REVIEW

Thyroid function disruptors: from nature to chemicals

Karen Jesus Oliveira¹, Maria Isabel Chiamolera², Gisele Giannocco²,³, Carmen Cabanelas Pazos-Moura⁴ and Tania Maria Ortiga-Carvalho⁵

¹Departamento de Fisiologia e Farmacologia, Laboratório de Fisiologia Endócrina e Metabologia, Universidade Federal Fluminense, Rio de Janeiro, Brazil
²Disciplina de Endocrinologia e Metabologia, Laboratório de Endocrinologia Molecular e Translacional, Escola Paulista de Medicina, Universidade Federal de São Paulo, São Paulo, Brazil
³Departamento de Ciências Biológicas, Universidade Federal de São Paulo, Diadema, Brazil
⁴Laboratorio de Endocrinologia Molecular, Instituto de Biofísica Carlos Chagas Filho, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil
⁵Laboratório de Endocrinologia Translacional, Instituto de Biofísica Carlos Chagas Filho, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Rio de Janeiro, Brazil

Correspondence should be addressed to T M Ortiga-Carvalho: taniaort@biof.ufrj.br

Abstract

The modern concept of thyroid disruptors includes synthetic chemicals and bioactive compounds from food that interfere with any aspect of the hypothalamus–pituitary–thyroid axis, thyroid hormone biosynthesis and secretion, blood and transmembrane transport, metabolism and local actions of thyroid hormones. This review highlights relevant disruptors that affect populations through their diet: directly from food itself (fish oil and polyunsaturated fatty acids, pepper, coffee, cinnamon and resveratrol/grapes), through vegetable cultivation (pesticides) and from containers for food storage and cooking (bisphenol A, phthalates and polybrominated diphenyl ethers). Due to the vital role of thyroid hormones during every stage of life, we review effects from the gestational period to adulthood, including evidence from in vitro studies, rodent models, human trials and epidemiological studies.

Key Words

- bioactive food compounds
- chemical disruptors
- TSH
- thyroid hormones

Introduction: temporal description, definitions, basic concepts

Although the terms endocrine disruption and endocrine disruptors were coined at the Wingspread Conference in 1991, the concept of endocrine alterations caused by synthetic chemicals has followed the larger context of the environmental movement that has had various influences on science since the 1960s and 1970s (Schug et al. 2016). Several factors have guided this journey, such as Rachel Carson’s 1962 book Silent Spring; several wildlife studies showing reproductive alterations in birds, alligators and fish (Gilbertson & Reynolds 1972, Aulerich et al. 1973, Aulerich & Ringer 1977, Semenza et al. 1997, Jobling et al. 2002); and the disastrous use of an estrogen analog, diethylstilbestrol, which was used by millions of women during pregnancy to reduce the risk of miscarriage: later this use was linked to the occurrence of unusual cancers and reproductive system malformations for those exposed in utero (McLachlan et al. 1980, Newbold et al. 1985). Dietary phytoestrogens, which are phytochemicals that are structurally similar to estrogens, are able to bind and activate estrogen receptors (ERs), mimicking or interfering with endogenous estrogen pathways. Therefore, these plant-derived compounds can induce many health
benefits but can also cause harmful effects, such as an elevated breast cancer risk and premature puberty (Sauve & Desrosiers 2014).

In 2012, the Endocrine Society published (Zoeller et al. 2012, Schug et al. 2016) a statement of principles and used the following definition to describe endocrine-disrupting chemicals (EDCs): chemicals, or mixtures of chemicals, which interfere with any aspect of hormone action (Patau & Jeffery 2010, Bode & Dong 2015). There are a great variety of agents that have been reported to have endocrine-related effects, including more than 1000 chemicals from several chemical classes (Diamanti-Kandarakis et al. 2009). These chemicals are not identified by their chemical structure or by a specific type of usage but rather by their mechanisms of action and ability to alter endocrine system function. In this context, there are a large number of chemicals with different structures present in food, and many of them have biological properties, including endocrine-disrupting-related effects (Gore et al. 2015), which extends the concept of endocrine disrupters beyond man-made chemicals. Several reviews have explored the effects of chemicals and bioactive compounds in food on the reproductive system. In addition to effects on the reproductive system, endocrine disruptors also affect different endocrine glands, including the thyroid, a fundamental gland for neural development. Thyroid hormones (THs) are released under the regulation of the hypothalamus–pituitary–thyroid (HPT, Fig. 1) axis and other factors that control the mechanism of action of THs (Fig. 2), which have been extensively reviewed elsewhere (Ortiga-Carvalho et al. 2014, 2016, Bernal et al. 2015). Importantly, endocrine disrupters may act at any level of this regulation. In this review, we explore the effects of natural and artificial compounds on thyroid function and regulation.

**Bioactive food compounds as thyroid-disrupting factors**

Some vegetables, such as cabbage, broccoli and cassava, are a source of antithyroid compounds, such as thiocyanate and thiocyanates, which are well-characterized goitrogens (Gaitan 1990, Felker et al. 2016, Willemin & Lumen 2017). However, the consumption of other foods and food-related compounds, such as soy protein and soybean isoflavones, as well as other flavonoids, may interfere with thyroid function. These effects of those compounds have been reviewed recently by several authors (Doerge & Chang 2002, Messina & Redmond 2006, Xiao 2008, de Souza Dos Santos et al. 2011). Here, we focus on reviewing the thyroid-disrupting properties of other less well-known bioactive food compounds.

