A role for steroid sulphatase in intracrine regulation of endometrial decidualisation

Douglas A Gibson1, Paul A Foster2, Ioannis Simitsidellis1, Hilary O D Critchley2, Olympia Kelepouri1, Frances Collins1 and Philippa T K Saunders1

1MRC Centre for Inflammation Research, The University of Edinburgh, QMRI, Edinburgh, UK
2Institute of Metabolism & Systems Research, University of Birmingham, Birmingham, UK.
3MRC Centre for Reproductive Health, The University of Edinburgh, QMRI, Edinburgh, UK

Correspondence should be addressed to D A Gibson: d.a.gibson@ed.ac.uk

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Abstract

In women, establishment of pregnancy is dependent upon ‘fine-tuning’ of the endometrial microenvironment, which is mediated by terminal differentiation (decidualisation) of endometrial stromal fibroblasts (ESFs). We have demonstrated that intracrine steroid metabolism plays a key role in regulating decidualisation and is essential for time-dependent expression of key factors required for endometrial receptivity. The primary aim of the current study was to determine whether sulphated steroids can act as precursors to bioactive sex steroids during decidualisation. We used primary human ESF and a robust in vitro model of decidualisation to assess the expression of genes associated with sulphation, desulphation and transport of sulphated steroids in human ESF as well as the impact of the steroid sulphatase (STS) inhibitor STX64 (Irosustat). We found evidence for an increase in both expression and activity of STS in response to a decidualisation stimulus with abrogation of oestrone biosynthesis and decreased secretion of the decidualisation marker IGFBP1 in the presence of STX64. These results provide novel insight into the contribution of STS to the intracrine regulation of decidualisation.

Key Words
- decidualisation
- steroid sulphatase
- sulphation
- Irosustat
- oestrone

Introduction

Decidualisation is a fundamental process of endometrial remodelling that is required for the establishment of pregnancy. It is associated with unique time-dependent transcriptomic and proteomic changes (reviewed in Gellersen & Brosens 2014, which are reported to be disordered in women with recurrent implantation failure (Ruiz-Alonso et al. 2013, Koot et al. 2016). Whilst the post-ovulatory rise in progesterone acts as an endocrine signal to stimulate decidualisation of oestrogen-primed human endometrial stromal fibroblast (hESF) (reviewed in Gellersen & Brosens 2014), we have demonstrated an important role for local (intracrine) steroid metabolism in fine-tuning the cellular differentiation of hESFs (Gibson et al. 2016b). Importantly, we have established that expression of CYP19A1 (aromatase, the key enzyme required for conversion of androgens to estrogens) as well as AKR1C3 and SRD5A1 (enzymes that convert precursor androgens into testosterone and dihydrotestosterone (DHT), respectively) are altered in a time-dependent manner (Gibson et al. 2013, 2016a). Expression of these...
enzymes results in increased biosynthesis of potent steroid receptor agonists (E2, testosterone, DHT) that in turn regulate the expression of genes important for receptivity and immune-cell-mediated vascular remodelling (Gibson et al. 2013, 2015, 2016a).

Bioavailability of sulphated steroids can affect intratissue concentrations of oestrogens and androgens, which may play a major role in regulating the function of both normal and malignant tissues (Mueller et al. 2015). Hydrolysis of sulphated precursors into biologically active steroids requires the enzyme steroid sulphatase (STS) (Fournier & Poirier 2009). STS primarily converts oestrone sulphate (E1,S) to oestrone (E1) and is expressed in many tissues, albeit at low levels (Miki et al. 2002, Suzuki et al. 2003). In addition to oestrogen-forming activities, STS also has the capacity to hydrolyse DHEAS to DHEA, which may also act as a precursor for the formation of oestrogens and androgens within target tissues (Purohit et al. 2011). The actions of STS are countered by sulphotransferases that promote inactivation and metabolism of steroids following conjugation of a sulphate moiety. Sulphotransferases are a diverse gene family and the isoforms SULT1A1, SULT1E1, SULT2A1 and SULT2B1 have been associated with sulphation of steroids (Mueller et al. 2015). SULT1E1 primarily catalyses the sulphation of oestrogens, although SULT1A1 is also reported to have this action (Gamage et al. 2005). Sulphation requires the co-substrate PAPS (adenosine 3’-phosphate 5’-phosphosulfate) that provides the universal sulphate donor compound for all sulphotransferase reactions. PAPS is synthesised via one of two PAPS synthase isoforms PAPSS1 and PAPSS2 (Schröder et al. 2012). Uptake and excretion of sulphated steroids from cells is regulated by membrane transporters which are reported to be expressed in endometrial and ovarian cancer cells and tissues (Mueller et al. 2015, Rizner et al. 2017). To date, few studies have examined the expression of STS, sulphotransferases, PAPSS or membrane transporters in endometrial tissues.

