Novel mitochondrial complex I inhibitors restore glucose-handling abilities of high-fat fed mice

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Abstract

Metformin is the main drug of choice for treating type 2 diabetes, yet the therapeutic regimens and side effects of the compound are all undesirable and can lead to reduced compliance. The aim of this study was to elucidate the mechanism of action of two novel compounds which improved glucose handling and weight gain in mice on a high-fat diet. Wildtype C57Bl/6 male mice were fed on a high-fat diet and treated with novel, anti-diabetic compounds. Both compounds restored the glucose handling ability of these mice. At a cellular level, these compounds achieve this by inhibiting complex I activity in mitochondria, leading to AMP-activated protein kinase activation and subsequent increased glucose uptake by the cells, as measured in the mouse C2C12 muscle cell line. Based on the inhibition of NADH dehydrogenase (IC₅₀ 27 µmol L⁻¹), one of these compounds is close to a thousand fold more potent than metformin. There are no indications of off target effects. The compounds have the potential to have a greater anti-diabetic effect at a lower dose than metformin and may represent a new anti-diabetic compound class. The mechanism of action appears not to be as an insulin sensitizer but rather as an insulin substitute.

Introduction

Whether administered in the early stages of the development of type 2 diabetes or as a combination therapy with injectables, metformin is the main drug of choice for treating the condition. Yet the therapeutic regimens, the side effects of the compound, and the probable off-target actions are all undesirable and can lead to reduced compliance (Donnan et al. 2002, Florez et al. 2010, Kirpichnikov et al. 2002). The principal mode of action of metformin responsible for its anti-diabetic effects centers around its role in inhibiting liver gluconeogenesis (Bailey & Turner 1996, Madiraju et al. 2014), antagonising glucagon action (Miller et al. 2013), and activating AMP-activated protein kinase (AMPK) (Li et al. 2015, Owen et al. 2000). This is achieved via either a reduction in the activity of complex I of the mitochondrial respiratory chain (Bridges et al. 2014,
Brunmair et al. 2004, El-Mir et al. 2000, Fontaine 2014, Owen et al. 2000) or the inhibition of mitochondrial glycerophosphate dehydrogenase (Madiraju et al. 2014). Other biguanides have similar and more potent effects on complex I (Bridges et al. 2014, Matsuzaki and Humphries 2015), with phenformin dramatically reducing the oxygen consumption rate in HepG2 cells (Bridges et al. 2014).

The effect of activating the intracellular metabolic sensor, AMPK, is to maintain ATP levels. This is achieved via further signaling resulting in an increase in glucose uptake by the cell (Pehmøller et al. 2009), an increase in fatty acid oxidation (Winder et al. 1997) and decrease in fatty acid synthesis (Munday et al. 1988), and a decrease in glycogen synthesis (Jørgensen et al. 2004) and an increase in glycolysis (Marsin et al. 2002). AMPK is activated during exercise as the cell seeks to replenish levels of ATP (Winder & Hardie 1996). Lowering high blood glucose levels in this way would appear to be as close to a natural physiological phenomenon as any other anti-diabetic mechanism of action.

This study deals with two related, novel compounds (PCT/EP2012/071286; see Fig. 1 and Supplementary Materials and methods (see section on supplementary data given at the end of this article)) with the aim of elucidating their mechanism of action. As can be seen, their principal mode of action is to inhibit complex I, thereby very effectively stimulating AMPK and increasing glucose uptake by muscle cells, a major site of insulin action within the body. These compounds represent a possible new anti-diabetic class and a move away from the well-known biguanides. In these respects, the compounds activate signaling pathways that are influenced in a similar way by exercise (Winder & Hardie 1996), and presumably also by metformin. Analyses also indicate that these compounds have no effects on other anti-diabetic targets and present no accepted toxic indicators.

