SFRP5 acts as a mature adipocyte marker but not as a regulator in adipogenesis

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Abstract

WNT/β-catenin signalling is involved in regulating adipogenesis, and its dysregulation occurs in obesity. Secreted frizzled-related protein 5 (SFRP5) is a WNT protein inhibitor; however, its role in adipogenesis and obesity is controversial. In this study, we observed that SFRP5 mRNA levels were increased in the fat tissues of obese humans and mice. Sfrp5 expression was gradually induced during differentiation of white and brown adipocytes and was highly increased in mature adipocytes rather than preadipocytes. However, the effects of the exogenous overexpression of Sfrp5 indicated that Sfrp5 may not directly regulate adipogenesis in vitro under the conditions studied. Moreover, SFRP5 did not inhibit the canonical WNT/β-catenin signalling pathway in preadipocytes. Subsequently, we measured the levels of circulating SFRP5 in obese patients and non-obese subjects using ELISA and did not find any significant difference. Collectively, these findings indicate that Sfrp5 represents a mature adipocyte marker gene. Our data provide new evidence concerning the role of SFRP5 in adipogenesis of white and brown adipocytes and obesity.

Key Words
- adipogenesis
- adipocyte differentiation
- WNT signalling pathway
- β-catenin
- obesity

Introduction

The obesity epidemic presents one of the largest worldwide threats to overall health. The elucidation of the molecular mechanisms that are involved in adipogenesis is critical to the development of effective strategies for the prevention and treatment of obesity and its metabolic complications (Kelly et al. 2008, Flegal et al. 2012). Obesity is frequently associated with insulin resistance (IR), which in turn is linked to the development of many chronic diseases, such as type 2 diabetes (T2DM), hyperlipidaemia, hypertension and atherosclerosis (WHO Expert Consultation 2004). Adipogenesis is related to glucose and lipid metabolism and plays a role in IR (Rosen & MacDougald 2006, Rosen & Spiegelman 2006). Adipogenesis involves the sequential activation of a cascade of transcription factors that coordinate the expression of genes responsible for the adipogenic phenotype (Wu et al. 1999, Rosen et al. 2000). Results from gene functional studies indicate that evoked WNT/β-catenin signalling in white and brown
preadipocytes leads to the maintenance of the undifferentiated state and prevents the induction of CCAAT/enhancer-binding protein α (Cebpa) and peroxisome proliferator-activated receptor γ (Pparg) (Ross et al. 2000, Kang et al. 2005, Rosen & MacDougald 2006). In contrast, the disruption of WNT/β-catenin signalling promotes adipogenesis (Bennett et al. 2002, Prestwich & Macdougald 2007).

WNT10b, which has been the most extensively studied endogenous WNT in the regulation of adipocyte differentiation, decreases during adipogenesis preceding the downregulation of cytoplasmic β-catenin. Furthermore, the ectopic expression of WNT10b activates WNT/β-catenin signalling and exhibits an anti-adipogenic action (Longo et al. 2004, Christodoulides et al. 2006, Aslanidi et al. 2007). Conversely, the extracellular antagonists of WNTs, such as dickkopfs, WNT inhibitory factor 1 and secreted frizzled-related proteins (SFRPs), have been shown to exert proadipogenic effects (Kawano & Kypta 2003). SFRPs prevent downstream WNT signalling by binding to and sequestering WNT ligands in the extracellular space (Bovolenta et al. 2008). Among SFRPs, SFRP5 may interact with both canonical and non-canonical WNT signalling pathways (Li et al. 2008, Satoh et al. 2008) and might be an import metabolic regulator (Ouchi et al. 2010, Mori et al. 2012, Carstensen et al. 2014). Results from an in vitro study indicated that SFRP5 impairs insulin signalling under normal conditions, but reduces tumour necrosis factor α-induced inflammation in primary human adipocytes (Carstensen et al. 2014). In murine mature adipocytes, SFRP5 expression is downregulated in drug-induced IR and upregulated by insulin sensitisers (Lv et al. 2012). However, in vivo studies regarding the role of SFRP5 in adiposity have yielded conflicting data. Four groups reported increased Sfrp5 expression in genetic and diet-induced obese mouse models (Koza et al. 2006, Lagathu et al. 2009, Okada et al. 2009, Mori et al. 2012), whereas another group detected decreased Sfrp5 expression in Lepob/ob and high-fat/high-sucrose diet-induced obese mice (Ouchi et al. 2010). As to the downstream signalling affected by SFRP5, Ouchi et al. (2010) reported that Sfrp5 deficiency in mice elevated phosphorylation of c-Jun N-terminal kinase, a downstream target of the non-canonical WNT signalling; however, the work from the MacDougald Laboratory indicated that SFRP5 inhibits canonical WNT signalling to suppress oxidative metabolism and stimulate adipogenesis (Mori et al. 2012). In contrast to the findings from studies using murine models, implicating SFRP5 in metabolic dysfunction, Ehrlund et al. (2013) indicated that SFRP5 in the WAT of Caucasians is not influenced by obesity. In light of the profound but conflicting effects of SFRP5 on adipogenesis and its unknown effect on brown adipocyte differentiation, we aimed to further investigate its function in adipogenesis.