**Fish oil and polyunsaturated fatty acids**

Essential fatty acids play important roles in plasma membrane integrity and function, energy production and as precursors of bioactive lipids at all stages of life. The long chain $n–3$ polyunsaturated fatty acid ($n–3$ PUFA) docosahexaenoic acid (22:6n-3; DHA) is especially important to brain development during gestation and infancy (Chilton et al. 2017). Male rat pups supplemented with DHA during the first 6 weeks of life exhibited higher serum thyroid-stimulating hormone (TSH) levels without changes in other pituitary hormones (Clandinin et al. 1998), which suggests that DHA interferes in the HPT axis (Fig. 1). However, other authors (Souza et al. 2010) studying rats supplemented with fish oil, which is enriched in $n–3$ PUFAs, from lactation until 11 weeks of age found no alterations in serum TSH or TH concentrations. The authors detected higher levels of the TH receptor β (THRβ) protein in the liver, accompanied by increased mitochondrial glycerophosphate dehydrogenase (mGPD) activity; mGPD is a THRβ-mediated triiodothyronine (T3) target, suggesting that fish oil increases TH sensitivity in this tissue. However, deiodinase type 1 (D1, DIO1) activity, another recognized marker of TH action in the liver (Fig. 2), was not changed, suggesting that the $n–3$ PUFAs present in fish oil might interfere with T3 action in a target-specific manner (Souza et al. 2011). Another study showed that hepatocytes treated with T3 and eicosapentaenoic acid (20:5n-3; EPA) showed a reduction of 70% in the effect of T3 on thyroid hormone responsive spot 14 (Thsrp) gene transcription (Jump et al. 1993). However, the authors showed that the Thsrp TH-response element (TRE) was not directly sensitive to EPA inhibition. The PUFAs-responsive element was found to be located in a region that potentiates T3-mediated activation of Thsrp gene transcription.

In hypothyroid patients, higher levels of free fatty acids were associated with lower symptom severity, along with lower serum TSH and higher thyroxine (T4) and T3 levels, than in patients with low levels of plasma free fatty acids (Makino et al. 2001). These authors also showed, in methimazole-induced hypothyroid rats, that chronic EPA supplementation for 28 days reduced the methimazole-induced drop in TH levels and the consequent increase in serum TSH levels (Makino et al. 2001). This study suggests that EPA exerts a direct stimulatory effect on the thyroid.
Pepper and piperine

Piperine is the main alkaloid found in the fruit *Piper nigrum*, which is also known as black pepper. Studies have shown that piperine enhances the bioavailability of several drugs and nutraceuticals and possesses important pharmacological effects, including anticancer, anti-inflammatory and antimicrobial activities (Chavarria et al. 2016). Although there is no evidence for the effects of piperine in humans, studies conducted in rodents point to a thyroid-disrupting effect of pepper and isolated piperine. Two different preparations of the *Piper* fruit were tested in mice. A water extract induced a reduction in serum T3 and T4 levels after 15 days of treatment. An ethanolic extract had the opposite effect and induced an elevation in TH levels (Panda & Kar 2003b). The study did not evaluate the chemical composition of the extracts to identify the bioactive component responsible for the different effects observed. Another study investigated the effect of isolated piperine administered orally for 15 days to mice. This study found suppressed hepatic D1 activity in the groups receiving high or low doses of piperine, with
the former showing reduced serum T3 and T4 levels and the latter only showing reduced serum T3 levels (Panda & Kar 2003a). These authors suggested that piperine has a direct suppressive effect on the thyroid at higher doses, but at lower doses, it directly inhibits the peripheral metabolism of THs mediated by the D1 in the liver.

Coffee and caffeine

Coffee contains over a thousand components, including many with biological activity, such as caffeine, diterpene alcohols and chlorogenic acid, which are potential nutraceuticals. Although the biological effects of coffee are not restricted to caffeine, this is the most studied component of coffee (O’Keefe et al. 2013). The effects of caffeine on thyroid function in animal models are inconclusive due to variations in the species, ages and doses studied. Caffeine has been used for more than 40 years as a prescribed drug in neonatal medicine, mostly for apnea treatment, but the overall effect of caffeine exposure at this age is poorly understood (Kreutzer & Bassler 2014). In newborn rats, a single administration of caffeine induced a biphasic response, with plasma T4 levels elevated after 4 h and then reduced 24 h after administration (Clozel et al. 1983). These authors suggested that the decrease in T4 concentration was a direct effect of caffeine on the thyroid gland since no significant change in thyrotropin-releasing hormone (TRH)-stimulated TSH levels were observed at the same point. However, after 10 days of caffeine administration, basal serum TSH and T4 concentrations were increased, but after TRH stimulation, TSH levels were attenuated by caffeine, suggesting that chronic exposure to caffeine may lead to exhaustion of the pituitary reserve (Clozel et al. 1983). A study performed in preterm infants showed that caffeine was negatively associated with TSH levels at the 7th postnatal day (PND) but was positively associated with TSH levels at the 14th day, and caffeine showed no correlation at the 28th day. The reverse T3 (rT3) level was negatively correlated with caffeine exposure at the 7th PND. Therefore, these authors suggested a mild and transitory effect of caffeine on thyroid function in newborn humans (Williams et al. 2005). In adult rats, an acute injection of caffeine induced a decrease in TSH levels after 1–6 h, followed by a reduction in T3 and T4 levels after 4 h. An injection of anti-somatostatin antiserum blocked the inhibitory effect of caffeine on TSH secretion, and the incubation of an isolated pituitary gland with caffeine did not elicit changes in TSH release, suggesting that the effect of caffeine on TSH release appears to be mediated by hypothalamic-derived somatostatin (Spindel et al. 1984). The chronic exposure to caffeine associated with regular intake induces tolerance related to hemodynamic and humoral effects (O’Keefe et al. 2013), and this seems to extend to the effects on thyroid function as well.
Spindel et al. (1984) observed that an acute injection of caffeine induced an inhibitory effect on serum TSH; however, a 7-day treatment with caffeine induced tolerance to its effect on serum TSH. A possible tolerance-inducing effect on thyroid function was also observed in Syrian golden hamsters treated for 90 days with caffeine. After 3 days of treatment, the authors observed a transient increase in serum T3 levels, without changes in T4 levels. After 90 days of treatment, no changes in serum T3 and T4 levels or in thyroid histopathology were identified (Bartsch et al. 1996). Evidence in humans is complex but also points to a transitory effect and a possible tolerance-related outcome. The acute intake of caffeine did not promote any changes in serum TSH or T3 levels over 4h in healthy adults who normally drank one to three cups of coffee a day (Spindel et al. 1984). In a study conducted by Friedrich et al. (2017), a strong positive correlation between the levels of free T4 (FT4) and urinary trigonelline, a urinary marker of coffee consumption in humans (Lang et al. 2011), was observed in cross-sectional analyses. However, this positive association was lost in longitudinal analyses, and higher trigonelline levels in urine samples were related to a greater decline in FT4.