STS immunoexpression has been detected in both stromal and epithelial cells in the endometrium across the menstrual cycle (Dassen et al. 2007). In a study using 12 endometrial cancer samples and adjacent endometrium, expression of both STS and SULT1E1 mRNAs were higher in the normal endometrium than in the malignant cells (Smuc et al. 2006). Notably, in both benign and normal endometrial tissues, concentrations of mRNAs for STS were higher than SULT1E1 (Smuc et al. 2006), which may favour an increase in the bioavailability of active steroid receptor agonists in the endometrium. Indeed, studies using STX64 (Irosustat), an irreversible inhibitor of STS (Day et al. 2009), suggest that STS can regulate formation of oestradiol (E2) in the uterus (Colette et al. 2011). STS inhibitors are also reported to decrease the oestrogenic growth of hormone-dependent tissues, including endometrial cancer tissues (Foster et al. 2008) and proliferation of human endometrial xenografts (Colette et al. 2011). Recently, Sinreih et al. extended their studies on endometrial cancer steroid intracrinology and concluded that E2 was formed from E1,S via the sulphatase pathway rather than via aromatase in endometrial cancer tissues (Sinreih et al. 2017). In contrast, previous studies in both human and mouse suggest intracrine regulation of the normal endometrium occurs via aromatase-dependent biosynthesis of oestrogens (Das et al. 2009, 2012, Gibson et al. 2013).

In the current study, we sought to complement and extend our previous investigations by assessing the contribution of STS to intracrine regulation of decidualisation. We used an in vitro model of decidualisation to assess the expression of genes associated with sulphation, desulphation and transport of sulphated steroids in hESFs and assessed the impact of the STS inhibitor STX64 (Irosustat). Our results provide new insight into the mechanisms that contribute to generation of an oestrogen-rich microenvironment during the establishment of pregnancy.

**Materials and methods**

**Human studies**

Primary human endometrial tissue (proliferative phase, n=9) was obtained from women undergoing surgery for non-malignant gynaecological conditions. None of the women were receiving hormonal therapy or suffering from endometriosis. Primary hESFs were isolated from proliferative phase endometrium and cycle phase determined as reported previously (Bombail et al. 2010). Briefly, endometrial tissue was minced using scalpel blades, followed by DNAse/collagenase digest for 2h at 37°C. The tissue homogenate was then sequentially drained through 70µm and 40 µm membrane filters to separate hESF from the glandular epithelium. Isolated hESFs were washed in warmed PBS and cultured in RPMI 1640, supplemented with 10% foetal calf serum (FCS) at 37°C in 5% CO2 and maintained for a maximum of five passages. Forty-eight hours prior to experimentation, hESF culture media were changed to phenol red-free RPMI 1640 supplemented with 10% charcoal-stripped FCS (CSFCS). Decidualisation was induced by addition of decidualisation (DEC) media (phenol red-free RPMI 1640, 2% CSFCS, 0.1 mg/mL 8-Br-cAMP (Sigma, B5386), 1 µM progesterone (Tocris, Abingdon, UK; Cat no. 2835)). Some cell cultures were
supplemented with the STS inhibitor Irosustat (STX64; 10μM, Sigma S1950) for the duration of the culture period. Control cultures were incubated with phenol red-free RPMI 1640, 2% CSFCS and equivalent volume of vehicle control (DMSO). 2 × 10³ hESFs were seeded per well of six-well plate, treatments were in duplicate and assessed in minimum of three individual patients for all treatments. For experiments assessing expression and activity of sulphation enzymes/transporters, cells from n = 6 patients were assessed. For experiments assessing the impact of STX64, cells from n = 3 patients were used. Each sample was assayed in duplicate and average values were analysed. To assess the time-dependent accumulation of secreted products, treatments were maintained for the duration of each time point. hESFs were treated for 24h (1 day), 48h (2 days), 4 days or 8 days as indicated; cells were treated in parallel with dedicated samples for each time point and patient. The expression of genes associated with sulphation, desulphation and transport of sulphated steroids was assessed by RT-qPCR.

