The LIM homeodomain protein ISL1 mediates the function of TCF7L2 in pancreatic beta cells

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Abstract

Pancreatic β-cell Tcf7l2 deletion or its functional knockdown suggested the essential role of this Wnt pathway effector in controlling insulin secretion, glucose homeostasis and β-cell gene expression. As the LIM homeodomain protein ISL1 is a suggested Wnt pathway downstream target, we hypothesize that it mediates metabolic functions of TCF7L2. We aimed to determine the role of ISL1 in mediating the function of TCF7L2 and the incretin hormone GLP-1 in pancreatic β-cells. The effect of dominant negative TCF7L2 (TCF7L2DN) mediated Wnt pathway functional knockdown on Isl1 expression was determined in βTCFDN mouse islets and in the rat insulinoma cell line INS-1 832/13. Luciferase reporter assay and chromatin immunoprecipitation were utilized to determine whether Isl1 is a direct downstream target of Tcf7l2. TCF7L2DN adenovirus infection and siRNA-mediated Isl1 knockdown on β-cell gene expression were compared. Furthermore, Isl1 knockdown on GLP-1 stimulated β-catenin S675 phosphorylation and insulin secretion was determined. We found that TCF7L2DN repressed ISL1 levels in βTCFDN mouse islets and in the rat insulinoma cell line INS-1 832/13. Wnt stimulators enhanced Isl1 promoter activity and binding of TCF7L2 on Isl1 promoter. TCF7L2DN adenovirus infection and Isl1 knockdown generated similar repression on expression of β-cell genes, including the ones that encode GLUT2 and GLP-1 receptor. Either TCF7L2DN adenovirus infection or Isl1 knockdown attenuated GLP-1-stimulated β-catenin S675 phosphorylation in INS-1 832/13 cells or mouse islets and GLP-1 stimulated insulin secretion in INS-1 832/13 or MIN6 cells. Our observations support the existence of TCF7L2- Isl1 transcriptional network, and we suggest that this network also mediates β-cell function of GLP-1.

Introduction

Following the recognition that TCF7L2 is a type 2 diabetes (T2D) risk gene by intensive and reproducible genome-wide association studies (GWAS) (Florez et al. 2006, Grant et al. 2006), great efforts have been made to explore functions of this Wnt signaling pathway effector in pancreatic β-cells and elsewhere (Schafer et al. 2007, Norton et al. 2011, 2014, Ip et al. 2012, 2015, Kaminska et al. 2012, Jin 2016). Previous in vitro and ex vivo investigations...
have collectively suggested the role of mouse and human TCF7L2 in facilitating β-cell proliferation and regulating β-cell metabolic functions (da Silva Xavier et al. 2009, Shu et al. 2009, 2012). Although the investigations on the three different pancreatic β-cell Tcf7l2−/− mouse models have generated inconsistent observations (Boj et al. 2012, da Silva Xavier et al. 2012, Mitchell et al. 2015), functional knockdown of the Wnt signaling cascade with dominant negative TCF7L2 (TCF7L2DNin humans and mice) in pancreatic β-cells by our group and by another team allowed us to suggest that TCF7L2 in mice is important for β-cell gene expression and function (Takamoto et al. 2014, Shao et al. 2015).

Utilizing RNAseq in human pancreatic islets and the rodent insulinoma cell line, Zhou et al. suggested recently that the LIM homeodomain protein ISL1 is among the primary downstream targets of TCF7L2 and the existence of the TCF7L2–ISL1 transcriptional network (Zhou et al. 2014).

In rodent species, ISL1 was shown to be expressed in pancreatic α, β, δ and PP cells (Karlsson et al. 1990, Dong et al. 1991, Ahlgren et al. 1997). A previous study by Du et al. revealed that Isl1-deficient pancreatic endocrine precursors cannot form functional islet cells, and that β-cell specific Isl1−/− mice were diabetic (Du et al. 2009). Several follow-up investigations have then revealed the role of ISL1 in regulating the expression of pancreatic islet genes including MafA, Arx, Pdx1 and Slc2a2 (Glut2) (Liu et al. 2011, Hunter et al. 2013, Ediger et al. 2014), and that ISL1 may cooperate with LIM domain binding 1 (LDB1) in exerting its regulatory functions in pancreatic islets (Hunter et al. 2013, Ediger et al. 2016), including the maintenance of the terminally differentiated state of pancreatic β-cells (Ediger et al. 2016).