SFRP5 is preferentially expressed in WAT and confined to adipocytes (Koza et al. 2006, Lagathu et al. 2009, Ouchi et al. 2010); however, whether it can be secreted into the blood for regulating metabolism is still a subject of debate. Ouchi et al. (2010) observed that detection of serum SFRP5 is only possible in mice after adenovirus-mediated i.v. injection of Sfrp5. Similarly, Mori et al. (2012) demonstrated that SFRP5 is not easily detectable in whole-cell lysates or extracellular matrix (ECM) fractions except when overexpressed in HEK293T and 3T3L1 cells. Results from several studies have indicated that circulating SFRP5 is detectable in humans (Schulte et al. 2012, Carstensen et al. 2013, Hu et al. 2013a,b, Lu et al. 2013). However, both positive (Carstensen et al. 2013) and negative (Hu et al. 2013a,b) associations between circulating SFRP5 concentrations and homeostasis model assessment of insulin resistance (HOMA-IR) were reported, whereas no statistically significant correlation between plasma SFRP5 levels and HOMA-IR was found in a Chinese cohort (Lu et al. 2013). Moreover, Ehrlund et al. (2013) reported that SFRP1, SFRP2 and SFRP4, but not SFRP3 or SFRP5, were actively secreted from human WAT and constituted adipokines. Additional investigations into the issue concerning whether SFRP5 acts in an endocrine manner are warranted.

In view of the inconclusive evidence regarding the role of SFRP5 in adiposity, we investigated the potential involvement of Sfrp5 in adipogenesis and obesity. We tested the expression level of SFRP5 in human and murine adipose tissue and further explored its effects on differentiation of white and brown preadipocytes in vitro. Additionally, we determined whether SFRP5 exhibited extracellular localisation. Finally, we detected serum SFRP5 concentrations in obese and non-obese subjects using a replicable ELISA kit to test its application as a biomarker of obesity. Our study provides a better understanding of the influence of Sfrp5 on adipocyte differentiation.

Materials and methods

Ethics statement

This study was approved by the Institutional Review Board of the Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, and was in accordance with the principle of the Helsinki Declaration II. Written informed consent was obtained from each participant.
Human adipose tissue samples
Subcutaneous and visceral adipose tissues were obtained during surgical procedures from seven non-obese and 13 obese subjects. The exclusion criteria for subjects included the following: acute or chronic infectious or immunological diseases, cancer, hepatitis, hyper- or hypocortisolism, hyper- or hypothyroidism, hypertension, diabetes mellitus and a family history of the above diseases. The detailed characteristics of the subjects are presented in Supplementary Table 1, see section on supplementary data given at the end of this article.

Animals
Leptin-deficient (Lep<sup>ob/ob</sup>) mice have an excessive appetite and become profoundly obese. Lgr4 homozygous mutant (Lgr4<sup>m/m</sup>) mice display reduced adiposity and are resistant to high-fat diet-induced obesity (Wang et al. 2013). They served as obese and lean mouse models in this study. Lep<sup>ob/ob</sup> mice and their WT littermates were age- and sex-matched throughout the experiments. Male mice, 24 weeks of age, were used in the experiments (n=6–7/group). Lgr4<sup>m/m</sup> mice were generated as described previously (Wang et al. 2012, 2013). Lgr4<sup>m/m</sup> mice show browning of white fat, which is more obvious when β-adrenergic signalling is activated by cold stress or isoprenaline treatment (Wang et al. 2013). To test gene expression in a lean mouse model regardless of experimental conditions, male Lgr4<sup>m/m</sup> mice, between 8 and 16 weeks of age, were subjected to cold room (4 °C) stress for 1 week or received injections of isoprenaline (7.5 mg/kg, Harvest Pharmaceutical, Shanghai, China) for 10 days before analysis as described previously (Wang et al. 2013). The dissected tissues were quickly frozen and then stored at −80 °C. All of the animal experiments were approved by the Animal Care Committee of Shanghai Jiao Tong University School of Medicine.