Grape and resveratrol

Resveratrol is a natural polyphenol stilbene found in foods such as grapes, berries and peanuts, as well as in red wine, which is the main source of resveratrol in the Mediterranean diet (Gambini et al. 2015). In vitro resveratrol incubation of the rat thyroid cell line FRTL-5 induced a rapid and transient stimulatory effect of iodide trapping, with the highest effect occurring between 6h and 12h, and after 24h, no changes were observed. The stimulatory effect was dose dependent and associated with increased levels of the solute carrier family 5A (also known as sodium-iodide symporter or NIS) protein. In addition, the effect of resveratrol on iodide uptake after 6h of incubation was additive to the effect TSH stimulation and was mediated by a CAMP-independent pathway (Sebai et al. 2010). Interestingly, after 48h of incubation, the effect of resveratrol on FRTL-5 cells was the opposite: it induced a reduction in iodide uptake accompanied by the reduced expression of NIS and the gene that encodes this transporter, Slc5a5 (Giuliani et al. 2014). At this time point, resveratrol also downregulated the expression of TSH receptor (Tshr), thyroid peroxidase (Tpo), thyroglobulin (Agic et al. 2007) and the related transcriptional factors NK2 homeobox 1 (Nkx2-1), forkhead box E1 (Foxe1) and paired box 8 (Pax8) mRNAs in FRTL-5 cells (Giuliani et al. 2017). In vivo, resveratrol administered to adult male rats for 14 days induced lower NIS protein expression levels and reduced radioiodine uptake by the thyroid, with no significant effects on thyroid growth or TSH and TH concentrations (Giuliani et al. 2014). In a chronic study, the treatment of adult male rats with resveratrol for 60 days promoted increased thyroid mass and higher serum TSH levels, without altering the serum concentration of THs, and the histological analyses showed an increased number of follicles, with lower thyroglobulin concentrations in the colloid (Giuliani et al. 2017). Therefore, these data suggest that resveratrol elicits a direct antithyroid effect and a mild goitrogenic effect when administered chronically. The thyroid-related effects of resveratrol can be different in females, as showed previously (Bottner et al. 2006). Ovariectomized female rats treated chronically with resveratrol for 3 months showed higher levels of serum T3, without changes in serum T4 or TSH levels, pituitary TSH beta subunit (Tshb) mRNA expression, thyroid weight or the gross morphology of the thyroid. In addition, resveratrol had no effect on TRH-stimulated TSH secretion from isolated pituitary cells (Bottner et al. 2006). These authors pointed out that the higher T3 levels observed in their study cannot be related to the estrogenic action of resveratrol since 17β-estradiol treatment of ovariectomized rats did not promote any change in the concentration of T3. In addition, the authors suggested that the higher T3 levels should be related to increased D1 activity or altered levels of T3-binding proteins such as thyroxine-binding protein (TBG) (Bottner et al. 2006). Resveratrol administration for 26 weeks to female rats fed a high-fat diet (HFD) attenuated the increase in total serum T4 levels and the reduction in total serum T3 levels induced by a HFD, while this treatment increased the levels of free T4 and T3 in HFD-fed mice. Resveratrol also increased levels of the Dio1 mRNA in the hearts of HFD-fed mice, restoring expression levels to those observed in rats fed a normal lipid diet, and resveratrol decreased Trα1 (Thrα) mRNA expression in the hearts of HFD-fed rats. These results indicate that resveratrol can affect the peripheral metabolism and actions of THs (Cheserek et al. 2016).

Human studies concerning the effects of grapes or resveratrol on thyroid function are scarce, and they do not point to a thyroid-disrupting effect; these studies have only explored a few parameters related to the thyroid physiology (Tome-Carneiro et al. 2012, 2013).
Cinnamon

Cinnamon is a common spice that is used worldwide and has been widely studied as a nutraceutical for the management of insulin resistance, obesity and dyslipidemia (Rafehi et al. 2012, Medagama 2015). According to the results of human, animal or in vitro studies, cinnamon and its active biological compounds interfere with the biosynthesis or action of many hormones, such as insulin, glucagon-like peptide-1 (GLP1), ghrelin and leptin (Hlebowicz et al. 2009, Rafehi et al. 2012, Camacho et al. 2015, Lopes et al. 2015, Medagama 2015, Bento-Bernardes et al. 2017). In this context, a study showed that rats supplemented with a water extract of cinnamon exhibited lower serum T3 levels accompanied by unaltered serum T4 and TSH levels. Cinnamon did not seem to impact pituitary or liver TH signaling or metabolism since Thrb or Dio expression in these tissues was similar to that in controls. However, cinnamon-treated rats showed a strong reduction of Thra and THRε expression in the cardiac ventricle, with important consequences for the expression of calcium handling proteins (Gaique et al. 2016).

Chemicals as thyroid-disrupting factors

Due to the need to increase the productivity of agriculture and industry, thousands of different chemicals have been developed. Many of these artificial chemicals (such as hexachlorobenzene (HCB) and dichlorodiphenyltrichloroethane (DDT)) have been used as pesticides to improve agricultural efficiency. Plastic components and additives, such as bisphenol A (BPA), phthalates and benzo(a)pyrene diol epoxide (BPDE), are frequently used in products that people use daily to improve industrial processes. The present manuscript will concentrate on describing published data for pesticides that are directly involved in food contamination and chemicals that contaminate food due to their presence in plastic containers (BPA, phthalates and BPDEs) to present a more comprehensive review of selected chemicals. However, several other substances have already been shown to disrupt thyroid function, such as perchlorate, thiocyanate, nitrate, polychlorinated biphenyls, triclosan, dioxins and furans, styrenes, sunscreens and lead. Some of these substances can also reach humans and other animals through food contamination (Pearce & Braverman 2009).

Pesticides are chemical agents that are known to act as endocrine disrupters. For example, in 2014, the European Food Safety Authority reported that of 287 pesticides tested, more than a third of them (103) had thyroid-disrupting properties (Sugiyama et al. 2005, Pearce & Braverman 2009, Bellanger et al. 2015, Leung et al. 2016).