To determine whether ISL1 also mediates the function of the Wnt signaling pathway effector β-catenin (β-cat)/TCF and GLP-1 in rodent pancreatic β-cells, here we assessed the effect of TCF7L2DN expression on Isl1 mRNA and ISL1 protein levels in our βTCFDN mouse model (Shao et al. 2015), and in the rat insulinoma cell line INS-1 832/13. We have also compared the effect of adenovirus-mediated TCF7L2DN expression and siRNA-mediated Isl1 knockdown on β-cell gene expression, as well as high glucose (HG) and HG plus GLP-1 stimulated insulin secretion. Our observations have collectively confirmed the existence of the TCF7L2–ISL1 transcriptional network in rodent islets. More importantly, our investigations have expanded this novel network study into the function of the incretin hormone GLP-1, on which a series of diabetes drugs have been generated during the past decade (Shigeto et al. 2015, Wu et al. 2015, Tian & Jin 2016).

Materials and methods

Cell cultures and experimental mice

The rat insulinoma cell line INS-1 832/13, the mouse MIN6 cell line and mouse pancreatic islets were cultured as previously described (Shao et al. 2010). No mycoplasma contamination has been verified in these two cell lines. Mouse islets were isolated by a collagenase-based method from 6- to 8-week-old male C57BL6/J mice (The Jackson Laboratory) as described previously (Shao et al. 2013b). The generation of the transgenic mouse line βTCFDN has been presented previously, via mating TCF7L2DNfet with Ins2-rtTA (Shao et al. 2015). The pregnant mothers were fed with doxycycline diet provided by Harlan (625 mg/kg doxycycline, TD.08434) until weaning. After weaning and genotype clarification at age of 3 weeks, both βTCFDN and the wild-type littermate controls were fed with the doxycycline diet before the experiments were performed (Shao et al. 2015). Male βTCFDN mice and littermate wild-type controls at the age of 6–8 weeks were utilized for pancreatic islets isolation. Mice were maintained in pathogen-free cages at room temperature and relative humidity of 50%, with free access to food and water under a 12h light/darkness cycle. Up to five mice were kept per cage. All animal protocols were approved by the Institutional Animal Care and Use Committee of the University of Health Network.

Reagents and plasmid

Recombinant human Wnt-3a was the product of R&D Systems (5036-WN). The Wnt signaling pathway inhibitor iCRT14 was purchased from Sigma-Aldrich. GLP-1(7–37) is the product of Abcam. Small interfering RNAs (siRNA) recognizing either a scrambled sequence or Isl1 were provided by Ambion Thermo Fisher Scientific. pGL3-Isl1 Luc plasmid was kindly provided by Dr Chunyan Zhou (Department of Biochemistry and Molecular Biology, School of Basic Medical Sciences, Peking University, China) (Lu et al. 2014).

Western blotting and immunohistochemistry (IHC) studies

Methods for Western blotting and IHC have been described previously (Shao et al. 2010, Ip et al. 2015). Antibodies
utilized in this study are listed in Table 1. Among them, the TCF7L2 antibody was shown to recognize the long and short isoforms (78 kDa and 58 kDa, respectively) of both human and mouse TCF7L2 (Shao et al. 2015). For Western blotting, a 1:1000 dilution solution was made for each of the antibodies. For IHC, a 1:10,000 dilution solution was prepared for the Isl1 antibody.

Quantitative RT-PCR (real-time RT-PCR or qRT-PCR)

RNAs were extracted from the INS-1 832/13 cell line or isolated mouse pancreatic islets, following the indicated treatment, with the use of the TRI Reagent (Sigma-Aldrich). Complementary DNA (cDNA) was synthesized for each of the antibodies. For IHC, a 1:10,000 dilution solution was made for each of the antibodies. For IHC, a 1:10,000 dilution solution was prepared for the Isl1 antibody.

Luciferase reporter assay

Transfection of luciferase (LUC) reporter plasmids into the mouse MIN6 cells was achieved using 2 µg of polyethylenimine (Sigma) and 1 µg LUC reporter gene construct in 12-well plates. Luciferase (LUC) reporter analyses were performed using firefly luciferin substrate.

Table 1 Antibodies utilized in the study.