Cell culture
3T3L1, C3H10T1/2 and HEK293T cell lines were obtained from the American Type Culture Collection (Manassas, VA, USA). The 3T3L1 preadipocyte line is the ‘gold standard’ for investigating differentiation of white preadipocytes (Tang & Lane 2012). The pluripotent C3H10T1/2 cells serve as a faithful mesenchymal stem cell (MSC) model for genetic studies of the brown adipocyte developmental programme. The stromal vascular fraction of brown adipose tissue (BAT SVF) possesses a resident population of pluripotent MSCs, which enable it to serve as a model for the differentiation of brown adipocytes (Tang & Lane 2012). The details of the isolation of SVF are presented in the Supplementary materials and methods, see section on supplementary data given at the end of this article. The HEK293T cell line is readily infected and is a widely used tool for studying protein function (Thomas & Smart 2005). Murine 3T3L1 preadipocytes, C3H10T1/2 and HEK293T cells were grown in DMEM (Gibco) containing 10% foetal bovine serum (FBS) (PAA, Pasching, Austria) and 100 U/ml penicillin–streptomycin (Gibco) at 37 °C with 5% CO2.

Cell infection
Adenoviral vectors overexpressing Sfrp5 (Ad-Sfrp5-Gfp) or empty vectors (Ad-Gfp) were manufactured to our specifications by Invitrogen Life Technologies. The cells were infected at 70% confluence.

Differentiation of 3T3L1 and C3H10T1/2 cells into adipocytes
Two days after confluence has been reached, the 3T3L1 preadipocytes were incubated with DMEM containing 0.5 mM 3-isobutyl-1-methylxanthine (IBMX; Sigma), 5 μg/ml insulin (Eli Lilly) and 1 μM dexamethasone (Sigma). After 2 days (day 2), the medium was replaced with DMEM containing only 10% FBS and 5 μg/ml insulin. On day 4, the medium was removed and a fresh medium with 10% FBS was added. The medium was changed every 2 days until the cells were collected. Differentiation of confluent C3H10T1/2 stem cells into brown adipocytes was performed in a growth medium supplemented with 0.5 mM IBMX, 5 μg/ml insulin, 1 μM dexamethasone, 50 nM tri-iodothyronine (T3) (Sigma) and 5 μM troglitazone (Sigma) for 48 h. The cells were placed in a growth medium supplemented with insulin, T3 and troglitazone for another 6 days. The medium was changed every 2 days. mRNA was isolated on days 0, 1, 2, 4, 6 and 8.

Oil Red O dye staining and triglyceride content
Fully differentiated 3T3L1 adipocytes and SVF (8 days after induction) were collected and washed twice with PBS, then fixed in 4% formaldehyde for 20 min and washed twice with water. Oil Red O dye (Sigma) was dissolved in isopropanol and filtered. Cells were stained with the dye for 1 h and washed with water until the background became transparent. The images were recorded using a microscope (Olympus). 3T3L1 cell triglyceride levels were measured using a commercial kit (Kehua Bio-engineering Company, Shanghai, China) according to the manufacturer’s instructions.
RNA isolation and real-time PCR

Total RNA was extracted from the cells and adipose tissue samples using TRIzol reagent (Invitrogen) according to the manufacturer’s instructions. The RNA integrity number (RIN) of samples was checked using an Agilent 2100 Bioanalyzer (Agilent Technologies, Santa Clara, CA, USA). Samples that had an RIN higher than 7.0 were eligible for the experiment and were adjusted to the same concentration. cDNA was generated from 1 μg RNA using the RT System (Promega). The quantification of transcripts was carried out using the LC480 real-time PCR detection system (Roche Diagnostics, Penzberg, Germany) and SYBR Green I Supermix (Takara Bio Inc., Otsu, Shiga, Japan). The cycling conditions were as follows: 95°C for 30 s, 45 cycles of 5 s at 95°C followed by 30 s at 60°C. The primers used in this study are presented in Supplementary Table 2, see section on supplementary data given at the end of this article.

