR, whilst MR is only a GR in a restricted range of tissues or under certain pathological conditions (Funder 1997). Although MR has high intrinsic affinity for the endogenous glucocorticoids cortisol and corticosterone, it is normally specific for the mineralocorticoid, aldosterone, by virtue of the activity of an enzyme, 11 β -hydroxysteroid dehydrogenase type 2 (11 β -HSD2) which converts the physiological glucocorticoids cortisol and corticosterone to cortisone and 11-dehydrocorticosterone, respectively, preventing their inappropriate activation of MR (Chapman *et al.* 2013). MR expression is largely restricted to mineralocorticoid target tissues, but it is also expressed in the heart without 11 β -HSD2 co-expression. Thus, in heart, MR binds endogenous cortisol/corticosterone avidly (Funder 2005). Of note, MR is poorly activated by betamethasone and dexamethasone, the potent synthetic glucocorticoids used in antenatal therapy.

GR has approximately tenfold lower affinity for physiological glucocorticoids than MR and does not bind aldosterone. In mice (Cole *et al.* 1995, Tronche *et al.* 1998) and most likely also humans (Charmandari *et al.* 2004), glucocorticoid action mediated via GR is essential for survival beyond birth. It is also important for survival in late gestation in mice; around 50% of GR^{-/-} fetuses die by E17.5 with characteristics of hydrops foetalis (Rog-Zielinska *et al.* 2013), a condition associated with intra-uterine death in humans that frequently results from heart failure (Bellini *et al.* 2009, Fritsch *et al.* 2012). In contrast, mice which lack MR survive birth, but die in the first 2 weeks of life of circulatory failure caused by renal salt-wasting (Berger *et al.* 1996, Bleich *et al.* 1999). Both GR and MR are expressed in the foetal heart (Brown *et al.* 2005) and while MR appears to play a detrimental role in adult cardiac remodelling (Messaoudi *et al.* 2012), how both receptors influence cardiac development and maturation remains poorly understood. GR is certainly present in the human heart from 12 weeks of gestation (Ballard & Ballard 1974) and in mouse heart from as early as embryonic (E) day E10.5, though it is not activated by endogenous glucocorticoids until E15.5 (Rog-Zielinska *et al.* 2013). A critical role for GR signalling in foetal heart maturation has recently been demonstrated (Rog-Zielinska *et al.* 2013). However, the relationship between *in utero* glucocorticoid exposure and heart maturation appears to follow an inverted U-shaped pattern, typical of many glucocorticoid actions (Munck *et al.* 1984), with both low and excessive glucocorticoids having a detrimental impact on cardiovascular development. Whether these effects share a common mechanism

remains unknown. Nonetheless, given the widespread use of antenatal glucocorticoids in preterm infants to promote lung maturation, it is important to understand the potential impact of this treatment on maturation and growth of the heart and vasculature.

Glucocorticoid signalling in the foetus

The timing and level of exposure to glucocorticoids *in utero* are the key determinants of their lifelong organisational effects upon tissues (Fowden *et al.* 1998, Seckl & Holmes 2007). Inadequate glucocorticoid action can have potentially fatal consequences, yet premature activation of glucocorticoid-dependent pathways or excessive glucocorticoid levels during pregnancy can cause lifelong 'programmed' adverse cardiovascular effects (Benediktsson *et al.* 1993, Louey & Thornburg 2005, Seckl & Holmes 2007; and see below). Endogenous physiological foetal glucocorticoid production is tightly regulated, and increases markedly over a narrow time window shortly before birth (Fig. 1). The foetal adrenal gland initiates *de novo* glucocorticoid synthesis during the second half of gestation, at around the 28th week of pregnancy in humans (Mastorakos & Ilias 2003) and