**Assessment of mRNA**

Isolation of mRNAs, preparation of cDNAs and analysis by RT-qPCR was performed according to standard protocols (Bombail et al. 2010); samples were analysed by the comparative ΔΔCt method with CYC (cyclophilin) as an internal control. Primers/probes are given in Supplementary Table 1 (see section on supplementary data given at the end of this article).

**ELISA**

Insulin-like growth factor-binding protein 1 (IGFBP1), E1 and E2 in culture supernatants were determined by ELISA as described previously (Gibson et al. 2013, 2016a). E1 and E2 were not detected in control supernatants (not shown). Antibody cross-reactivity for E1 and E2 ELISA was <5% for other steroids (Supplementary Tables 2 and 3).

**Measurement of steroid sulfatase activity**

STS activity was determined in cell lysates as described previously (Purohit et al. 1997). Briefly, cells were lysed in RIPA buffer and protein concentration measured by BCA assay. Hundred micrograms of protein were incubated for 4h with PBS containing [6,7-^3^H] E$_1$S (4 × 10^6 dpm) adjusted to a final concentration of 20μM with unlabelled E$_1$S. [4-^1^4^C] E$_1$ (1 × 10^4 dpm) was used to monitor procedural losses. E$_1$ was separated from E$_1$S by tolune partition and ^3^H and ^1^4^C radioactivity was measured by liquid scintillation spectrometry. Results were expressed as E1 formed pmol/h/mg protein.

**Statistics**

Statistical analysis was performed using GraphPad prism. Two-way ANOVA was used to determine the significance between treatments in grouped data; interaction between time and treatment was assessed, Sidak’s multiple comparisons test was used to assess the differences between treatments for individual time points. Non-parametric testing was utilised where sample sizes were insufficient to confirm normality of data distribution; Mann–Whitney test was used to assess differences between treatments at each time point for RT-qPCR data. Criterion for significance was P<0.05. All data are presented as mean ± s.e.m.

**Study approval**

Written informed consent was obtained from all subjects prior to surgery; ethical approval was granted by the Lothian Research Ethics Committee (LREC/07/S1103/29 and LREC 10/S1402/59). Methods were carried out in accordance with NHS Lothian Tissue Governance guidelines. Studies using these cells have previously been reported in (Gibson et al. 2018).

**Results**

**Expression of sulphation and desulphation enzymes during decidualisation of human ESF**

We assessed expression of STS as well as members of the sulphotransferase gene family in cells recovered 1–8 days after incubation with control (VC) or DEC culture media (Figs 1 and 2). Expression of STS was increased at all time points in hESF stimulated with DEC; results reached statistical significance in samples recovered on Day 1 (Fig. 1A; n = 6; P<0.0001), 2 (Fig. 1B; n = 6; P<0.0001) and 4 (Fig. 1C; n = 6; P<0.01). To complement these findings, STS activity was assessed in cell lysates and was significantly increased in lysates from hESF treated with DEC media compared to VC after 4 days of treatment (Fig. 1E, n = 6; P<0.05).

To determine whether E1 and E2 formed by decidual cells (Gibson et al. 2013) might also be subject to inactivation, we also assessed expression of sulphotransferases in the same samples. In these experiments, expression of SULT1E1 and SULT2A1 mRNAs...
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Expression of PAPSS isozymes during decidualisation of hESF

To complement and extend the studies on sulphotransferase enzymes, we also assessed expression of PAPSS isozymes PAPSS1 and PAPSS2. Expression of mRNAs encoded by PAPSS1 (Fig. 3A) was increased in DEC-treated hESF compared to controls on days 1 (n=6; P<0.05) and 4 (n=6; P<0.01) days, whereas PAPSS2 mRNA concentrations were only higher on day 1 (n=6; P<0.01) and unchanged at other time points (Fig. 3B).