Protein preparation and western blotting

3T3L1 preadipocytes were treated with 3 nM WNT3a (R&D company, Minneapolis, MN, USA) and 1 μg/ml recombinant murine SFRP5 protein (rmSFRP5) (R&D company) individually or in combination. In another treatment, 3T3L1 preadipocytes were infected with Ad-Gfp or Ad-Sfrp5-Gfp respectively. After 72 h, the nuclear and cytoplasmic protein extracts were isolated using NE-PER reagent according to the manufacturer’s protocol (Pierce Biotechnology, Inc., Rockford, IL, USA). The protein concentration was determined by the BCA protein assay (Pierce Biotechnology, Inc.). Total protein was denatured by boiling, separated on a 10% SDS–PAGE gel and transferred onto a nitrocellulose membrane (Amersham Biosciences). After blocking in Odyssey blocking buffer (LI-COR, Lincoln, NE, USA) for 1 h, the membranes were incubated overnight at 4°C with primary antibodies directed against β-catenin (Cell Signaling Technology (CST), Danvers, MA, USA), α-tubulin (CST), and Lamin B (Santa Cruz Biotechnology, Inc.). The secondary antibody used was anti-rabbit IgG (LI-COR) or anti-goat IgG (LI-COR), and the resulting bands were visualised using the Odyssey infrared imaging system (LI-COR).

Localisation of SFRP5

The separation of cell lysates, ECM, and conditioned medium fractions was performed as described previously (Lee et al. 2004, Mori et al. 2012). Briefly, the conditioned media were collected and concentrated by centrifuging at 3000g for 15 min. The confluent HEK293T cells were released from the culture dishes by incubation with PBS containing 5 mM EDTA at 37°C in an atmosphere of 5% CO2 and were gently shaken every 10 min, for a total of 40 min. The cells were then verified to be completely released from the dishes using a microscope and collected. After washes, the remaining ECM components were extracted with 1× SDS Laemmli sample buffer containing 5% β-mercaptoethanol and subjected to western blot analysis.

Measurement of SFRP5 in human serum

A total of 160 young adults living in the eastern area of China (Han Chinese) were recruited for this study. A total of 80 patients with obesity were collected consecutively from January 2010 to October 2012 from the specialised outpatient clinic for obesity in Ruijin Hospital, Shanghai Jiao Tong University School of Medicine. The inclusion criteria for patients with obesity were as follows: aged 18–30 years, with BMI ≥30 kg/m2, previously untreated obesity, fasting blood glucose level lower than 7.0 mmol/l and 2h-postprandial glucose levels according to the oral glucose tolerance test (OGTT) lower than 11.1 mmol/l. Patients with syndromic obesity, diabetes mellitus, hyper- and hypocortisolism, hyper- and hypothyroidism, growth hormone deficiency, severe hepatic or renal disease, acute or chronic infectious or immunological disease, cancer and pregnancy or breastfeeding, or being treated with glucose-lowering drugs such as metformin were excluded from our study. Conforming to the age and sex distribution of the obese patients, 80 unrelated non-obese subjects were recruited from volunteers of Shanghai Jiao-Tong University School of Medicine. The inclusion criteria for controls were as follows: aged 18–30 years, with BMI <23 kg/m2 (WHO Expert Consultation 2004), currently physically well and with no past history of obesity. Non-obese subjects with clinical evidence of major diseases, hypertension, impaired glucose regulation and diabetes mellitus were excluded from our study. Blood samples were obtained after an overnight fast. Goat and mouse anti-human SFRP5 antibodies (Santa Cruz Biotechnology, Inc.) were used to produce the ELISA kit by Shanghai Senxiong Biotech (Shanghai, China). The assay was proven to be highly specific to human SFRP5 (intra-assay coefficient of variation (CV) <10%; inter-assay CV <15%; analytical-sensitivity, 16 ng/ml). The ELISA was performed according to the instructions from Shanghai Senxiong Biotech.

Statistical analysis

The Shapiro–Wilks test was used to test the normality of distribution for the data, and variables with P values <0.05...
were logarithmically transformed for statistical analyses. The correlations between circulating SFRP5 levels and other metabolic parameters were calculated using Spearman’s correlation coefficients. When we explored the correlation between SFRP5 and other metabolic parameters, we further adjusted for age and sex. All of the analyses were performed using SAS (version 9.3; SAS Institute, Inc., Cary, NC, USA). The data are presented as the means ± S.E.M. unless stated otherwise. Two-tailed Student’s t-test or ANOVA followed by posthoc analysis were used for comparisons between groups, as appropriate. Differences were considered significant for P values < 0.05.

**Results**

**SFRP5 is increased in adipose tissues of obese humans and mice**

To investigate the role of SFRP5 in WAT biology, we first detected the mRNA expression levels of SFRP5 in visceral WAT (vWAT) and subcutaneous WAT (sWAT) from obese and non-obese subjects. The results of real-time PCR showed that the SFRP5 mRNA expression was robustly increased in vWAT from obese patients compared with non-obese controls (Fig. 1A). No significant difference in SFRP5 mRNA levels was detected in sWAT from obese patients compared with non-obese subjects (Fig. 1B).