**Materials and methods**

**Animal studies**

All procedures were approved by the UK Home Office and local ethical committee of the University of Leeds, UK. In a prevention study, wildtype C57Bl/6 male mice (~20g; Charles River Laboratories) were housed (4 mice per cage) in a conventional animal facility with a 12h light:12h darkness cycle, given access to water and an obesogenic diet ad libitum (Diet F3282; 5450 kcal kg−1; Bio-serv, Flemington, NJ, USA). Each group comprised 8 mice, which were fed for 8 weeks with the high-fat diet (HFD) containing DMSO (vehicle control) or RTB-70 at 0.04% (w/w) relative to the diet (~1mg/mouse/day). In a second series of experiments, mice were fed normal chow or the HFD for 16 weeks. The mice then underwent a glucose tolerance test which indicated that those on the high-fat diet indeed had impaired glucose handling abilities (mean AUC values – normal chow 17.64 ± 3.7, high-fat diet 36.08 ± 2.9). This latter HFD group was then sub-divided (each group had 8 mice) with one group maintained on the diet and the other on the diet plus the addition of RTC-1 at 0.04% (w/w) relative to diet (~1mg/mouse/day), for a further 16 weeks. At the end of both feeding paradigms, glucose (GTT) and insulin tolerance tests (ITT) were carried out. Following a period of fasting (12h and 4h for GTT and ITT, respectively), repeated whole-blood sampling was carried out on conscious mice at 30-min intervals after an initial i.p. injection of either glucose (1 mg kg−1 body weight; GTT) or insulin (10 µU kg−1 body weight; ITT). Body weight was recorded throughout the protocol.

**Cell culture**

Mouse muscle cells (C2C12; passages 5–25; ECACC (STR authenticated)) were differentiated in DMEM supplemented with 2% (v/v) horse serum for 3 days. RTC-1 and RTB-70 were dissolved in 100% DMSO (to 50 µmol L−1) and diluted to 1 µmol L−1 in Krebs Ringers Buffer (KRB; in mmol L−1: 136 NaCl, 20 HEPES, 4.7 KCl, 1 MgSO4, 1 CaCl2, 4.05 Na2HPO4, 0.95 NaH2PO4; pH 7.4) including glucose (5.5 µmol L−1). The cells were treated with RTC-1 or RTB-70 at 10 µmol L−1 in DMEM plus 2% (v/v) horse serum overnight or, as indicated, at 37°C. In other experiments, C2C12 cells were treated with Insulin (100 µmol L−1; 30min, Sigma), cytochalasin B (10 µmol L−1; 30min, Sigma, Flemington, NJ, USA), or metformin (500 µmol L−1; 6h, Sigma). In a further experiment,
Compound C (10 µmol L⁻¹; GE Healthcare) was added to the cells 1h before the addition of RTC-1 (10 µmol L⁻¹ for 6h). CHO cells (passages 15–25; ECACC) and primary human red blood cells were also treated with RTC-1 (10 µmol L⁻¹; 16h) before the addition of ³H deoxy-2-glucose.

**Glucose uptake**

Glucose uptake was monitored using a tritiated version of deoxy-2-glucose, a derivative of glucose that cannot be metabolized (Yun et al. 2009). Following treatment, the cells were washed in KRB and ³H deoxy-2-glucose (1 µCi mL⁻¹; specific activity 8 mCi mmol⁻¹; PerkinElmer) added for 10min at 37°C. The cells were washed 3 times in ice-cold KRB, solubilized in 0.1% (w/v) SDS for 30min, and 500 µl (~250 µg protein) of the extract added to 2 mL scintillation fluid (Ultima Gold, PerkinElmer). The results are expressed as counts per minute per mg protein (c.p.m. mg⁻¹; Wallac MicroBeta, PerkinElmer) as assayed by the BCA method (Smith et al. 1985).