Expression of ATP-binding cassette transporters during decidualisation of human ESF

We assessed expression of cellular transporters ABCC1 and ABCC4, which are associated with efflux of sulphated steroids. Expression of mRNAs encoding ABCC1 was increased in a time-dependent manner in decidualised hESF (Fig. 4A). ABCC1 mRNA expression was significantly increased in DEC compared to controls at each time point; 1 (n=6; P<0.0001), 2 (n=6; P<0.0001) and 4 (n=6; P<0.01) days, with the greatest increase in expression detected after 8 days (n=6; P<0.0001). In contrast, ABCC4 mRNA expression was significantly decreased in hESF treated with DEC compared to controls at 2 (n=6; P<0.01), 4 (n=6; P<0.0001) and 8 (n=6; P<0.01) days (Fig. 4B). Expression of organic anion-transporting
polypeptide transporters (SLC gene family; SLC01A2, SLC01B1, SLC01B3 and SLC02B1), which mediate influx of sulphated steroids were not detected in either control or decidualised hESF (not shown).

In light of these results and as a complement to our previous studies in which we demonstrated that both E1 and E2 are secreted into culture media during our decidualisation protocol (as determined by ELISA (Gibson et al. 2013)), we used sensitive LC-MS to assess the concentrations of E1S or E2S in conditioned media from hESF, however, neither steroid was detected in either the control or DEC samples (not shown; LLOD for E1S and E2S = 0.5 ng/mL).

**Irosustat (STX64) disrupts bioavailability of oestrogens and alters decidualisation responses**

We investigated the potential importance of STS activity during decidualisation of hESF by assessing the impact of the selective, irreversible STS inhibitor Irosustat (STX64). hESFs were stimulated with DEC media or co-incubated with STX64 (DEC STX) for 1, 2, 4 or 8 days. Incubation with STX64 was associated with altered production of oestrogens from hESF and mean concentrations of secreted E1 were decreased at each time point in hESF treated with DEC STX compared to DEC alone (Fig. 5A). The impact of STX on E1 secretion was time-dependent with the greatest reduction detected after 1 day (32% decrease in E1; n=3 patients, P<0.05) but at later time points, there was no significant difference between hESF treated with DEC and DEC STX. In contrast to E1, concentrations of secreted E2 were not altered between hESF treated with either DEC alone or DEC STX (Supplementary Fig. 1). We next assessed if altered bioavailability of oestrogens affected decidualisation of hESF. Notably, STX64 altered secretion of the decidualisation marker IGFBP1; with concentrations reduced at each time point and significantly decreased after 1 day in hESF treated with DEC STX compared to DEC alone (Fig. 5B, n=3 patients, P<0.05).

**Discussion**

Oestrogens within tissues can be derived from two pathways; *de novo* synthesis, which requires aromatase, and activation of circulating sulphated steroids which requires
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However, in the presence of the aromatase inhibitor letrozole, both decidualisation and implantation were impaired indicating that ovarian oestrogens were dispensable for decidualisation (Das et al. 2009). In further experiments using a mouse model of artificially induced decidualisation, they went on to show that factors required for stromal remodelling and neovascularisation were dependent on aromatase-dependent intrauterine de novo biosynthesis of oestrogens (Das et al. 2009, 2012). Our recent studies have extended these observations in mice and found evidence for complementary mechanisms in human endometrial stromal fibroblasts. We have shown that hESF have the capacity to utilise DHEA as a substrate and that hESF synthesise aromatisable androgens during decidualisation (Gibson et al. 2013, 2016a, 2018). We have demonstrated that decidualisation is characterized by an increase in the expression and activity of aromatase (CYP19A1) leading to local biosynthesis of both E1 and E2 within the microenvironment of the endometrial stroma (Gibson et al. 2013). We have also demonstrated that oestrogens regulate cellular cross-talk within the endometrium during the establishment of pregnancy by directly regulating immune cell function and promoting vascular remodelling (Gibson et al. 2015). Given that oestrogens play such an essential role in modulating the endometrial environment, in the present study, we sought to further examine the mechanisms that regulate the availability of oestrogens by assessing the contribution of STS to intracrine oestrogen bioavailability during decidualisation.