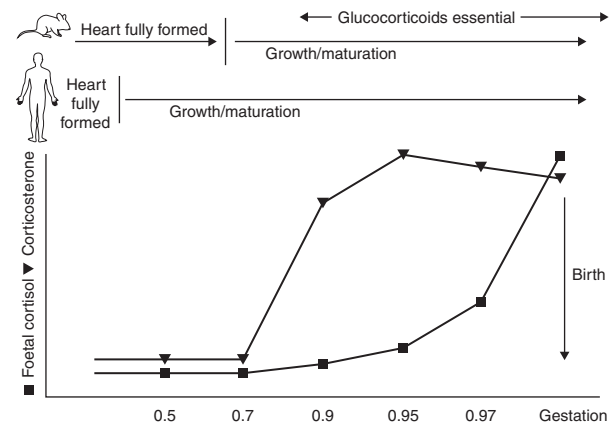


Figure 1

Glucocorticoids rise in the foetus in late gestation, coincident with the growth and maturation of the foetal heart. The foetal heart is fully formed in both mice (~E13.5–E14.5) and humans (~40 days; indicated by vertical bars, top) before the glucocorticoid surge that occurs in the few days before birth in most mammals (Michelson & Anderson 1992, Fowden *et al.* 1998, Henderson & Anderson 2009). In mice, the late gestational increase in foetal plasma corticosterone (triangles in the schematic representation above) is essential for maturation of the foetal heart (Rog-Zielinska *et al.* 2013). In humans, the exact role of the rise in plasma cortisol levels (squares in the representation above) in maturation of the foetal heart remains poorly understood. However, both insufficient or excessive/mis-timed glucocorticoid exposure may have adverse effects on heart function over the long term (see text for details).

E14.5 in mice (Michelsohn & Anderson 1992). In mice, endogenous foetal plasma glucocorticoid levels increase rapidly from E15 (Michelsohn & Anderson 1992) though in humans this physiological increase happens in the week before birth (Fowden *et al.* 1998). In addition, as pregnancy progresses into the final stage, maternal plasma glucocorticoid levels rise dramatically as a result of increased maternal glucocorticoid production and prolonged plasma half-life (Donaldson *et al.* 1991, Mastorakos & Ilias 2003), though placental 11 β -HSD2 restricts the access of maternal glucocorticoids to the foetus (Chapman *et al.* 2013). Placental 11 β -HSD2 activity declines in the last week of gestation in humans (Murphy & Clifton 2003) and in mice (Brown *et al.* 1996, Thompson *et al.* 2002), allowing for transfer of maternal glucocorticoid to the foetus as birth approaches (Cottrell *et al.* 2012). These different effects combine to cause a surge in foetal glucocorticoid levels in late gestation, which act through GR to elicit the critical maturation of foetal lung (Cole *et al.* 1995, Nemati *et al.* 2008), skin (Bayo *et al.* 2008), liver (Opherke *et al.* 2004) and, importantly, heart (Rog-Zielinska *et al.* 2013). In addition to administration of exogenous glucocorticoids, foetal glucocorticoid exposure is potentially affected by a number of mechanisms including prematurity at birth, maternal stress, alterations in placental metabolism and adrenal disease.

Late gestation cardiac maturation

By late gestation, although the heart has attained its adult anatomical configuration, it continues to undergo extensive growth, remodelling and maturation to support the rapidly growing foetus and prepare for life after birth. Both systolic and diastolic function improve in late gestation (Harada *et al.* 1997, Corrigan *et al.* 2010), reflecting the improved contractility and relaxation properties of the cardiomyocytes. The ventricle walls progressively thicken (Webb *et al.* 1996, Christoffels *et al.* 2000), driven predominantly by cardiomyocyte hyperplasia (Oparil *et al.* 1984). As hyperplastic growth gives way to hypertrophic growth following birth, cardiomyocytes become binucleated, less spherical in shape and more rod-shaped allowing for more efficient mechanical and electrical coupling (Burrell *et al.* 2003, Hirschy *et al.* 2006). As these rod-shaped cardiomyocytes align with each other, the muscle fibres in the outermost layer of the compact myocardium form into spiraling bundles to create the typical architecture of the mature mammalian heart. The change in cardiomyocyte shape is underpinned by ultrastructural changes. Contractile proteins accumulate

and switch from foetal to adult isoforms (myosin heavy chain and titin, for example) as myofibrils grow and align parallel to the long axis of the cardiomyocyte, acquiring more sarcomeres and well-defined Z-discs (Ehler *et al.* 1999, Siedner *et al.* 2003). Towards the end of gestation, the sarcoplasmic reticulum also swells and undergoes structural changes (Canale *et al.* 1986) in order to facilitate calcium-induced calcium release (Rapila *et al.* 2008). This is accompanied by marked changes in ion channel expression and activity, as electrical coupling in the heart matures in the time period close to birth (Davies *et al.* 1996). The late gestation heart also undergoes a significant metabolic transformation as it prepares to adjust to the high-oxygen extra-uterine environment and increased haemodynamic work load after birth. The number of mitochondria increase as cardiomyocytes switch from utilising carbohydrates (mainly glucose) as the predominant source of energy to fatty acids, a more efficient source of ATP (Bartelds *et al.* 2000). All of these morphological and functional changes enable the late gestation foetal heart to increase cardiac output as the foetus grows and prepare for the increase in cardiac work load at birth.