Plasticizers

Plastic items are produced and used intensely worldwide, generating several environmental concerns associated with not only individual plastic use but also related to contamination due to leakage of plastic components into the environment. Two important substances used in the plastics industry that are correlated with endocrine disruption, particularly HPT axis disruption, are BPA and phthalates (Talsness et al. 2009). Humans can ingest these components because they can leach from food cans, microwave containers and polycarbonate bottles.

BPA was shown to interfere with the binding of T3 to its receptor by acting as an antagonist, both in vitro (Moriyama et al. 2002, Gayathri et al. 2004) and in vivo (Zoeller et al. 2005), particularly to the beta isoisform of THR (THRβ), mainly disrupting the negative feedback of THs in the pituitary (Zoeller et al. 2005). It was also shown, in vitro, that BPA suppressed expression of the gene encoding RXR gamma, which forms heterodimers with THR (Iwamuro et al. 2006). Similarly, several phthalate metabolites that act as THR antagonists were detected in Chinese rivers (Ibhaeziehbo & Koibuchi 2011, Shi et al. 2011, Hu et al. 2013, Li et al. 2014), which was corroborated by a transient transfection study with low phthalate doses (Hansen et al. 2016).

The capacity of BPA to interfere with THR transcription activity has been analyzed by several studies, with some of them showing interference at the nuclear transcriptional level, particularly for T3-related genes (Xu et al. 2007, Heimeier et al. 2009, Hansen et al. 2016, Jiang et al. 2016), and one study demonstrated interference at a nongenomic level (Sheng et al. 2012). BPA and several BPA analogs that have recently been introduced in the plastics industry were tested on different TH-responsive cell lines (GH3, FRTL-5 and rat cerebellar cells), and the results showed that they interfered with the expression of genes involved in TH synthesis (Sk5, Tg and Tpo), thyroid cell activity/proliferation (Tshr, Tshb, Thrh, Thra, Dio1 and Dio2) and thyroid transcriptional regulation (Pax8, Nkx2-1 and FoxE1) (Gentilcore et al. 2013, Lee et al. 2017). From these results, one can presume that BPA and BPA analogs may affect thyroid function even at low doses, with a possible mechanism being through NF-kB and RAR/RXR pathways (Ghisari & Bonefeld-Jorgensen 2005, Okada et al. 2007, Somogyi et al. 2016). Phthalate metabolites could affect...
iodine uptake through the regulation of NIS and of Slc5a5 mRNA expression (Breous et al. 2005, Wenzel et al. 2005) and by modulating hormone sulfation (Turan et al. 2005).

Several studies using animal models were controversial in demonstrating disruption of thyroid function after BPA exposure using different doses and windows of exposure. Zoeller et al. showed that there was an increase in serum concentrations of total T4 in pups after perinatal exposure, as early as PND 15, with no difference in TSH levels compared to levels in animals not exposed to BPA (Zoeller et al. 2005). However, another group did not detect any difference in the F1 generation of rats exposed to BPA prenatally (Kobayashi et al. 2005). Pregnant female rats exposed to BPA experienced transient hypothyroidism, while their male pups underwent transient hyperthyroidism, followed by hypothyroidism (Xu et al. 2007). Infant rats exposed during the neonatal period did not present any alterations, but the adult female animals developed a TH status similar to hypothyroidism, and evidence in a primary pituitary cell line showed that BPA alters TSH release through direct action on the pituitary (Fernandez et al. 2018). Studies with pregnant ewes, which have a gestational period more comparable to that of humans, that were exposed to BPA both subcutaneously and by dietary exposure demonstrated lower levels of THs in pregnant females and newborns, and those females showed evidence of a modified deiodination balance (Vigie et al. 2013, Guignard et al. 2017). In analyzing other windows of exposure, mice exposed to BPA during puberty have been shown to have lower free T4 levels (Jiang et al. 2016).

Animal studies of phthalates exposure have shown similar discrepancies; earlier studies suggested that phthalate metabolites could show thyromimetic effect (Price et al. 1988, Badr 1992, Gayathri et al. 2004). However, later studies were more consistent in showing a thyroid antagonistic effect, causing reductions in TH levels and signs of multigenerational and persistent effects (Pereira et al. 2007, Erkekoglu et al. 2012, Liu et al. 2015, Dong et al. 2017, Mahaboob Basha & Radha 2017, Sun et al. 2018). Evidence shows disruption of the TSH/TSHR pathway, with involvement of the hypothalamus (Dong et al. 2017, Sun et al. 2018), and of the TH synthesis machinery (downregulation of NIS, TPO, D1s and transthyretin), as well as increased TH metabolism caused by increased expression of hepatic enzymes (Liu et al. 2015). Phthalates, particularly di-n-butyl phthalate (DBP), might also exacerbate chronic lymphocytic thyroiditis by inducing oxidative stress and changes in serum TBG levels and thyroid interleukin-7 (IL-7) levels (Johns et al. 2016).

In humans, the available data are even more controversial; two prospective studies, CHAMACOS (Chevrier et al. 2013) and HOME, have evaluated the possible associations of BPA with THs during pregnancy and in neonates, and the results are somewhat similar regarding neonatal thyroid profiles, with a negative correlation between BPA exposure and TSH levels only in male newborns in the CHAMACOS study and in female newborns in the HOME study, with the strongest association found when exposure occurred later in pregnancy (Chevrier et al. 2013, Romano et al. 2015). Only the CHAMACOS study observed an inverse association between BPA exposure and total T4 levels in pregnant women (Chevrier et al. 2013).

Regarding possible interference from phthalates during pregnancy, the majority of studies have reported a negative correlation between levels of several metabolites and THs, particularly free and total T4 and free T3 levels, and a positive correlation with TSH levels, and these effects may depend on the timing of exposure during gestation (Huang et al. 2007, 2016, Johns et al. 2015, Yao et al. 2016, Gao et al. 2017). However, other studies have shown very different results, with an inverse association with TSH and a positive association with free and total T4 levels (Huang et al. 2007, 2016, Johns et al. 2015, 2016, Yao et al. 2016, Gao et al. 2017). Regarding phthalate exposure and neonatal thyroid status, some studies have shown a correlation between phthalate levels and TH levels in newborns; a Dutch prospective cohort detected changes in free T4 levels in girls but not in boys (de Cock et al. 2014), and a study in Taiwan found an association of phthalate exposure with reduced levels of THs in young children (Huang et al. 2017a,b).