We speculated that in addition to the classic de novo biosynthesis pathway, oestrogens may also be synthesised via de-sulphation of E$_1$S. Using a robust model of in vitro decidualisation and hESF, we detected increased expression and activity of STS. Notably, we found that the STS inhibitor STX64 (Irosustat) had a time-dependent impact on decidualisation and decreased secretion of IGFBP1 from days 1 to 4 of the time course. Decreased secretion of IGFBP1 was associated with concomitant decreases in E1 concentrations suggesting STS-dependent regulation of intracrine oestrogens occurs during decidualisation. Notably, the aromatase inhibitor letrozole is reported to completely inhibit uterine oestrogen production in ovariectomised mice (Das et al. 2009). However, in our previous study, we co-incubated hESF with the aromatase inhibitor letrozole during decidualisation, and this only reduced secretion of E1 and E2 by approximately 65% (Gibson et al. 2013). In the current study, E1 synthesis was reduced by up to 30% during decidualisation consistent with both STS- and aromatase-dependent production of E1 as growth and development of implanted embryos, (Das et al. 2009).

### Figure 5

Irosustat (STX64) disrupts oestrogen bioavailability and alters decidualisation responses. The impact of the potent irreversible STS inhibitor STX64 (STX) on decidualisation of hESF was investigated following treatment for 1–8 days as indicated. (A) Secretion of E1 was significantly decreased after 1 day in hESF treated with DEC + STX compared to DEC alone (P < 0.05). Two-way ANOVA; global analysis time factor P = 0.0158, treatment factor P = 0.0111. Sidak’s multiple comparisons test; treatment effect significantly different day 1 P < 0.05. (B) Concentrations of IGFBP1 tended to be lower at all time points in hESF treated with DEC + STX compared to DEC alone. STX significantly decreased secretion of the decidualisation marker IGFBP1 after 1 day of treatment (P < 0.05). Two-way ANOVA; global analysis time factor P = 0.0026, treatment factor P = 0.0008. Sidak’s multiple comparisons test; treatment effect significantly different day 1 P < 0.05. E1 and IGFBP1 secretion was determined by ELISA. n = 3 patients, duplicate treatments. *P < 0.05. DEC, decidualisation media; E1, oestrone; hESF, human endometrial stromal fibroblasts; IGFBP1, insulin-like growth factor-binding protein-1.
during decidualisation. Notably, the effect of STX64 on 1 concentrations was most pronounced at day 1 but absent by day 8. This reflects our previous observations regarding the role of intracrine sex steroids during decidualisation, whereby modulation of local androgen action resulted in time-dependent changes in the expression of decidualisation and implantation-associated factors in the presence of the antiandrogen flutamide (Gibson et al. 2016a). Thus, if sulphated oestrogens are contributing to the ‘oestrogen pool’ during decidualisation, this effect may also be temporally regulated, with increased de-sulphation of steroid precursors during the early stages of decidualisation preceding aromatase-dependent synthesis of oestrogens during the latter stages of the differentiation process. Our results therefore suggest sulphation pathways contribute to regulation of decidualisation via rapid as well as gradual mechanisms. Taken together, our findings suggest bioavailability of intracrine oestrogens modulates initiation of stromal decidualisation and that tightly controlled regulation of intracrine steroids, via sulphation and export, is required as decidualisation progresses for appropriate paracrine signalling to immune and vascular endothelial cells within the tissue microenvironment during endometrial remodelling.

Notably, the most abundant oestrogen detected in the current analysis and in our previous study (Gibson et al. 2013) was 1. This was surprising as 1 is classically considered to be weak/inactive oestrogen. However, our results are consistent with Huhtinen et al. who performed intra-tissue profiling of human endometrial tissues using LC-MS. They reported increased concentrations of 1 in secretory phase endometrium compared to serum concentrations and that mean intrauterine concentrations of 1 are greater than 2 in the secretory phase (Huhtinen et al. 2012). Although the role of 2 in regulating establishment of pregnancy is well established, the predominance of 1 in the intrauterine environment during the secretory phase may suggest an independent role for 1 in regulating endometrial function. We have previously demonstrated that equimolar concentrations of 1 (10nM) can directly regulate increases in ER-dependent migration of uterine NK cells to a similar extent as 2 (Gibson et al. 2015) and in the current study, it was notable that reduction of 1 concentrations but not 2 by STX64 abrogated secretion of the decidualisation marker IGFBP1. Whether 1 acts as a direct agonist or by modulating intracrine metabolism in this context requires further investigation. However, the findings in the current study provide new evidence that availability of 1 may independently contribute to regulation of the endometrial microenvironment.