**SDS–PAGE and Western immunoblotting**

Following drug treatments, the cells were washed in PBS and a lysis buffer (in mmol L⁻¹: 50 HEPES (pH 7.5), 150 NaCl, 10 Na₂HPO₄, 50 NaF, 1 EDTA, 1.5 MgCl₂, 2 Na₃VO₄, 1 Na₄P₂O₇, 1 PMSF, 1X SigmaFAST protease inhibitor cocktail, 10% (v/v) glycerol, and 1% (v/v) Triton X-100) was added. Following incubation for 1h, the mixture was centrifuged at 12,000g and 30 µL supernatant were loaded on 12% SDS–PAGE mini gels. The proteins were transferred to activated PVDF membranes and probed with anti-phospho (Thr172) AMPK alpha (diluted 1/1000; Cell Signalling) for 2h at RT. Secondary antibody (anti-rabbit; 1/5000, Dako) was added for 1h at RT. The labeled bands were detected by enhanced chemiluminescence.

**PPARγ lance assay**

The LanthaScreen TR-FRET PPARγ Co-activator Assay (Life Technologies) was performed according to the manufacturer’s protocol. In an agonist mode, the test compounds and agonist control GW1929 (1 µmol L⁻¹; Sigma) were incubated with fluorescein-labeled TRAP220/DRIP-2 coactivator peptide (125 nmol L⁻¹), terbium-labeled anti-GST antibody (5 nmol L⁻¹), and PPARγ LBD (20 nmol L⁻¹) for 1h at 25°C. The signals at 520 nm were normalized to those obtained from the terbium emission at 495 nm, and the 520nm/495 nm ratios were used as a measure for the TRAP220/DRIP-2 coactivator recruitment potential of the tested compounds. The ratio of the emission intensity of the acceptor (Fluorescein: λ = 520 nm) divided by the emission intensity of the donor (Tb: λ = 490 nm) was then calculated to determine the degree of nuclear receptor coactivator binding. Each measurement

![Figure 2](https://example.com/f2.png)

**Figure 2**

RTB-70 and RTC-1 restore normal glucose handling in high-fat fed mice. Mice fed a high-fat diet and RTB-70 (0.04% (w/w); ∆) for 8 weeks had improved glucose handling abilities compared with mice on high-fat diet alone (○) when assessed by GTT (A) and ITT (B). Mice fed a high-fat diet for 16 weeks and then a high-fat diet and RTC-1 (▲) for a further 16 weeks had improved glucose handling abilities compared with mice on high-fat diet alone (●) when assessed by GTT (D) and ITT (E), with levels mimicking control fed mice (●). Total body weights (×-RTB-70 and f-RTC-1) are also shown. Bar charts depict area under the curve (AUC) for GTT and ITT, and final body weights. Values are mean ± s.e.m.; n=8 per group; *p < 0.05, **p < 0.01; student’s t-test vs control.
was performed in duplicate. The 520 nm/495 nm ratio was plotted against the test compound concentration.

Homogeneous time-resolved fluorescence (GLP-1R)

The compounds were assayed for GLP-1 activation using a HTRF cAMP detection kit (Cisbio, Bedford, MA, USA). Briefly, 32,000 hGLP1R-expressing Flp-In HEK293 cells were plated in 25 μL/well of PBS with IBMX (Sigma) in 384-well solid bottom white plates and 15 μL/well compound in DMSO solution or controls were added. Following 30min incubation at RT, 25 μL/well of labeled d2 cAMP and 25 μL/well of anti-cAMP antibody (both diluted 1:20 in lysis buffer) were added to each well. Signals were recorded using the Omega Polarstar (BMG LabTech) with excitation at 330 nm and emissions at 615 nm and 660 nm.