Glucocorticoids, through GR, promote foetal heart maturation

Mechanical and hormonal factors, including glucocorticoids, play important roles in cardiac maturation, although understanding the contribution made by endogenous glucocorticoids, particularly in rodents, can be difficult due to compensation between foetal and maternal hypothalamic–pituitary–adrenal (HPA) systems (Montano *et al.* 1993). Moreover, endogenous glucocorticoids, cortisol and corticosterone may act through either GR or MR. Recent work has shown that glucocorticoids act via GR to promote foetal heart maturation through both direct and indirect effects on cardiomyocytes (Rog-Zielinska *et al.* 2013). In the absence of glucocorticoid signalling, the late gestation foetal heart is immature and functions poorly. In GR^{-/-} mice or mice with tissue-specific disruption of GR in cardiomyocytes and vascular smooth muscle, the initial phase of contraction takes longer and the normal macro- and micro-architecture of the compact myocardium is disrupted (Rog-Zielinska *et al.* 2013). This is associated with immature cardiomyocyte ultrastructure featuring short and disorganised myofibrils (Rog-Zielinska *et al.* 2013). The normal maturational change in the expression of genes involved in calcium handling and energy metabolism that occurs between

E14.5 and E17.5 (spanning the time of the glucocorticoid surge) is absent in GR^{-/-} mice (Rog-Zielinska *et al.* 2013). Not all of these gene-expression changes are a direct result of GR in cardiomyocytes and/or vascular smooth muscle cells (Rog-Zielinska *et al.* 2013), suggesting that extra-cardiac glucocorticoid actions mediated by GR are also key to late gestation heart maturation, possibly via haemodynamic changes. GR^{-/-} fetuses exhibit oedema, which must be due, at least in large part, to GR actions in cardiomyocytes and/or vascular smooth muscle, as specific knockout of GR in these cells also causes foetal oedema, albeit to a lesser extent. In contrast to prematurity, when birth occurs before the normal rise in endogenous cortisol levels (see below), the hydrops phenotype of late gestation GR^{-/-} fetuses is manifested following the onset of the glucocorticoid surge (Rog-Zielinska *et al.* 2013), raising the possibility that, in some cases, foetal glucocorticoid insufficiency or even glucocorticoid resistance may be a cause of non-immune hydrops foetalis in humans. Clinically, cardiogenic hydrops typically presents with impaired diastolic ventricular filling (Bellini *et al.* 2009), as seen in GR^{-/-} fetuses (though not in fetuses with cardiomyocyte-specific GR knock-out). Glucocorticoids are an effective treatment in hydrops foetalis associated with congenital cystic adenomatoid malformations (CCAM; Tsao *et al.* 2003, Curran *et al.* 2010), although the therapeutic mechanism remains to be ascertained (Leung *et al.* 2005) and may relate to reduced mediastinal shift secondary to glucocorticoid-induced maturation or involution of the lung lesions, restoring cardiac venous return and improving contractility (Knox *et al.* 2006).

Whether MR plays a role in the glucocorticoid-induced maturation of cardiac structure and function is important and remains to be determined. However, cardiac MR mRNA levels are not affected by GR deficiency *in utero* (Rog-Zielinska *et al.* 2013).

Insufficient glucocorticoid action: prematurity

Preterm birth increases the risk of cardiovascular disease in later life (Crump *et al.* 2011). As preterm birth occurs before the physiological late gestation rise in foetal glucocorticoids and before foetal organs are mature enough for extra-uterine life, it is possible that perinatal glucocorticoid deficiency underlies this increased risk. Endogenous glucocorticoid action occurs under the control of the foetal HPA axis. Although the HPA axis can respond to a variety of stressors in prenatal and neonatal life, the magnitude and kinetics of the responses

are dependent on developmental age (Walker *et al.* 1991). Thus, developmental immaturity in the preterm infant may be associated with a limited adrenal reserve. Relative adrenal insufficiency in preterm infants, with low serum cortisol or an inability to mount an adequate cortisol stress response, has been suggested as a contributory factor to haemodynamic instability and increased mortality in preterm infants (Ng *et al.* 2004, Nykänen *et al.* 2007, Fernandez & Watterberg 2009).