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<th>Antibody</th>
<th>Company</th>
<th>Catalog number</th>
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<td>TCF7L2</td>
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<td>-1000</td>
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<td>HA</td>
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<td>MMS-101P-200</td>
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<td>GAPDH</td>
<td>Santa Cruz Biotechnology</td>
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Table 2 Primers utilized in the study.

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<tr>
<th>Gene (mouse/rat)</th>
<th>Forward</th>
<th>Reverse</th>
<th>Product size (bp)</th>
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<td>5-CCAAGGCTGAAGATTCCCG-3</td>
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<td>Insulin2 (rat)</td>
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<td>5-CTTGTTGGCTCTCCATTCG-3</td>
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<td>Insulin2 (mouse)</td>
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<td>5-GCTGTTGCGAGACTGATGCT-3</td>
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<tr>
<td>Axin2</td>
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<td>5-AACCTGCTGACCAAACAAT-3</td>
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<td>Glp1r</td>
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<td>5-TCACATTTTTCATCTC-3</td>
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<td>Nkx6.1 (rat)</td>
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<td>Slc2a2</td>
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<td>TCF7L2 (rat)</td>
<td>5-GAGAAGAGCAAGCAGCATCGAC-3</td>
<td>5-GTGACGAGGAGGAGGAGGAGG-3</td>
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<td>C-Myc</td>
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<td>5-TGACAGGCGAGAAGACACC-3</td>
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<td>Cyclin D1</td>
<td>5-ATGGAAGAATCTCATTCCA-3</td>
<td>5-ATGGAAGAATCTCATTCCA-3</td>
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(BioShop, Burlington, ON, Canada) as described previously (Ip et al. 2015).

**Chromatin immunoprecipitation (ChIP)**

The ChIP procedure has been described in our previous study (Ip et al. 2015). Following ChIP, PCR was performed using primers recognizing the *Isl1* promoter surrounding the TCF binding site, as provided in Table 2.

**Insulin secretion assay**

For the INS-1 832/13 cell line, we followed the method by Deisl et al. for measuring insulin secretion (Deisl et al. 2013). Briefly, INS1 832/13 cells were subject to siRNA-mediated *Isl1* knockdown for 48h. After an equilibration period of 2h in the KRB with 2.8mM glucose, cells were incubated for 60min in fresh KRB buffer containing 2.8mM glucose (defined as low glucose, LG), 16.7mM glucose (defined as HG), or 16.7mM glucose plus 10nM GLP-1. Supernatants were then harvested for insulin detection with the insulin enzyme immunoassay kit (AIS, Hong Kong) (Shao et al. 2015). For the MIN6 cell line, cells were incubated in KRB with 2.0mM glucose for 6h, followed by 30min incubation with 2.8mM glucose (LG), 25mM glucose (HG) or HG plus 10nM GLP-1. For cellular insulin content determination, cells from each of the treatment groups were harvested, followed by insulin content detection with the above kit. Values for cellular insulin content were normalized against cellular protein content. Insulin secretion in each group of cells was presented as percentage of cellular insulin content in 60min (for INS-1 832/13) or 30min (for MIN6).

**Calcium imaging**

Fura-2 ratiometric Ca$$^{2+}$$ sensitive dye was used to measure intracellular Ca$$^{2+}$$ levels in single cells, as described previously (Gardzinski et al. 2007). INS-1 832/13 cells were pre-loaded with Fura-2 AM (5μM, Molecular Probes) in dark for 40min. Fura-2 Ca$$^{2+}$$ signal was acquired at alternate excitation wavelengths of 340 and 380nm, while cells were perfused with the control/loading solution containing 120mM NaCl, 3.5mM KCl, 1mM MgCl, 0.4mM KHPO$$_{4}$$, 5mM NaHCO$$_{3}$$ and 1.3mM CaCl$$_{2}$$ (pH: 7.3–7.4), adjusted to 2.8mM glucose, 16.7mM glucose or 16.7mM glucose with or without 10nM GLP-1. Signals were digitized by an intensified charged-coupled device camera (PTI), and fluorescence intensity ratios were calculated using EasyRatioPro. Response to stimulus was quantified by extracting and evaluating the change in F/F$$_{0}$$ before and after each stimulus. Results are normalized to the response to 2.8mM glucose (Gardzinski et al. 2007, Soltani et al. 2011).