No evidence of an association between phthalate exposure and changes in TH levels was detected in children exposed during the use of extracorporeal oxygenation or after using DEHP-tainted foodstuffs (illegal use of DEHP in food from Taiwan); however, in the second group, a reduction in TSH levels was detected just after exposure (Rais-Bahrami et al. 2004, Wu et al. 2013, Tsai et al. 2016a,b). There is evidence in the literature of changes in TH levels, with a negative association of phthalate exposure with serum levels of THs, primarily in girls (Boas et al. 2010, Morgenstern et al. 2017). By contrast, another study showed a different thyroid profile, with sex-specific and metabolite-specific positive changes in TH levels (Weng et al. 2017).

Results from epidemiological cross-sectional studies vary for the different populations analyzed. In the National Health and Nutrition Examination Survey
Disruptors of thyroid function
Sriphrapradang 

There is no evidence that disruption of TH levels in adults; however, other studies, both with animal models and humans, demonstrated that BPA could influence thyroid carcinogenesis, particularly through an association with excess iodine (Zhang et al. 2017a, Zhou et al. 2017). One proposed mechanisms for this process includes interference with the action of ERs, particularly the alpha isoform (Zhang et al. 2017a,b). Additionally, a recent a study based on microarray experiments, showed that BPA exposure impairs cellular defense against DNA damage, resulting in follicular cells being more likely to die or develop a genetic impairment (Porreca et al. 2017).

Pesticides

Many toxic effects of thyroid-disrupting pesticides have been recognized at various stages of development in various adult animal models and in humans. The effects on humans have mainly focused on somatic changes, as well as changes that can be inherited by subsequent generations through epigenetic mechanisms of transgenerational inheritance (Tabb & Blumberg 2006, Andersen et al. 2008, Schug et al. 2011).

Human epidemiological studies have shown that genetic nucleotide polymorphisms in the enzyme paraoxonase 1 (PON1) gene, which encodes a serum enzyme with antioxidant and detoxification properties (Furlong et al. 2005, Eskenazi et al. 2014), can alter the activity of the enzyme and may determine the level of sensitivity to pesticide toxicity (Furlong et al. 2010). Changes in serum lipid and lipoprotein concentrations occur frequently in thyroid dysfunction; in addition, a significant reduction in PON1 activity was observed in both hyperthyroid and hypothyroid patients (Azizi et al. 2003). This supports the idea that pesticides act as thyroid disruptors in humans, and their potential detrimental effects are more pronounced in individuals who are genetically more susceptible to thyroid dysfunction (Lacasana et al. 2010).

Some pesticides have long been banned in many countries but are still present in the environment, such as DDT, HCB and chlorpyrifos (CPF), and many of these have been examined and investigated for their thyroid-disrupting abilities (Pearce & Braverman 2009). In vitro studies on DDT exposure showed that DDT inhibits TSH release, mainly by affecting cAMP production at the post-receptor step (Santini et al. 2003), and DDT can inhibit the activity of TSHR (Rossi et al. 2007, 2009, 2018). In addition, it has been demonstrated that, in rats, a low dose of DDT increases the concentration of T3 and reduces the level of TSH (Yaglova & Yaglov 2014). Furthermore, the stable metabolite of DDT, p,p′-DDE, has also been investigated as a thyroid
disruptor in a study of male Mexican floriculture workers who presented with low serum levels of THs and no significant changes in TSH levels (Blanco-Munoz et al. 2016).

In a study of prenatal HCB exposure, the effect of HCB depended on the genotype of the gene encoding the D1 (Dio1) enzyme, suggesting that D1s play a role in the disruption of THs following exposure to HBC (Llop et al. 2017). Similar studies have shown that diminished T3 levels caused by HBC exposure during pregnancy can interfere with the TH levels in early newborns and affect neurodevelopment (Takser et al. 2005, Maervoet et al. 2007, Berg et al. 2017).

CPF is an organochlorine pesticide commonly used in households and agriculture in the United States. CPF can cross the placental barrier and mainly acts by inhibiting acetylcholinesterase. Even at low doses, this chemical can cause endocrine disruption related to neurodevelopmental defects, resulting in decreased IQ scores and impaired memory skills in children (Crumpton et al. 2000), as well as reductions in serum T4 levels (De Angelis et al. 2009, Zhang et al. 2013).

A cross-sectional study of spray paint workers showed that TSH levels were elevated in this group of workers and that T4 levels were suppressed, while T3 levels were not altered (Zaidi et al. 2006). Furthermore, the evaluation of serum samples from greenhouse workers exposed to pesticides between spring and the fall showed reductions in free T4 and free T3 levels and an increase in TSH levels (Toft et al. 2006).

Glyphosate-based herbicides (GBHs) are among the most widely used pesticides in the world, and they cause several types of endocrine disruption (Toft et al. 2006, Bai & Ogbourne 2016). de Souza demonstrated that glyphosate could also disrupt the HPT axis (Fig. 1) (de Souza et al. 2017). The hormonal profiles of the exposed group showed decreased TSH concentrations, with no variation in the levels of THs. In addition, hypothalamic gene expression analysis of the exposed group revealed that there was a reduction in the expression of genes encoding D1 (Dio2) and 3 (DIO3), and TH transporters Slco1C1 (formerly Oatp1C1) and Slc16a2 (formerly Mct8). On the other hand, in the pituitary of the exposed group, Dio2, Thr (Thra1 and Thrβ1), and Slc16a2 genes were expressed at higher levels. Thus, the role of GBH exposure on HPT axis disruption should be considered in populations exposed to these herbicides (de Souza et al. 2017).