Cardiovascular complications in premature infants include persistence of the ductus arteriosus (the foetal shunt that connects the pulmonary artery to the aorta, bypassing the lungs) and the foramen ovale (that allows blood to flow directly from the right to the left atrium, bypassing the right ventricle and the lungs), a delay in the normal decrease in pulmonary vascular resistance (the foetal lungs have a high resistance) and a delay in the necessary increase in cardiac output required to manage the additional volume load after birth (Huhta 2005). Consequently, haemodynamic instability is a well-recognised problem in premature infants (Ng *et al.* 2004, Kluckow 2005, Sehgal 2011) with hypotension and low systemic blood flow associated with increased mortality (Ibrahim *et al.* 2011). Many of these markers of prematurity are improved by exogenous glucocorticoid treatment. Glucocorticoids facilitate the closure of the ductus arteriosus by inhibition of prostaglandin E₂ efficacy (Clyman *et al.* 1981, Eronen *et al.* 1993). They are also an effective treatment for refractory systemic hypotension in preterm infants, either alone or in combination with other volume expansion/ionotropic agents (Noori *et al.* 2006, Ibrahim *et al.* 2011). In foetal sheep, the normal late gestation increase in arterial blood pressure is abolished by adrenalectomy and restored by cortisol replacement (Unno *et al.* 1999), supporting a crucial role for endogenous cortisol secretion in achieving normal circulatory function.

Systolic function of the left ventricle, measured by cardiac output, fractional shortening, ejection fraction or left ventricular mass and diameter, is typically impaired in the preterm foetus compared with full-term neonates (Harada *et al.* 1999, Kozák-Bárány *et al.* 2001, Ciccone *et al.* 2011). In addition, impaired diastolic function is increasingly recognised as an equally, if not more, important determinant of postnatal outcome. In a series of clinical studies, parameters of diastolic function such as the E-wave velocity and early-to-atrial filling ratios were significantly lower in preterm infants (Kozák-Bárány *et al.* 2001, Schmitz *et al.* 2004, Ciccone *et al.* 2011). Myocardial performance index, a measure of systolic and

diastolic function that is independent of heart rate and size, is higher in preterm infants and strongly correlates with severity of pulmonary dysfunction and poor outcomes (Yates *et al.* 2008). Given the importance of GR signalling for the maturation of cardiac function in mice shortly before birth, it is plausible that insufficient or mistimed glucocorticoid signalling may contribute to these common cardiovascular complications. The short-term effects of glucocorticoids on heart function itself have been little investigated and are difficult to dissect from the more general haemodynamic effects. However, in immature baboons in which adrenal insufficiency was associated with impaired heart function (measured by fractional shortening and velocity of circumferential fibre shortening), a short course of neonatal cortisol led to a sustained improvement in cardiac performance (Yoder *et al.* 2002), consistent with a beneficial effect of glucocorticoid therapy on heart maturation.

Whether adrenal insufficiency and its associated haemodynamic instability results from prematurity *per se*, or as a consequence of antenatal glucocorticoid therapy, is an important question. A meta-analysis of 49 human studies assessed the impact of antenatal treatment with synthetic glucocorticoids and concluded that although basal HPA axis function recovered within 2 weeks of delivery, there was a sustained (>4–8 weeks) suppression of the cortisol response to pain (Tegethoff *et al.* 2009), suggesting that antenatal glucocorticoid treatment itself may cause a persistent inability to mount a cortisol stress response. Thus, both immaturity and antenatal glucocorticoid therapy may result in a similar limited adrenal reserve in premature birth.

Although in humans it is difficult to separate the long-term effects of prematurity from the effects of glucocorticoid treatment and possible over- or mis-timed exposure, animal models are providing valuable information to support a 'programming' effect of early or excessive glucocorticoid exposure on the foetal cardiovascular system (Seckl & Holmes 2007, Cottrell & Seckl 2009).