**Statistics**

All results are expressed as mean ± S.D. Statistical significance was assessed by the Student’s t-test or by one-way ANOVA. P value less than 0.05 was considered to be statistically significant.

**Results**

**TCF7L2DN-mediated β-cat/TCF functional knockdown leads to reduced Isl1 expression**

We reported previously that in βTCFDN mouse islets, PDX1- and NKX6.1-positive pancreatic islet cell numbers were reduced (Shao et al. 2015). With both qRT-PCR and Western blotting approaches, we show here that in βTCFDN mouse islets, both *Isl1* mRNA and ISL1 protein levels were reduced (Fig. 1A and B). βTCFDN mice also showed approximately 10% reduction of ISL1-positive pancreatic islet cells, assessed by the immunohistochemistry approach (Supplementary Fig. 1, see section on supplementary data given at the end of this article).

As ISL1 is known to be expressed in other pancreatic islet cells, we cannot attribute the observed reduction fully to pancreatic β-cells. Further investigations were then conducted in the clonal insulinoma cell line INS-1 832/13 with adenovirus-mediated TCF7L2DN expression. For this purpose, INS-1 832/13 cells were infected with adenovirus that expresses wild-type TCF7L2 or TCF7L2DN, generated in our previous study (Ip et al. 2015) (Fig. 1C), followed by Western blotting and qRT-PCR. As shown in Fig. 1D and E, Ad-TCF7L2DN infection reduced ISL1 expression at both protein and mRNA levels. Ad-TFC7L2WT infection, however, increased *Isl1* mRNA levels (Fig. 1E) although we did not see appreciable activation on ISL1 protein expression (Fig. 1D). Thus, similar to the INS-1 832/13 cell Tcf7l2 depletion model presented by Zhou et al. (Zhou et al. 2014), *in vivo or in vitro* TCF7L2DN expression also reduces ISL1 expression in rodent pancreatic β-cells.

**ISL1 is likely a direct downstream target of β-cat/TCF7L2**

Mouse Isl1–LUC reporter gene plasmid (Lu et al. 2014) was then utilized to test whether ISL1 promoter activity
ISL1 mediates function of TCF7L2

Shao et al. (2013) showed that ISL1–LUC activity can be stimulated by Wnt3a or lithium chloride (LiCl) treatment. iCRT14, a chemical inhibitor of the β-caten/TCF signaling (Marlow et al. 2013), repressed ISL1–LUC and blocked Wnt3a or LiCl induced activation (Fig. 2A). The co-transfection of the constitutively active S33Y β-caten mutant (Ni et al. 2003) generated a robust activation on Isl1–LUC (Fig. 2B), while Ad-TCF7L2DN infection repressed Isl1–LUC expression (Fig. 2C).

An evolutionarily conserved TCF binding motif was located within the Isl1 promoter region, designated as TBE1 (Fig. 2D) (Lu et al. 2014). Via the utilization of the ChIP approach in the INS-1 832/13 cell line, we found that TCF7L2 antibody, but not the control IgG, precipitated DNA fragment that contains TBE1 (Supplementary Fig. 2). We define this as the indication of the interaction between TCF7L2 and TBE1. Figure 2E/F shows our qChIP results, indicating that the interaction between TCF7L2 and TBE1 was stimulated by Wnt3a treatment or exogenous TCF7L2 expression but repressed by TCF7L2DN expression. These observations collectively suggest that ISL1 is likely a directly downstream target of TCF7L2 in mouse pancreatic β-cells.

Unexpectedly, Wnt3a treatment generated no appreciable effect on ISL1 protein expression in the INS-1 832/13 cell line (Fig. 2G). We repeated the experiment with 16h Wnt3a treatment in INS-1 832/13 cells and performed the same Wnt3a treatment in mouse islets. Again we did not see the stimulation on ISL1 protein level (Supplementary Fig. 3). We then tested effects of β-caten/TCF activity inhibition with iCRT14 and stimulation with S33Y β-caten expression in the INS-1 832/13 cell line. As shown in Fig. 2H and I, iCRT14 treatment repressed, while S33Y β-caten expression stimulated ISL1 protein expression, supporting our suggestion that β-caten/TCF positively regulates β-cell ISL1 expression.