Rat liver mitochondria isolation

Rat liver mitochondria were isolated by the method described by Chappell and Hansford (1972). Immediately after extraction, liver samples were washed free of excess blood, trimmed of any fat and connective tissue, finely chopped, and placed in 50 mL ice-cold (0–4°C) isolation medium (in mmol L\(^{-1}\): 250 sucrose, 5 Tris–HCl, 1 EGTA, pH 7.4). The tissue was homogenized and centrifuged at 750 \(g\) for 5 min at 4°C. The supernatant was filtered through a sieve before being centrifuged at 12,000 \(g\) for 10 min at 4°C. The resulting mitochondrial pellet was resuspended in ice-cold 25 mL isolation medium supplemented with 2% (w/v) defatted BSA and washed twice by centrifugation (12,000 \(g\) for 10 min at 4°C). The final mitochondrial pellet was resuspended in a small volume of isolation medium, and the aliquots were frozen at −20°C.

NADH: ubiquinone oxidoreductase activity analysis

Immediately before the assay, mitochondria were diluted in a hypotonic buffer (25 mmol L\(^{-1}\) K\(_2\)HPO\(_4\), 5 mmol L\(^{-1}\) MgCl\(_2\)) and permeabilized with three cycles of freeze thawing. The assay was carried at 30°C in a 1 mL cuvette (Shimadzu UV-2550, Milton Keynes, Buckinghamshire, UK). Permeabilized mitochondria were incubated with 50 mmol L\(^{-1}\) K\(_2\)HPO\(_4\) (pH 7.5), 3 mg mL\(^{-1}\) fatty acid-free BSA, 300 μmol L\(^{-1}\) KCN and 100 μmol L\(^{-1}\) NADH, and baseline activity was measured at 340 nm for 1 min. The reaction was initiated with the addition of 60 μmol L\(^{-1}\) ubiquinone and the resulting decrease in absorbance was measured for 3 min. Varying concentrations of RTC-1 or RTB-70 were then added and absorbance was measured for a further 3 min. The effect of the compounds on NADH:ubiquinone oxidoreductase was determined by comparing activity before and after addition, relative to the vehicle control, DMSO. Rotenone (1 μmol L\(^{-1}\)) was used as a positive control in this assay.

Oxygen consumption of rat liver mitochondria

Oxygen consumption rates were measured by an Oxygraph Respirometer (Oroboros, Innsbrück, Austria) as previously described (Breen et al. 2006). Mitochondria (6.2 μg mL\(^{-1}\)) were incubated at 37°C in a respiration
medium (5 mmol L\(^{-1}\) HEPES, 120 mmol L\(^{-1}\) KCl, 10 mmol L\(^{-1}\) K\(_2\)HPO\(_4\), 1 mmol L\(^{-1}\) EGTA; pH 7.4) to which 0.1% fatty acid-free bovine serum albumin was added on the day of use. Oxygen consumption rates were measured as the steady-state rates achieved on addition of glutamate (5 mmol L\(^{-1}\)) and malate (3 mmol L\(^{-1}\)). The sensitivity of these steady-state rates to various concentrations of RTB-70, RTC-1 or the DMSO vehicle control was then determined. Recovery of the steady state was then assessed by the addition of 10 mmol L\(^{-1}\) succinate (succinate-KOH, pH 7.4). The Oroboros Oxygraph Respirometer was calibrated according to the procedure of Reynafarje et al. (1985), assuming that 406 nmol of oxygen atoms was dissolved in 1 mL of ionic incubation medium at 37°C.

**ATP assay**

The Luminescent ATP Detection Assay (Abcam) was performed according to the manufacturer’s instructions. C2C12 cells (100 µL) were cultured in a black, clear bottom 96-well plate, and treated with RTC-1 (10 µmol L\(^{-1}\); 1 and 2 h) or rotenone (25 µmol L\(^{-1}\); 1 h). The cells were lysed by addition of detergent solution (50 µL) and incubated for 5 min. Reconstituted substrate buffer (containing d-Luciferin and Luciferase; 50 µL) was added to the cells and incubated for 15 min before luminescence was read. ATP concentration was determined via an ATP standard curve.