Dimethoate is an organophosphate insecticide used to kill mites and aphids and is applied to crops such as citrus, cotton, fruit, olives, potatoes, tea, tobacco and vegetables. Dimethoate was classified as an endocrine disruptor after studies showed that the treatment of lactating rats with the insecticide resulted in reduced TH secretion in pups, as well as increased plasma T3 and T4 levels and reduced iodine uptake (Mahjoubi-Samet et al. 2005).

Fipronil is used worldwide as an insecticide. It is most commonly applied to crops such as corn, sunflowers, apples, rice and beans and is also used for household and veterinary pest control (Tingle et al. 2003). It accumulates in adipose tissue and the brain (Hainzl & Casida 1996, Simon-Delso et al. 2015). An in vitro study using fipronil and its metabolites showed that fipronil sulfone had anti-THR alpha activity (Lu et al. 2015). It also has been associated with an increase in the incidence of thyroid tumors in rats, decreased plasma T4 concentrations and increased T4 clearance in the rat liver, but these effects have not been observed in occupationally exposed humans (Takagi et al. 2002, Blair et al. 2005, Leghait et al. 2009, Herin et al. 2011). These different results might be related to the exposure dosage of the toxicant and the metabolites present in different environments (Leghait et al. 2010).

Ioxynil (IOX) is an iodine-containing herbicide used for the control of weeds (Takahashi et al. 2010). Ioxynil has been shown to bind to the TH transporter protein TBPA, in different species (Ogilvie & Ramsden 1988, Akiyoshi et al. 2012). It has also been shown to cause epigenetic changes in DNA, resulting in post-receptor changes (Otsuka et al. 2014).

Mancozeb is an agricultural fungicide that, at different oral doses, shows the ability to reduce iodine uptake and reduce serum T4 levels in dams and adult rats (Kackar et al. 1997, Axelstad et al. 2011). In addition, a study of the thyroid axis using adult male birds exposed to Mancozeb for 30 days showed an increase in thyroid size, an increase in plasma TSH levels and a decrease in T4 and T3 levels, mainly during the breeding phase (Pandey & Mohanty 2015).

Other widely used pesticides as acetochlor, amitrole and cyhalothrin have been tested in zebrafish and amphibian models and have been shown to affect the development and metamorphosis (Crumpton et al. 2002, Helbing et al. 2006, Pan et al. 2011, Tu et al. 2016), as well as the expression of a large group of genes that could affect HPT axis and thyroid function (Li et al. 2009, Pan et al. 2011, Tu et al. 2016, Yang et al. 2016, Chang et al. 2018). However, studies in mammals and humans are necessary to further study the impact of the use of these pesticides.

Thus far, compelling evidence has been presented on pesticides as endocrine disruptors; however, more
toxicology screening is needed to identify the toxicity levels and elucidate the mechanisms underlying the effects of these pesticides.

**Polybrominated diphenyl ethers**

There are over 200 brominated flame retardant (BFR) compounds that are used in many industrial applications (de Wit 2002, de Wit et al. 2006, Kabir et al. 2015). Polybrominated diphenyl ethers (PBDEs) are the most common chemicals used BFRs (Mazdai et al. 2003, Streets et al. 2006, de Wit et al. 2010, Covaci et al. 2011). As a consequence, PBDEs are present in plastics, paints, textiles, furniture and other items. In general, they are commercially available in mixtures classified as pentaBDE, octaBDE and decaBDE (Talsness 2008). The compound 2,2′,4,4′,5-pentaBDE (BDE-99) in one of the most prevalent congeners found in humans, and the presence of PBDEs in pregnant women, the accumulation and transfer of this compound from the mother to the infant through the placenta and breast milk has been a concern of society for more than 10 years (Gomara et al. 2007, Schuhmacher et al. 2009).

Several published studies have shown a correlation between PBDE levels and changes in thyroid function to different degrees (Turyk et al. 2008, Chevrier et al. 2010, 2011, Zhang et al. 2010, Lin et al. 2011, Zota et al. 2011). How PBDE affect thyroid function is still unclear. It has been suggested that due to the chemical nature of PBDEs, this chemical could bind directly to THR and affect the expression of T3 target genes (Fig. 2). In an in vitro study, Ren demonstrated that lower-brominated OH-PBDE bind the inner side of the THR-binding pocket, acting as agonists (Ren et al. 2013). Additionally, as hydroxylated metabolites of PBDE, OH-PBDEs are structurally similar to T3 and can compete to bind to THR (Kojima et al. 2009, Schreiber et al. 2010). However, the relationship between PBDEs and thyroid function was suggested based on the cognitive alterations, as many studies showed cognitive alterations associated with impaired TH function (Bowers et al. 2015). Single doses of 2,2′,4,4′-teraBDE (BDE-47) on PND 10 (Dingemans et al. 2007) or chronic administration of decaBDE (BDE 209) during rat gestation (Xing et al. 2009) caused reductions in levels of a key factor in learning and memory, brain-derived neurotrophic factor (BDNF), which is regulated by THs (Koibuchi et al. 1999, Gilbert & Lasley 2013, Shulga & Rivera 2013). Blanco and collaborators treated pregnant rats from embryonic day 6 (E6) to PND 21 and observed a delay in the spatial learning task in the water maze concomitant with decreased BDNF levels in the hippocampus (Blanco et al. 2013). Effects on cognitive function, a downregulation of BDNF and a concomitant decrease in TH serum levels have been reported in several papers (Kodavanti & Derr-Yellin 2002, Darnerud et al. 2007, Bowers et al. 2015).

However, there is controversy regarding the effect of the PBDEs on serum TH levels in animal models depending on the specific chemical, administration time and dose, as well as the hormone that is being evaluated. The majority of the papers found a decrease in total serum T4 levels (Stoker et al. 2004, Kuriyama et al. 2007, van der Ven et al. 2008, Blanco et al. 2013, Kim et al. 2013, Bowers et al. 2015) and total and free T3 levels (Stoker et al. 2004, van der Ven et al. 2008, Blanco et al. 2013, Bowers et al. 2015). A few papers have shown an increase in TH level (Reverte et al. 2014) or no correlation at all (Huang et al. 2014).