Figure 1
TCF7L2DN-mediated β-caten/TCF functional knockdown leads to reduced ISL1 expression. (A and B) Comparison of Isl1 mRNA and ISL1 protein expression in β-TCFDN (Tg) and littermate control (WT) mouse islets. (C) Schematic of constructs of human Ad-TCF7L2DN (TCFDN) and Ad-TCF7L2 (TCF). The β-catenin interaction domain and the DNA binding HMG domain are illustrated. (D and E) Detection of ISL1 protein and Isl1 mRNA in INS-1 832/13 cells infected with Ad-TCF7L2DN or Ad-TCF7L2. HA antibody detects adenovirus expressed exogenous TCF7L2 or TCF7L2DN. n = 3 for panels A, B and E. Panel D is a representative blot for 3 independent Western blotting experiments. * or #, P<0.05. Values are represented as mean ± s.e.m. A full color version of this figure is available at https://doi.org/10.1530/JME-17-0181.

TCF7L2DN expression and Isl1 knockdown generate similar effect on β-cell gene expression

We found previously that Ad-TCF7L2DN infection in the INS-1 832/13 cell line repressed the expression of Pdx1, Nkx6.1 and other β-cell genes (Shao et al. 2015). Here, we conducted the adenovirus infection experiments in both the INS-1 832/13 cell line and mouse pancreatic islets, and expanded the qRT-PCR examination to include endogenous Tcf7l2, Tcf7, genes that encode GLP-1 receptor (Glp1r), GIP receptor (Gipr) and Glut2. The virus infection in mouse pancreatic islets was monitored by visualizing GFP 48h after the adenovirus infection and HA-tagged protein detection with Western blotting (Supplementary Fig. 4). Figure 3A and B shows that in both INS-1 832/13 cell line and mouse islets, TCF7L2DN expression did not reduce endogenous Tcf7l2 or Tcf7 expression.
The expression of a battery of β-cell genes including Glp1r, Gipr and Glut2 were repressed by Ad-TCF7L2DN expression. siRNA-mediated Isl1 knockdown was then performed in the INS-1 832/13 cell line. Figure 3C shows that expression of tested β-cell genes, including Glp1r, Gipr and Glut2 was also repressed by Isl1 knockdown. The similar repression pattern on β-cell gene expression via Ad-TCF7L2DN infection and Isl1 knockdown allows us to suggest that SL1 mediates the function of β-cat/TCF7L2 on β-cell gene expression. As anticipated, Isl1 knockdown did not affect the expression of Tcf7l2 or Tcf7 (Fig. 3C), as Isl1 is likely downstream of the bipartite transcription factor β-cat/TCF.

To further verify that ISL1 mediates the effect of TCF7L2 on Glp1r expression, we tested the stimulation on Glp1r expression by Ad-TCF7L2 infection in the INS-1 832/13 cell line (Fig. 4A) and observed that the stimulation can be blocked by simultaneous Isl1 gene knockdown (Fig. 4B).

GLP1 stimulated β-cat S675 phosphorylation and insulin secretion can be attenuated by Isl1 knockdown

The repressive effect on Glp1r level by either TCF7L2DN expression or Isl1 knockdown prompted us to ask whether ISL1 mediates β-cell function of the incretin hormone GLP1. In pancreatic β-cells, β-cat/TCF activity can be stimulated by GLP1 as this incretin hormone stimulates β-cell S675 phosphorylation (Liu & Habener 2008, Shao et al. 2013a). We repeated the observation by Liu et al. (Liu & Habener 2008) that GLP1 (10 nM) can stimulate β-cat S675 phosphorylation in the INS-1 832/13 cell line, and demonstrated that this stimulation can be blocked with siRNA-mediated Isl1 knockdown (Fig. 5A). Figure 5B and C shows that in both the INS-1 832/13 cell line and mouse pancreatic islets, adenovirus-mediated TCF7L2DN expression also attenuated GLP1-stimulated β-cat S675 phosphorylation.
We then assessed the effect of Isl1 knockdown on glucose and GLP1 regulated insulin secretion. In the INS-1 832/13 cell line with our insulin secretion assay protocol, HG as well as HG plus GLP1- (10nM) stimulated insulin secretion were recognizable (Fig. 5D), but were not as profound as we observed previously in mouse islets (Shao et al. 2015). Nevertheless, following siRNA-mediated Isl1 knockdown, it appeared that HG plus GLP1-facilitated insulin secretion was attenuated, when compared with cells transfected with the scramble siRNA (Fig. 5D/E).