Precocious or excessive glucocorticoid action

For two decades it has been recognised in animal studies that exposure to excessive glucocorticoid levels *in utero*, by bypassing or overwhelming placental 11 β -HSD2, causes short-term effects such as foetal growth restriction and long-term effects that include increased risk of cardiovascular disease when the offspring reach adulthood (Benediktsson *et al.* 1993, Nyirenda *et al.* 1998). Maternal nutritional restriction, which prematurely activates the

foetal HPA axis (Cottrell *et al.* 2012), also increases foetal glucocorticoid levels and 'programmes' greater risk of cardiovascular disease once adult (Langley-Evans *et al.* 1998, Roseboom *et al.* 2001), as does maternal stress, probably through increased maternal glucocorticoid levels (Seckl & Holmes 2007, Bingham *et al.* 2013). Glucocorticoid and/or nutritional programming of adult disease has been extensively reviewed (for recent reviews see Seckl & Holmes (2007), Langley-Evans (2013) and Reynolds (2013)). In addition, the reader is referred to a comprehensive recent review of the acute effects of betamethasone and dexamethasone on human foetal behaviour (movement/breathing) and haemodynamics (heart rate, Doppler velocity measurements of blood flow in the umbilical artery) (Mulder *et al.* 2009). In this study, we restrict our discussion to the impact of elevated glucocorticoid signalling on the maturation of the prenatal heart and its possible long-term consequences.

In vivo studies have largely focused on the ability of exogenous glucocorticoids to increase foetal heart size, which occurs despite a general restriction of foetal growth in humans and in animal models. However, whether this involves glucocorticoid-induced alterations in cardiomyocyte proliferation, binucleation and/or hypertrophy remains controversial. Moreover, foetal blood pressure, a well-known stimulus of myocardial maturation, is increased by exogenous glucocorticoids, further confounding interpretation. In some studies in rats and sheep, the glucocorticoid-induced increase in heart weight and protein:DNA ratio has been interpreted as late gestation cardiomyocyte hypertrophy, without proliferation (Slotkin *et al.* 1991, Rudolph *et al.* 1999, Jensen *et al.* 2002). Other studies report a higher proliferative index in hearts following antenatal glucocorticoid administration, suggesting hyperplasia rather than hypertrophy as the mechanism of glucocorticoid-induced heart enlargement (Torres *et al.* 1997, Giraud *et al.* 2006). Many studies do not report foetal haemodynamic effects of glucocorticoid administration. However, when cortisol was infused into the coronary artery of foetal sheep, hyperplasia was induced with only a modest increase in intracardiac cortisol (threefold) and without blood pressure changes (Torres *et al.* 1997, Giraud *et al.* 2006), suggesting a pro-proliferative effect of cortisol on cardiomyocytes. A recent study on mice has administered dexamethasone from E12.5 to E15 (before the normal glucocorticoid surge; O'Sullivan *et al.* 2013). Although this caused a transient decrease in foetal weight and a corresponding though non-significant decrease in foetal heart weight at E14.5, this had recovered by E17.5,

possibly as a result of increased cardiac expression of insulin-like growth factor 1, and cardiomyocyte number in adulthood was unchanged (O'Sullivan *et al.* 2013), suggesting compensatory myocardial growth once dexamethasone treatment stopped (though glucocorticoid-regulated mRNAs remained elevated at E17.5, indicating a lasting effect of glucocorticoid action on foetal cardiomyocytes). However, the possibility of foetal HPA axis suppression by dexamethasone treatment was not examined in that study, so it is unclear whether the normal increase in glucocorticoid levels at E16.5–E18.5 occurred in the treated mice and it is likely that the late gestation trajectory of heart maturation was altered in response to dexamethasone. Differences in study outcomes are likely to result from different steroid regimes: dose, steroid used – cortisol (which binds to both GR and MR), betamethasone or dexamethasone (GR-selective) and/or timing. For example, experimental models using rats commonly use dexamethasone in late gestation, after the onset of the endogenous glucocorticoid surge, whereas sheep models have typically treated either much earlier (around 0.6 gestation) or in late gestation (shortly before the normal increase in foetal glucocorticoid levels) with dexamethasone, betamethasone or cortisol. Recent data implicate MR in cortisol-induced cardiomyocyte hyperplasia, with GR exerting anti-apoptotic effects in late gestation in sheep (Feng *et al.* 2013). Moreover, in mice with cardiomyocyte/vascular smooth muscle knockout of GR (and normal plasma glucocorticoid levels), heart size was similar to littermate controls at E17.5 (Rog-Zielinska *et al.* 2013), suggesting that the smaller hearts of GR^{-/-} fetuses are an indirect result of glucocorticoid signalling and not a direct effect of GR in cardiomyocytes. Thus, increased heart size may result from stimulation of myocyte proliferation mediated by cortisol signalling through MR, or may reflect indirect effects, most likely haemodynamic, mediated by GR elsewhere. This has implications for antenatal therapy if the synthetic glucocorticoids used cause hypertrophy but do not cause the normal increase in myocyte proliferation that normally occurs near term. It will be of interest to test whether mice with cardiomyocyte-specific knock-out of MR (Fraccarollo *et al.* 2011) have smaller hearts in late-gestation.