**Cell cytotoxicity assay**

Rat hepatocytes were used to test the cytotoxicity of RTC-1. The cells were plated at 1 × 10\(^6\) per well and treated with each compound for 4 h at 100 µg mL\(^{-1}\). At the end of the treatment, a Trypan blue exclusion test was carried out and cell viability was assessed using the Countess automated cell counter (Life Technologies). The experiments were repeated three times and carried out by Pharmidex (London, UK).

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**Figure 4**

RTC-1 induces glucose uptake in a specific, time-dependent manner. Dose dependency of RTC-1 (16 h) on glucose uptake in C2C12 cells, with an EC\(_{50}\) estimated at 11.42 µmol L\(^{-1}\) (dashed line; A). Glucose uptake following RTC-1 (10 µmol L\(^{-1}\); 16 h) treatment of CHO cells (B) and primary erythrocytes (C). RTC-1-induced (10 µmol L\(^{-1}\); 16 h) glucose transport inhibited by cytochalasin B (cyto B; 10 µmol L\(^{-1}\); 30 min; D) and co-addition of 5 mmol L\(^{-1}\) ‘cold’ glucose (E). The ability of metformin (500 µmol L\(^{-1}\); 16 h) to increase glucose uptake in C2C12 cells compared with RTC-1 (10 µmol L\(^{-1}\); 16 h; F). The values are mean ± s.e.m.; n = 4 per group; **p < 0.01, ***p < 0.001; student’s t-test vs basal.
Effects of RTC-1 on the hERG channel

Effects of RTC-1 on the hERG channel currents were determined by the two-electrode voltage clamp technique as described previously (Elliott et al. 2009). In brief, cRNA was prepared from the hERG cDNA (acc. No. GI:4557729) clone in the pSP64 vector using the SP6 Megascript Kit (Ambion). Stage V or VI oocytes were isolated from Xenopus toads, injected with 50 nL of cRNA (50 pg nL⁻¹), and incubated in ND-96 solution (in mmol L⁻¹: 96 NaCl, 2 KCl, 1.8 CaCl₂, 1 MgCl₂, 5 HEPES, 2.5 sodium pyruvate, pH 7.5) supplemented with 100 μmol L⁻¹ DTT solution at 18°C. After 2–4 days, currents were recorded in Ringer’s solution (in mmol L⁻¹: 115 NaCl, 2 KCl, 1.8 CaCl₂, 10 HEPES, pH 7.2) using microelectrodes made from borosilicate glass, filled with 3 M KCl, which had resistances between 0.5 and 2.0 M. The effects of compounds on tail currents at −50 mV were examined using a repeated pulse protocol. For this, control currents were first measured in Ringer’s solution during repeated depolarizing steps (+30 mV) delivered from a holding potential of −80 mV. The cells were then superfused with RTC-1 (10 μmol L⁻¹) and the current recordings continued until a steady-state effect was achieved. All experiments were repeated at least three times.

Compound synthesis

Details of compound synthesis for this study are given in the Supplementary Materials and methods (see section on supplementary data given at the end of this article).

Statistical analysis

The values are expressed as the mean ± s.e.m. Statistical comparisons are made using two-tailed student’s t-test compared with basal levels, unless otherwise stated.

Results

Effects on glucose handling

The initial compound (RTB-70) and one of its derivatives, RTC-1, were assessed for their ability to improve glucose
handling in mice subjected to a high-fat diet. In the case of RTB-70, the compound was added to the diet for 8 weeks from the start of the study. With RTC-1, the compound was administered for 16 weeks after a preceding 16 week period on the diet alone when insulin resistance became evident. Fig. 2A and B reveal that RTB-70 significantly improved glucose handling in both glucose and insulin tolerance tests. This was also true in the intervention study with RTC-1 (Fig. 2D and E). Comparisons with an untreated chow-fed group indicated that RTC-1 restored glucose handling to levels not significantly different to controls. Both compounds also reduced the weight gain exhibited by the control groups of animals (Fig. 2C and F). There were no significant changes in food intake between the control and treated groups. The mice exhibited no adverse reaction to the compounds and no hyperactivity was observed as well as no deaths occurred.