We have also tested whether Isl1 knockdown reduces calcium influx in the INS-1 832/13 cell line in response to HG and GLP1 treatment, with the ratiometric Fura-2 Ca^{2+} imaging assays. In INS-1 832/13 cells treated with scrambled siRNA, HG or HG plus GLP1-1 enhanced calcium influx as compared to low glucose treatment (Fig. 5F and G, n = 16). In this cell line treated with Isl1 siRNA, HG or HG plus GLP1-1 enhanced calcium influx was attenuated (Fig. 5F and G, n = 16). Representative traces of INS-1 cells treated with scrambled siRNA and Isl1 siRNA are shown in Fig. 5F. KCl (50mM for 15s) was then applied to cells at the end of the experiments to confirm that cells are viable.

As the stimulatory effect of HG on insulin secretion in the INS-1 832/13 cell line was relatively moderate, we tested the effect of Isl1 knockdown in the MIN6 cell line. The knockdown efficiency on Isl1 protein expression in this cell line was presented in Fig. 6A. Figure 6B/C shows that after Isl1 knockdown, HG (25mM) or HG plus GLP1-stimulated insulin secretion was attenuated.

Figure 3
Isl1 knockdown mimics the effect of TCF7L2DN expression on β-cell gene expression. (A) INS-1 832/13 cells were infected for 48h with either the control adenovirus (GFP) or AdTCF7L2DN (TCFDN). (B) Mouse islets were infected for 48h with either the control adenovirus (GFP) or AdTCF7L2DN (TCFDN). (C) siRNA-mediated Isl1 knockdown was performed in the INS-1 832/13 cells. n = 3 for panels A, B and C. *P < 0.05; **P < 0.01. Values are represented as mean ± s.d. A full color version of this figure is available at https://doi.org/10.1530/JME-17-0181.

Figure 4
Isl1 mediates the stimulation on Gp1r expression by TCF7L2 over-expression. (A) Ad-TCF7L2 (Ad-TCFWT) infection in the INS-1 832/13 cell line increased Isl1 mRNA expression, accompanied with increased Glp1r mRNA level. The right panel shows the detection of TCF7L2/TCFWT (the 78kDa long isoform) by Western blotting 48h after indicated virus infection with the HA tag antibody or the TCF7L2 antibody. (B) In INS-1 832/13 cells, simultaneous Isl1 knockdown (with siRNA against Isl1) attenuated the stimulation of TCF7L2 over-expression (Ad-TCFWT infection) on Gp1r mRNA level. The right panel shows the verification of siRNA-mediated Isl1 knockdown by Western blotting and the detection of adenovirus infection mediated TCF7L2 expression by RT-PCR. ** or ****; P < 0.01. n = 3 for Panels A and B. A full color version of this figure is available at https://doi.org/10.1530/JME-17-0181.
ISL1 mediates function of HG/GLP-1. When the HG was utilized to drive the expression of the mouse TCF7L2DN, a TCF member (Shao et al. 2015), the avoidance of the compensation by different TCF members and the potential bi-directional functional feature of ISL1/GAPDH (protein) expression in pancreatic β-cell genes (Savic et al. 2013) was observed. Since 2006, reproducible GWAS have shown that TCF7L2 is a T2D risk gene (Florez et al. 2006, Grant et al. 2006, Cauchi et al. 2007). To explore metabolic functions of this Wnt pathway effector, efforts have been made in assessing its role in the liver, adipose tissues and pancreatic islets with various tools (Lyssenko et al. 2007, Schafer et al. 2007, Norton et al. 2011, Boj et al. 2012, Savic et al. 2013, Jin 2016).