The physiological rise in endogenous foetal glucocorticoids is important for the maturation of electrical coupling in the foetal heart, and abnormalities in sodium channel expression linked to dysregulated plasma cortisol levels have been suggested as a cause of sudden infant death syndrome in premature infants

(Fahmi *et al.* 2004). In immature foetal sheep, cortisol infusion (before the normal rise) precociously increased expression/transcripts of the cardiac sodium channels, SCN1b and SCN5a, altering the normal developmental trajectory of expression (Fahmi *et al.* 2004), suggesting potential consequences for cardiac function. Similarly, prenatal dexamethasone treatment in rats can cause lasting decreases in noradrenaline levels and turnover that persist into adulthood (Bian *et al.* 1993), suggesting that abnormal glucocorticoid exposure in early life could have a long lasting impact on cardiac sympathetic responses.

In late gestation, glucocorticoids accelerate the maturation of energy metabolism in the foetal heart. In rats, foetal dexamethasone exposure (at a time when endogenous corticosterone levels have peaked) increased ATP levels and delivery of ATP to myofibrils (Tsuzuki *et al.* 2009, Mizuno *et al.* 2010). It also increased levels of pyruvate and lactate in foetal heart and blood respectively (Jensen *et al.* 2002, Tsuzuki *et al.* 2009), suggesting increased glycolysis. Increased capacity to generate ATP in the late gestation foetal heart is likely to be critical to cope with the increased energy demand at birth. On the other hand, precocious maturation of oxidative energy production in a very immature foetal heart could be detrimental if energy substrates or oxygen levels subsequently become limiting and could increase reactive oxygen species.

Long-term consequences of prematurity and/or antenatal glucocorticoid therapy: alterations in the normal trajectory of heart maturation?

Animal studies have clearly shown that prenatal glucocorticoid administration increases blood pressure in adult offspring (reviewed Barker & Fall (1993), Benediktsson *et al.* (1993) and Dodic *et al.* (2003)). However, these models usually involve early and/or extended intervention in otherwise healthy foetuses. In humans it is difficult to disentangle the effects of glucocorticoid insufficiency associated with immaturity from the effects of antenatal glucocorticoid therapy, as these situations normally go hand-in-hand. Moreover, glucocorticoid administration preceding premature delivery has only become routine in the last 30–40 years, so the longer term effects on cardiovascular outcomes may yet unfold. Nonetheless, most studies to date do not report any deleterious effects of antenatal glucocorticoid therapy on blood pressure, heart rate or overall cardiac performance (systolic or diastolic)