In order to ascertain the mechanism by which these compounds exert their anti-diabetic effects, their ability to activate well-known type 2 diabetes targets was assessed. Neither RTB-70 nor RTC-1 was agonist of GLP-1R (Fig. 3A) or PPARγ (Fig. 3B). The compounds were also tested for their effects on glucose uptake in a mouse muscle cell line, C2C12. Glucose uptake was assayed by using 3H-deoxy-2-glucose, a derivative of glucose which cannot be metabolized. RTC-1 induced a significant increase in glucose uptake compared with vehicle-treated cells (Fig. 3C), which was more pronounced than the normal response to the positive control, insulin (100 nmol L⁻¹; 30 min). Perhaps meaningfully, RTB-70 did not affect glucose uptake significantly. Furthermore, the derivatives of RTC-1 that passed the Eli Lilly ‘Open Innovation Drug Discovery’ novelty screen were assessed for their efficacy in their anti-diabetic screenings. These compounds did not induce the secretion of insulin or GLP-1 from INS-1e and NCI cell lines, respectively (data not shown).

RTC-1 induced a concentration-dependent increase in glucose uptake when incubated with muscle cells for 16 h, with an EC₅₀ calculated at ~11 µmol L⁻¹ (Fig. 4A). This action of RTC-1 is not restricted to muscle cells as treatment of CHO cells with RTC-1 (10 µmol L⁻¹; 16 h) also induced a significant increase in glucose uptake compared with vehicle-treated controls (Fig. 4B). RTC-1 is not exerting its effect by directly activating the glucose transporter, as no glucose uptake was stimulated in erythrocytes (Fig. 4C). However, the ability of cytochalasin B (10 µmol L⁻¹; an inhibitor of the glucose transporter) to prevent this compound-induced uptake (Fig. 4D) indicated that the uptake was facilitated by a glucose transporter. This conclusion was reinforced by the observation that the addition of excess ‘cold’ glucose (5 mmol L⁻¹) along with the 3H deoxy-2-glucose inhibited the amount of detectable tritium in the cells (Fig. 4E).

Mechanism of action
AMPK, being the major metabolic biosensor of the cell, was the logical protein to investigate. Phosphorylation of AMPK at threonine 172 is an indicator of AMPK activation. Using C2C12 cells, Western immunoblotting

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**Figure 6**

RTC-1 is not cytotoxic and does not activate hERG significantly. Assay of RTC-1 (100 µg/mL; 4 h) cytotoxicity as measured by Trypan Blue exclusion in rat hepatocytes compared with the known toxic agents Chlorpromazine and Chloroform (both 100 µg/mL; A). Representative peak of the effect of RTC-1 (10 µmol L⁻¹) on tail currents of hERG-expressing oocytes prepared from Xenopus toads, as determined by a repeated pulse protocol (insert: representative current traces before (black trace) and after (gray trace) the application of RTC-1; B). Normalized data (n = 3; C), before and after RTC-1 application. The values are mean ± s.e.m.; n = 3 per group; **p < 0.01, ***p < 0.001; student’s t-test vs control.
showed that RTC-1 (10 µmol L⁻¹) induced an increase in Thr172 phosphorylation of AMPK in a time-dependent manner, the first increase occurring at 45 min (Fig. 5A). In order to confirm the sequence of events through which glucose enters the cell, pre-treatment with an inhibitor of AMPK activation, Compound C (1 h; 10 µmol L⁻¹), before RTC-1 (10 µmol L⁻¹; 6 h) was seen to attenuate the RTC-1-induced increase in glucose uptake (Fig. 5B). Therefore indicating that AMPK is a major intermediate in the effect of these compounds.