As discussed previously, different functional outcomes were reported on assessing mouse models with β-cell Tcf7l2 deletion (Boj et al. 2012, da Silva Xavier et al. 2012, Mitchell et al. 2015, Shao et al. 2015). To resolve the potential discrepancy, we and another team employed a functional knockdown approach via TCF7L2DN expression in pancreatic β-cells (Takamoto et al. 2014, Shao et al. 2015). The advantages with TCF7L2DN include the avoid of the compensation by different TCF members and the potential bi-directional functional feature of a TCF member (Jin 2016). When the RIP promoter was utilized to drive the expression of the mouse TCF7L2DN, the generated transgenic mouse lines showed impaired glucose tolerance and decreased insulin secretion, associated with a marked reduction of β-cell mass and decreased expression of a battery of β-cell genes (Takamoto et al. 2014). We generated βTCFDN in which TCF7L2DN

Figure 5
Isil1 knockdown attenuates the effect of GLP1 on β-cat S675 phosphorylation as well as GLP1 stimulated insulin secretion. (A) Effect of siRNA-mediated Isil1 knockdown on GLP-1 (10 nM) stimulated β-cat S675 phosphorylation in the INS-1 832/13 cell line. (B) and (C) Effect of Ad-TCF7L2DN (Ad-TCFDN) infection on GLP1 (10 nM) stimulated β-cat S675 phosphorylation in the INS-1 832/13 cell line (B) and in mouse islets (C). A, B and C show representative blot of three independent experiments. (D/E) Assessment of the effect of Isil1 knockdown on insulin secretion (LG vs HG) or HG plus GLP1 (10nM) induced insulin secretion in the INS-1 832/13 cell line. Panel E shows the insulin contents in cells received control (Scramble) or Isil1 siRNA transfection. (F and G) Average change in 340/380 ratio values to HG, and HG plus GLP-1 induced calcium influx, normalized to LG response in INS-1 cells treated with scrambled siRNA or Isil1 siRNA. n=5 for Panel D. n=16 for Panel G. * or t, P<0.05; ** or t, P<0.01. * or **, GLP1 treatment vs control for Panels A, B and C, or HG vs LG for Panels D and F; * or t*, HG plus GLP-1 vs HG, for Panels D and G. A full color version of this figure is available at https://doi.org/10.1530/JME-17-0181.

Discussion
Since 2006, reproducible GWAS have shown that TCF7L2 is a T2D risk gene (Florez et al. 2006, Grant et al. 2006, Cauchi et al. 2007). To explore metabolic functions of this Wnt pathway effector, efforts have been made in assessing its role in the liver, adipose tissues and pancreatic islets with various tools (Lyssenko et al. 2007, Schafer et al. 2007, Norton et al. 2011, Boj et al. 2012, Savic et al. 2013, Jin 2016).

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Figure 6
Isil1 knockdown attenuates the effect of HG plus GLP1 on insulin secretion in the MIN6 cell line. (A) The effect of Isil1 knockdown on ISL1 protein expression, assessed by Western blotting. (B/C) Assessment of Isil1 knockdown on HG (25 mM) and HG plus GLP1 (HG/GLP1, 10nM) on insulin secretion in the MIN6 cell line. n=3 for Panel A and n=5 for Panel B/C. For Panel B, **, HG vs LG; **, HG plus GLP1 vs HG. A full color version of this figure is available at https://doi.org/10.1530/JME-17-0181.
expression was driven by the Ins2 promoter, inducible upon doxycycline feeding. βTCFDN shows impaired glucose tolerance and reduced β-cell mass in adulthood when TCF7L2DN expression was induced during the embryonic developmental stage by doxycycline feeding starting in their pregnant mothers (Shao et al. 2015). Attenuated response to GLP1-stimulated insulin secretion was also shown in βTCFDN islets, associated with reduced expression of a battery of β-cell genes, including Pdx1, Nkx6.1, Mafa, Ins1, Ins2, Ngn3, Glp1r and Gipr (Shao et al. 2015).

Here, we expanded the investigation by including the LIM homeobox gene Isl1, a β-cat/TCF downstream target identified previously in other cell lineages (Lin et al. 2007, Lu et al. 2014) and suggested very recently by Zhou et al. with RNAseq in human islets (Zhou et al. 2014). We show here the repressive effect of TCF7L2DN on Isl1 in both the βTCFDN mouse islets and in the INS-1 832/13 cell line.