The decrease of serum TH levels could be explained by impaired TH biosynthesis impairment (Fig. 2). However, very few studies have found changes in the thyroid. Rats exposed to BDE-47 showed a significant increase in the presence of cellular debris in the follicular lumen (+71%), together with a decrease in iodide uptake (Maranghi et al. 2013, Wu et al. 2016). However, no effect on TPO activity was observed (Wu et al. 2016).

As discussed before, the effects of THs may be affected by cellular metabolism of T4 and T3, as well as interactions with THR (Fig. 2). Several papers studied alterations in D1 activity and THR expression in different animal models, such as birds, zebrafish and others organisms (Butt et al. 2011, Butt & Stapleton 2013, Francois & Verreault 2018). However, few studies have addressed these points in rodent models.

In vitro, PBDEs decreased DIO1 activity in liver cells (Butt et al. 2011) and DIO2 activity in cultured human glial cells (Robert et al. 2015). The possible mechanism for this inhibition was suggested by Marsan and Baise who showed that OH-PBDEs compete with THR through D1s (Marsan & Bayse 2017).

Although PBDEs consistently cause a decrease in the serum levels of THs in rodent models, in humans, many epidemiological papers present inconsistent results (Turyk et al. 2008, Chevrier et al. 2010, 2011, Han et al. 2011, Lin et al. 2011, Kim et al. 2012, 2013, 2015). The differences in the evaluations of cause and effect from these contradictory results can be explained by the highly complex correlations that are possible with over 200 different PBDEs. For example, serum T3 levels were positively correlated with BDE-99 and BDE-209 and negatively correlated with BDE-17, BDE-28, BDE-47, BDE-183 and pentaPBDEs (Huang et al. 2014).
In a Canadian Study evaluating pregnant women, there was a negative association between PBDEs and serum total and free T3 levels (Abdelouahab et al. 2013). In contrast with these studies, Bloom and collaborators observed an increase of free T3 levels with an increase of PBDEs in older women (Bloom et al. 2014). On the other hand, Leonetti found no correlation between BRFs and THs in the placenta (Leonetti et al. 2016). However, a positive association with HO-tetraBDEs and TSH was found in thyroid cancer patients with a decrease in free T4 levels (Liu et al. 2017).

The explanation for low levels of THs could be decreases in biosynthesis and secretion by the thyroid caused by direct effects on the gland or a decrease in TSH stimulation of the thyroid (Fig. 1). However, there are once again many inconsistencies in the results. TSH was correlated with BDE-17, BDE-28, BDE-47 and BDE-183 and inversely correlated with BDE-99 (Huang et al. 2014). However, in cancer patients, Liu and collaborators showed a positive correlation of HO-tetraBDEs and TSH (Liu et al. 2017).

It is important to note that TH levels in blood are tightly regulated within an individual; hence, intra-individual variation would often be negligible compared to the inter-individual variation or the wide range of reference values. Therefore, it has been suggested by some authors that small changes in TH levels in response to exposure to environmental chemicals may not be easy to detect in small human population (Boas et al. 2012). Recently, to try to end this controversy, a meta-analysis was published, indicating that the effect of PBDEs on thyroid function depends on the PBDE exposure dose and its serum levels, suggesting a u-shaped curve (Zhao et al. 2015).

**Take home message**

Here, we reviewed a relevant group of thyroid disruptors to which the general population is exposed to daily. These compounds are present not just in industrialized food but also in fresh preparations. Controlling the consumption of natural bioactive compounds by a population is difficult so increase the population’s awareness of their potential undesirable effects is fundamental. On the other hand, there is an enormous amount of data proving the deleterious effects of the chemicals, and as a consequence, several of them have already been banned from industrial processes or have an identified reference exposure dose. It is important to note that these reference exposures dose values are based on regulatory and often non-comprehensive tests that not always take into account how the endocrine system works, especially the thyroid gland.

THs are vital for neurological development in the early stages of life (Bernal 2017), and several studies have shown that low doses of these chemicals, closer to the usual environmental contamination dose, have effects on the endocrine system, particularly in developmentally vulnerable windows (Vandenberg et al. 2012).

Thyroid function is essential during all stages of life as it affects growth, development, metabolism and the cardiovascular and immune systems, and the continuous exposure to natural and artificial thyroid disruptors will deeply affect quality of life.

**Declaration of interest**

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this review.

**Funding**

This work was supported by grants from the Ministério da Ciência, Tecnologia e Inovação, Conselho Nacional de Desenvolvimento Científico e Tecnológico, CNPq (grant numbers: 304667/2016-1, 305427/2013-0, 422441/2016-3); Fundação Carlos Chagas Filho de Amparo ao Pesquisa do Estado do Rio de Janeiro (grant numbers: CNE 2015/E26/202924/2015, CNE 2015/E26/203.190/2015) and Fundação de Amparo a Pesquisa do Estado de São Paulo (grant number: 2013/26851-7).

**References**


Disruptors of thyroid function

Aulerich RJ & Ringer RK 1977 Current status of PCB toxicity to mink, and effect on their reproduction. *Archives of Environmental Contamination and Toxicology* 6 279–292. (https://doi.org/10.1007/BF02097769)


Butt CM & Stapleton HM 2013 Inhibition of thyroid hormone sulfotransferase activity by brominated flame retardants and halogenated phenolics. *Chemical Research in Toxicology* 26 1692–1702. (https://doi.org/10.1021/tx300432k)


tissue-specific expression of thyroid hormone receptor and target genes in rats. Journal of the Science of Food and Agriculture 96 2889–2895. (https://doi.org/10.1002/jsfa.7460)


Gayathri NS, Dhanya CR, Indu AR & Kupur PA 2004 Changes in some hormones by low doses of di (2-ethyl hexyl) phthalate (DEHP), a commonly used plasticizer in PVC blood storage bags and medical tubing. Indian Journal of Medical Research 119 139–144.