The mode of action of these compounds could be a direct effect on the enzyme or through the normal biological mechanism, namely an increase in the AMP to ATP ratio. RTC-1 induced a decrease (not statistically different from basal levels) in ATP levels 1 h after addition to cells; there was no discernible difference observed at 2 h (Fig. 5C) and thereafter. Rotenone (25 µmol L⁻¹), a known complex I inhibitor, acted as a positive control. Interestingly, RTC-1 (IC₅₀ 27 µmol L⁻¹), and to a lesser extent RTB-70 (IC₅₀ 138 µmol L⁻¹), both inhibited oxygen consumption by mitochondria, (Fig. 5D and E, respectively). The addition of succinate restored the oxygen consuming ability of these mitochondria (Supplementary Fig. 1). These observations led to the identification of the target protein. Assays of NADH oxidation using purified disrupted mitochondria in the presence of ubiquinone indicated that RTC-1 and RTB-70 (Fig. 5F and G, respectively) inhibit the NADH dehydrogenase in complex I of the electron transport chain of mitochondria (IC₅₀ for RTC-1 is 27 µmol L⁻¹). The data illustrated in Fig. 4F indicate that metformin, which has previously been shown to inhibit complex I in muscle cells (Brunmair et al. 2004), has a similar effect on glucose uptake in C2C12 mouse muscle cells to that of RTC-1. The response is very modest compared with RTC-1, despite a 50-fold higher dose.

The cell toxicity profile of RTC-1 confirmed the lack of adverse reactions observed in the animal studies. RTC-1 did not induce cell death in rat hepatocytes following 4 h treatment at 100 µg mL⁻¹ compared with the known cytototoxic agents chlorpromazine and chloroform (Fig. 6A). The ability of druggable compounds to affect hERG, a voltage-gated potassium channel involved in cardiac action potential repolarization, is a recognized preclinical safety test (Bowlby et al. 2008, Priest et al. 2008). RTC-1 (10 µmol L⁻¹) did not significantly affect hERG tail current amplitudes as measured by voltage clamp in oocytes from Xenopus toads (Fig. 6B and C).

**Discussion**

The results above present a potential new class of anti-diabetic compounds which possess a targeted effect on the NADH dehydrogenase of complex I of the mitochondrial
respiratory chain, the result of which is to activate AMPK. This enzyme, in turn, activates the pathway by which the GLUT4 transporter is mobilized to the plasma membrane of most mammalian cells with the consequent increase in glucose uptake by the cell. In this respect, the mechanism has elements in common with physical exercise (Winder & Hardie 1996). In both cases, there is a requirement for increased ATP production, probably partly effected through an increase in the AMP to ATP ratio through the action of adenylate kinase, which is met by elevated glucose mobilization/influx and utilization by the cell. At the levels studied, cellular ATP levels remain constant, though there may be a non-significant transient decrease on first exposure of cells to the compounds. This contrasts strongly with the effects of rotenone, an insecticide which produces long-lasting significant decreases in ATP levels (Fendel et al. 2008). These compounds appear to have a similar mode of action to metformin (Bridges et al. 2014, Brunmair et al. 2004, El-Mir et al. 2000, Fontaine 2014, Owen et al. 2000, Zhou et al. 2001), the drug of choice for treating type 2 diabetes. Although metformin has some major effects on the liver cells (Bailey & Turner 1996, Madiraju et al. 2014, Miller et al. 2013), it also affects muscle cells to inhibit complex I activity (Brunmair et al. 2004, Wessels et al. 2014), activate AMPK (Brunmair et al. 2004, Li et al. 2015, Musi et al. 2002) and induce glucose uptake (Kumar & Dey 2002), which are its other important anti-diabetic effects. RTC-1 appears to be more potent in inhibiting mitochondrial complex I, with an IC_{50} calculated at 27 \, \mu\text{mol} \, \text{L}^{-1} compared with previously stated IC_{50} values of between 1.2 and 27 \, \mu\text{mol} \, \text{L}^{-1} for metformin (Jenkins et al. 2013, Piel et al. 2015). One possible drug-dependent side effect of complex I inhibition is reported to be lactic acidosis (Brown et al. 1998, Misbin 1977, Wang et al. 2003). Although this was not specifically tested for in these mice, the characteristic symptoms of lactic acidosis, such as nausea, vomiting, muscle weakening and rapid breathing, were not evident in the mice that were treated with the drug for up to 16 weeks. The gastrointestinal side effects associated with metformin treatment, such as diarrhoea, retching and abdominal pain (Florez et al. 2010), were also absent. There are no effects on other potential drug targets for type 2 diabetes such as the GLP-1 receptor and PPAR\gamma. Based on the toxicology screenings, there is also no evident toxicity at the doses tested.