Our examinations with the LUC reporter assay and qChIP collectively suggest that Isl1 is likely a direct downstream target of β-cat/TCF7L2 in mouse pancreatic β-cells, involving the binding of β-cat/TCF7L2 on TBE1, a conserved cis-element within the human ISL1 and mouse Isl1 gene promoters. The stimulation on Isl1 promoter activity was observed with the Wnt ligand Wnt3a, the GSK-3 inhibitor LiCl, and the constitutively active S33Y β-cat mutant. Wnt3a treatment, however, generated no stimulation on mouse ISL1 at protein level. This could be due to the off-target effect of the recombinant Wnt3a as a chemical reagent, or the existence of yet-to-be-identified negative feedback loop on ISL1 expression at the protein level. Indeed, we are far away from clear on the overlapping, redundant or even opposite effect of 19 Wnt ligands on a given target gene in a given tissue, or an organ, or a cell lineage. As ISL1 protein expression was repressed by iCRT14 and stimulated by S33Y β-cat, we suggest that it is a direct downstream target of β-cat/TCF7L2, the key canonical Wnt signaling pathway effector.

We suggest that ISL1 serves as the key mediator for β-cat/TCF7L2 in regulating β-cell gene expression as AD-TCF7L2DN infection and siRNA-mediated Isl1 knockdown generated similar repressive effect on β-cell gene expression. More importantly, we revealed that ISL1 knockdown attenuated the effect of the incretin hormone GLP1 in pancreatic β-cells. As illustrated in Fig. 7, ISL1 is a downstream target of β-cat/TCF7L2 and its expression can also be positively regulated by GLP1 via stimulating β-cat S675 phosphorylation. In addition, GLP1R expression can be positively regulated by β-cat/TCF7L2, likely using ISL1 as a mediator, although the molecular mechanism underlying this regulation is yet to be determined. Whether this signaling cascade also plays a role for GLP1 to exert its other pancreatic β-cell functions, such as the stimulation of β-cell survival signaling pathway, as well as the exertion of PKA, Epac and other signaling cascade-mediated insulin secretion, remains to be further investigated (Brubaker & Drucker 2004, Holz & Chepurny 2005, Kolic & MacDonald 2015, Shigeto et al. 2015, Wu et al. 2015, Takeda et al. 2016).

ISL1 was initially identified as a Wnt pathway downstream target in cardiac cells. The loss of Wnt/β-cat signaling in anterior heart field resulted in defective outflow tract and frayed ventricle development, associated with the decrease in ISL1 positive progenitor cells, while Wnt gain of function led to the expansion of ISL1-positive progenitor cells (Cohen et al. 2007). Transcriptional stimulation of ISL1 expression by Wnt activation was then demonstrated with the cellular models including the embryonic carcinoma cell line P19 (Lin et al. 2007, Lu et al. 2014). ISL1 was also claimed as a direct downstream target of forkhead transcription factors in second-heart-field-derived mesoderm (Kang et al. 2009). Although ISL1 was reported as the first insulin gene enhancer (Karlsson et al. 1990), scientists paid less attention to its β-cell function when comparing with that of PDX1 and NKX6.1, possibly for its expression in other pancreatic endocrine cell lineages as well (Dong et al. 1991, Du et al. 2009, Liu et al. 2011, Hunter et al. 2013, Ediger et al. 2014, 2016). Considering the recognition of ISL1 in regulating islet gene expression (Liu et al. 2011, Hunter et al. 2013, Ediger et al. 2014), the TCF7L2/ISL1 transcriptional network (Zhou et al. 2014), and our current study showing its role
in mediating the function of GLP1, we suggest that more attention needs to be paid in the future for this islet cell transcription factor.

It is worth to mention that a recent study shows that β-cells from Gipr−/− mice display greater sensitivity to apoptosis and markedly lower islet expression of Tcf7 (Campbell et al. 2016). We observed reduced Gipr expression but not Tcf7 expression after TCF7L2DN expression or siRNA-mediated Isl1 knockdown. Further investigations are needed to clarify the relationship among the 3 TCF members (Columbus et al. 2010),Isl1, as well as 2 incretin receptors in mediating the function of the two incretin hormones in pancreatic β-cells. We, however, cannot properly detect pancreatic TCF7 or TCF7L1 expression with reliable antibodies by Western blotting (Columbus et al. 2010).

In summary, we presented in the current study that Isl1 is a direct downstream target of β-cat/TCF in pancreatic β-cells, and it is among the mediators of human and mouse TCF7L2 in exerting its metabolic functions including β-cell gene expression. Our investigation has not only verified the existence of TCF7L2/ Isl1 transcriptional network in rodent islet β-cells, but also expanded the implication of this network into the function of the incretin hormone GLP-1.

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