in neonates, children, adolescents or in adults at 30 years of age (Dalziel *et al.* 2004, 2005, de Vries *et al.* 2008). A few follow-up studies report that children exposed to antenatal corticosteroids have moderately heightened systolic and diastolic blood pressures at the age of 7 (Wilson *et al.* 2006) and 14 (Doyle *et al.* 2000), although none were in the hypertensive range. However, prematurity and/or antenatal glucocorticoid therapy may alter the normal trajectory of postnatal heart development and function (Bensley *et al.* 2010). Interestingly, the hearts of young adults that were born premature have been reported to have a unique three-dimensional left ventricular shape with a displaced apex and shorter ventricular length and increased left ventricular mass (Lewandowski *et al.* 2013). This is associated with reduced systolic torsion/rotation of the ventricle. Whether this functional change is prenatally determined (including via antenatal glucocorticoid treatment) or due to a premature adaptation to postnatal life is currently unclear. Nevertheless, the lack of alignment of the outermost circumferential muscle fibres in GR^{-/-} foetuses (Rog-Zielinska *et al.* 2013) suggests that maturation of systolic twisting could be a glucocorticoid-regulated process. These aspects of prematurity and/or glucocorticoid action will be important to further investigate in animal models. Whether these structural and functional changes result in abnormal responses to cardiac insults or haemodynamic load in later life is clearly of great interest and remains unproven at this stage. Other aspects of cardiac development, including the electrical conducting system of the heart, are known to be affected by prematurity. For example, adults born premature show a prolonged QT interval and increased QT dispersion, making them more vulnerable to ventricular arrhythmias (Bassareo *et al.* 2011), supporting the notion of increased cardiovascular risk.

A further difficulty encountered clinically is distinguishing the effects of small foetal size due to immaturity from other causes of intra-uterine growth restriction (IUGR). Haemodynamic adaptations to glucocorticoids may differ in IUGR foetuses (Hodges & Wallace 2012) and indeed, foetal growth restriction can be associated with glucocorticoid excess, for example as a result of 11 β -HSD2 deficiency (Holmes *et al.* 2006, Seckl & Holmes 2007). Because IUGR foetuses are at increased risk of premature birth, they are also likely to receive antenatal glucocorticoid therapy. The wisdom of this has recently been questioned (Hodges & Wallace 2012). IUGR is associated with altered heart shape (more globular-shaped ventricles) and function (reduced stroke volume and subclinical systolic dysfunction) in children, the majority

of which also receive antenatal glucocorticoid treatment (Crispi *et al.* 2010). Whilst there is no evidence that the risks of antenatal glucocorticoid therapy outweigh the benefits in this group, more follow-up studies focussed on the cardiac consequences are clearly warranted.

Implications and concluding remarks

Medical advances have led to more widespread use of antenatal glucocorticoid therapy, with intervention earlier in gestation, and greater use of repeat courses of glucocorticoids. Whilst it is clear that this therapy has reduced perinatal mortality and morbidity, clinicians have focused on early neonatal outcomes. Long-term effects of the high doses in routine use, particularly on the developing heart, remain unclear. Data from animal models, including non-human primates, suggest dose-dependent programming effects of antenatal dexamethasone on cardiovascular risk factors (Bian *et al.* 1990, de Vries *et al.* 2007). The large-scale randomised clinical trials that first demonstrated the benefits of antenatal glucocorticoid therapy included few below 30 weeks gestation, yet the same therapy is now applied from 24 weeks, often with repeat doses.

Most clinical trials suggest that sustained exposure or repeated doses of corticosteroids are more effective in improving foetal outcomes (Abbasi *et al.* 2000, Crowther *et al.* 2006, Murphy *et al.* 2008, Asztalos *et al.* 2010). However, there is also a considerable body of evidence to suggest that multiple dosing does not carry any additional respiratory or cardiovascular benefits for the infant and is more detrimental for foetal growth and postnatal neurodevelopment, as well as adrenal function, compared with a single dose (Banks *et al.* 1999, Spinillo *et al.* 2004, Nair & Omar 2009, Bontis *et al.* 2011, Murphy *et al.* 2012).

Clinically, betamethasone or dexamethasone is used, yet actions of the endogenous hormone, cortisol via MR, may be important for some of the normal maturational effects of endogenous glucocorticoids (such considerations do not apply to lung, where MR is co-expressed with 11 β -HSD2). Individual sensitivity to glucocorticoids may influence responses to therapy; the exon 9 β polymorphism in the NR3C1 gene encoding GR is associated with relative glucocorticoid resistance and also with higher blood pressure and left ventricular mass in children (Geelhoed *et al.* 2011). Thus, there is an important need to understand the normal actions of glucocorticoids on the foetal heart to address how timing and dose of glucocorticoid treatment might impact upon its developmental trajectory. Increasingly, therapy can be tailored to the

individual and in future detailed foetal echocardiography should allow better diagnosis of possible cardiac dysfunction and distinction between the growth-restricted foetus and the immature foetus. This tailored therapy approach might ensure that the most appropriate steroid regime is used to minimise long-term adverse effects, particularly on the heart.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review reported.

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