Studies on mice fed a high-fat diet reflect this mechanism of action. Given either at the start of the diet or once insulin resistance/type 2 diabetes has been established, the compounds effected a very significant restoration of the normal GTT and ITT parameters. Again there were no indicators of toxicity: no deaths, no lethargy or hyperactivity, no diarrhoea, and no change in food intake, even on continuous treatment for up to 16 weeks. The weight gain in the mice was normalized, this was particularly evident in the epididymal fat pad (C Redondo, personal communication), which was greatly increased in the diabetic untreated animals but very similar to chow-fed controls in the compound-treated groups. The ability of these compounds to improve glucose handling in high-fat fed mice is similar to studies carried out with metformin (Matsui et al. 2010) and other compounds that target complex I (Jenkins et al. 2013).

One unexplained result was the success of RTB-70 in vivo and in the NADH dehydrogenase assay, as measured in disrupted mitochondria (IC_{50} 6.3 \, \mu\text{mol} \, \text{L}^{-1}), but it showed reduced efficacy on the inhibition of oxygen consumption as measured in intact mitochondria (IC_{50} 138 \, \mu\text{mol} \, \text{L}^{-1}) and no significant effect on glucose uptake as measured in whole cells. These results suggest that the compound may not penetrate membrane lipids very efficiently and therefore not influence the AMP to ATP ratio over the time course of the cellular experiments. This inconsistency could, however, be the result of the nature and time differences between the two kinds of experiment. It may well be that RTB-70 has a longer residence time in the animals and is therefore able to partition successfully into cells to produce similar responses over a time frame of weeks rather than hours. There is also the theoretical possibility that the compound may be metabolized in vivo to a more permeable/active derivative.

All the parameters studied thus far showed that these compounds have the potential to have much more effective anti-diabetic effect at a very much lower dose than metformin. The lower doses may prevent some of the intestinal problems associated with the higher doses required with metformin treatment (Florez et al. 2010, Kirpichnikov et al. 2002); however, both preclinical and clinical testing would need to be carried out to confirm this. The mechanism of action (schematic: Fig. 7) appears not to be as an insulin sensitizer but rather as an insulin substitute, and in that respect may also be of value to individuals with type 1 diabetes. Since there also promises to be a positive effect on weight, that in itself might attenuate the deleterious effects of excess visceral adipose tissue. It is possible in the future that these compounds can become an important tool in the treatment of type 2 diabetes.
Supplementary data
This is linked to the online version of the paper at http://dx.doi.org/10.1530/JME-15-0225.

Declaration of interest
The funding agencies had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. The authors have declared that no competing interests exist.

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Author contribution statement
JBCF, JCS, RKP, AS, DSDM and CJB conceived and designed the experiments; DSDM, SL, RD, CR, CJB, GKK, TSM, and MFR performed the experiments; DSDM, SL, RD, CR, and GKK analyzed the data and DSDM, SL, GKK, JCS and JBCF wrote the paper.

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