Helbing CC, Oraska K & Ji L 2006 Evaluation of the effect of acetylcholine on thyroid hormone release from feto-placental explant cultures but have limited influence on selected thyroid cell functions in vitro. Thyroid 21 701–706. (https://doi.org/10.1089/thy.2010.0449)


Huang PC, Tsai CH, Liang WY, Li SS, Huang HB & Kuo PI 2016 Early phthalates exposure in pregnant women is associated with alteration of thyroid hormones. PLoS ONE 11 e0159398. (https://doi.org/10.1371/journal.pone.0159398)


Huang HB, Pan WH, Chang JW, Chiang HC, Guo YL, Jaakkola JJ & Huang PC 2017b Does exposure to phthalates influence thyroid function and growth hormone homeostasis? The Taiwan Environmental Survey for Toxics (TEST) 2013. Environmental Research 153 63–72. (https://doi.org/10.1016/j.envres.2016.11.014)

Ibhaizheho K & Kobuchi N 2011 Thyroid hormone receptor-mediated transcription is suppressed by low dose phthalate. Nigerian Journal of Physiological Sciences 26 143–149.

Iwamuro S, Yamada M, Kato M & Kikuyama S 2006 Effects of bisphenol A on thyroid hormone-dependent up-regulation of thyroid hormone

Janjua NR, Mortensen GK, Andersson AM, Kongsjoh B, Skakkebaek NE & Wulf HC 2007 Systemic uptake of diethyl phthalate, dibutyl phthalate, and butyl paraben following whole-body topical application and reproductive and thyroid hormone levels in humans. Environmental Science and Technology 41 5564–5570. (https://doi.org/10.1021/es062875l)


Johns LE, Ferguson K, McElrath TE, Mukherjee B & Meeker JD 2016 Associations between repeated measures of maternal urinary phthalate metabolites and thyroid hormone parameters during pregnancy. Environmental Health Perspectives 124 1808–1815.

Jump DB, Clarke SD, MacDougald O & Thelen A 1993 Polyunsaturated fatty acids inhibit S14 gene transcription in rat liver and cultured hepatocytes. PNAS 90 8454–8458. (https://doi.org/10.1073/pnas.90.18.8454)


Kim UJ, Kim MY, Hong YH, Lee DH & Oh JE 2012 Assessment of impact of internal exposure to PBDEs on human thyroid function – comparison between congenital hypothyroidism and normal paired tissue distribution and thyroid hormone levels. Environmental Health Perspectives 120 369–375. (https://doi.org/10.1093/biolreprod.66.2.272)


Koibuchi N, Fukuda H & Chin WW 1999 Promoter-specific regulation of the brain-derived neurotropic factor gene by thyroid hormone in the developing rat cerebellum. Endocrinology 140 3955–3961. (https://doi.org/10.1210/endo.140.9.6997)


Lee S, Kim C, Yoon H & Choi K 2017 Thyroid hormone disrupting potentials of bisphenol A and its analogues – in vitro comparison study employing rat pituitary (GH3) and thyroid follicular (FRTL-5) cells. Toxicology in Vitro 40 297–304. (https://doi.org/10.1016/j.tiv.2017.02.004)


Leung AM, Korevaar TI, Peeters RP, Zoeller RT, Kohrle J, Duntas LH, Brent GA & Demeneix BA 2016 Exposure to thyroid-disrupting chemicals: a transatlantic call for action. Thyroid 26 479–480. (https://doi.org/10.1089/thy.2016.0077)

Li W, Zha J, Li X, Yang L & Wang Z 2009 Effects of exposure to acetochlor on the expression of thyroid hormone related genes in larval and adult rare minnow (Gobio biros) Aquatic Toxicology 94 87–93. (https://doi.org/10.1016/j.aquatox.2009.06.002)

Li J, Ren S, Han S, Lei B & Li N 2014 Identification of thyroid-receptor antagonists in water from the Guanting Reservoir, Beijing, China. Archives of Environmental Contamination and Toxicology 67 68–77. (https://doi.org/10.1007/s00244-014-0027-5)

Lin SM, Chen FA, Huang YE, Hung LL, Chen LJ, Wu LS, Liu TS, Chang-Chien GP, Chen KC & Chao HR 2011 Negative associations between PBDE levels and thyroid hormones in cord blood. International
Meeker JD, Calafat AM & Hauser R 2007 Di(2-ethylhexyl) phthalate metabolites may alter thyroid hormone levels in men. Environmental Health Perspectives 115 1029–1034. (https://doi.org/10.1289/ehp.9852)
Meeker JD, Calafat AM & Hauser R 2010 Urinary bisphenol A concentrations in relation to serum thyroid and reproductive hormone levels in men from an infertility clinic. Environmental Science and Technology 44 1458–1463. (https://doi.org/10.1021/es902892k)
Mendez W Jr & Eftim SE 2012 Biomarkers of perchlorate exposure are correlated with circulating thyroid hormone levels in the 2007–2008 NHANES. Environmental Research 118 137–144. (https://doi.org/10.1016/j.envres.2012.05.010)
Messa M & Redmond G 2006 Effects of soy protein and soybean isoflavones on thyroid function in healthy adults and hypothyroid patients: a review of the relevant literature. Thyroid 16 249–258. (https://doi.org/10.1089/thy.2006.16.249)
Ortiga-Cevallos TM, Sidhaye AR & Wondisford FE 2014 Thyroid hormone receptors and resistance to thyroid hormone disorders. Nature Reviews Endocrinology 10 582–591. (https://doi.org/10.1038/nrendo.2014.143)
Panda S & Kar A 2003b Water and ethanol extracts of piper nigrum in regulating thyroid function and lipid peroxidation in mice.
Disruptors of thyroid function


Volpi E, Tataranni PA, Pacca V, Fanti F, & Visani L 2000 Thyroid hormones and fear learning but not anxiety are affected in adult apoE-deficient mice. *Physiology and Behavior* **68** 256–263. (https://doi.org/10.1016/s0031-9384(00)00072-9)


© 2019 Society for Endocrinology
Published by Bioscientifica Ltd.
Printed in Great Britain

Downloaded from Bioscientifica.com at 04/03/2019 10:06:02AM
via free access


Wu MT, Wu CF, Chen BH, Chen EK, Chen YL, Shiea J, Lee WT, Chao MC & Wu JR 2013 Intake of phthalate-tainted foods alters thyroid functions in Taiwanese children. PLoS ONE 8 e55005. (https://doi.org/10.1371/journal.pone.0055005)


Received in final form 28 March 2018
Accepted 12 July 2018
Accepted Preprint published online 12 July 2018