Sulphation of steroids is a primary route for metabolism of active oestrogens in healthy peripheral tissues. Previous studies have demonstrated that mRNA expression of the main oestrogen sulphating enzyme SULT1E1 is detected at all stages of the menstrual cycle in total endometrial tissue extracts (Dassen et al. 2007). In the current study, SULT1E1 was not detected in hESF, consistent with classic metabolism studies, which only detected formation of E,S from isolated endometrial epithelial, but not stromal, cells following incubation with E (Liu & Tseng 1979). We did detect mRNA expression of SULT1A1 which is reported to be expressed in the endometrium (Rubin et al. 1999) and also has oestrogen sulphating activity (Falany & Falany 1996), albeit in the micromolar range. Notably, increased concentrations of sulphotransferases (SULT1A1, SULT2B1) were detected concurrent with increased expression of PAPSS isozymes; however, we could not detect any sulphated oestrogens in conditioned media from hESF. There are limited data on the expression of transmembrane transporters that mediate cellular influx and efflux of sulphated steroids in the endometrium. Nishimura and Naito assessed mRNA expression of 46 ABC transporters and 108 SLC transporters in a range of tissues including 3 human uterine samples (Nishimura & Naito 2005). Notably, relative expression of mRNAs (ratio of target genes to PPIA housekeeping gene) encoding SLC01A2, SLC01B1 and SLC02B1 as well as ABCC1 and ABCC4 were reported in uterine samples; although expression of SLC isoforms was relatively low compared to other tissues (Nishimura & Naito 2005). In the current study, we could not detect expression of SLC isoforms in hESF, however, ABC transporters (ABCC1 and ABCC4) were detected and dynamically regulated consistent with a possible role in export of sulphated steroids during decidualisation. It is a limitation of the current study that only the contribution of stromal compartment was assessed. Given the reported expression and activity of SULT1E1 in endometrial epithelial cells (Liu & Tseng 1979), further studies are needed to assess the contribution of epithelial cells and stromal–epithelial interactions in regulating oestrogen bioavailability within the endometrium. Our data support STS activity is prominent in the stromal compartment during decidualisation and additionally we found some evidence for time-dependent increases in expression of sulphotransferase enzymes. We did not detect any secretion of sulphated oestrogens in conditioned medium for hESF, which may suggest that any sulphated oestrogens formed during decidualisation are subject to direct hydrolysis by STS.
In our study, we used primary human cells and a well-characterised model of in vitro decidualisation; however, it is conceivable that adaptive metabolism and alternative pathways may be more prominent in disease states such as endometriosis and endometrial cancer where increased expression of STS has been reported (Utsunomiya et al. 2004, Piccinato et al. 2016, Sinreih et al. 2017). It is notable that balanced availability of oestrogens is required to regulate establishment of pregnancy. Increasing or decreasing E2 outside the normal physiological range is detrimental to implantation in mice (Ma et al. 2003). Furthermore, supraphysiological levels of E2 and progesterone as a result of controlled ovarian hyperstimulation during in vitro fertilization treatment protocols is associated with low implantation rates, possibly as a result of an altered steroid milieu (Pellicer et al. 1996). ‘Out of phase’ endometrial remodelling is a common feature of implantation failure but whether dysregulation of endometrial intracrine metabolism occurs in sub/infertility requires further investigation. Notably, drugs that target steroid metabolism such as aromatase inhibitors, STS inhibitors, DASI (dual aromatase–STS inhibitors) and 17BHSD1 inhibitors may affect endometrial function, and these potential actions should be considered in future therapeutic applications of these drugs in reproductive-aged women.

Conclusions

In the current study, we assessed the expression of sulphation and desulphation enzymes, sulphate donors and transmembrane transporters in hESFs during a time course of in vitro decidualisation. We found that inhibition of STS activity disrupted bioavailability of oestrogens and inhibited decidualisation responses consistent with stromal utilisation of sulphated steroids as precursors to active hormones during decidualisation. Elucidation of the complex intracrine metabolism of steroids within the endometrium during decidualisation will be critical to understanding the relevance of these findings to reproductive health and disease. The results of the current study provide new insight into the contribution of sulphated steroids to the regulation of decidualisation and expand our understanding of intracrine regulation of the endometrium during the establishment of pregnancy.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of this review.

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Supplementary data

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