We examined Sfrp5 mRNA levels in epididymal WAT (eWAT) and inguinal WAT (iWAT) of Lepob/ob mice and WT mice. Sfrp5 displayed a higher level in eWAT of Lepob/ob mice than in WT mice, although this trend did not reach statistical significance (Fig. 1C). Sfrp5 expression was significantly enhanced in iWAT of Lepob/ob mice (Fig. 1D). Consistent with the findings from studies using the obese mouse model, Sfrp5 mRNA expression was reduced in eWAT of a lean mouse model, Lgr4m/m mice (Wang et al. 2013), under normal conditions, cold stress or isoprenaline treatment (Fig. 1E). Notably, isoprenaline treatment itself induced expression of Sfrp5 (Fig. 1E). These results indicate that SFRP5 is increased in the fat tissue of obese human vWAT and murine iWAT, which indicates that SFRP5 expression in WAT might be an indicator of obesity.

**Sfrp5 level is gradually elevated during differentiation of white and brown adipose tissue**

To study the biological function of Sfrp5 in adipogenesis, we assessed the changes in Sfrp5 expression during differentiation of white and brown adipocytes. We found that Sfrp5 expression incrementally increased during the differentiation of 3T3L1 cells, which was consistent with upregulation of the key transcription factors for adipogenesis, such as CEBPA and PPARγ, and reached maximal expression 8 days after the initiation of differentiation (Fig. 2A, B and C).

During differentiation of C3H10T1/2 cells, although the groups were statistically underpowered, Sfrp5 mRNA demonstrated a tendency towards higher levels that paralleled the increased expression of brown-adipocyte-differentiation-associated genes, including PR domain containing 16 (Prdm16), Pparg coactivator 1 alpha (Ppargc1a) and cell death-inducing DNA fragmentation factor, alpha subunit-like effector A (Cidea) (Fig. 2D, E, F and G). Moreover, we detected similar gene expression patterns during the differentiation of BAT SVF (Supplementary Figure 1).

![Figure 1](http://jme.endocrinology-journals.org)

**Figure 1**

SFRP5 mRNA is increased with obesity in humans and mice. (A) and (B) SFRP5 mRNA was examined by real-time PCR in human visceral WAT (vWAT) and subcutaneous WAT (sWAT) from obese (n = 13) and non-obese (n = 7) subjects. *P < 0.05 vs non-obese controls. Mean values are plotted with S.E.M. (C) and (D) Sfrp5 mRNA was examined by real-time PCR in epididymal WAT (eWAT) and inguinal adipose tissue (iWAT) of 24-week-old male wild-type (WT, n = 11) and Lepob/ob (n = 11) mice. WT, wild type; ob/ob, Lepob/ob. (E) Sfrp5 mRNA in eWAT of WT and Lgr4m/m (m/m) mice under different conditions (n = 6–9 for wild type; and n = 5–8 for m/m). NC, normal conditions; CR, cold-room stimulation; ISO, isoprenaline treatment. WT, wild type; m/m, Lgr4m/m. Differences among groups were examined by ANOVA followed by the Bonferroni’s multiple comparison analysis. *P < 0.05 vs WT mice under normal conditions. For (C), (D) and (E), mean values are plotted with S.E.M.; *P < 0.05, **P < 0.01 and ***P < 0.001 vs the corresponding WT mice.
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Figure 2

Sfrp5 mRNA is induced during differentiation of white and brown adipose tissue. Total RNA was isolated at the indicated time-points, subjected to real-time PCR and expression was normalised with respect to acidic ribosomal phosphoprotein P0 (Rplp0) mRNA expression. Expression levels of (A) Sfrp5, (B) Cebpa and (C) Pparg were elevated incrementally during differentiation of 3T3L1 adipocytes. Expression levels of (D) Sfrp5, (E) Prdm16, (F) Pparg1a and (G) Cidea were elevated incrementally during differentiation of 3T3L1 cells. Mean values are plotted with S.E.M. (n = 3). Differences among groups were calculated by ANOVA followed by post-hoc Dunnett’s test. *P<0.05, **P<0.01 and ***P<0.001 vs day 0.

Effects of Sfrp5 overexpression on adipocyte differentiation

We then examined whether SFRP5 could promote adipogenesis of white or brown adipose tissue. 3T3L1 preadipocytes, C3H10T1/2 cells and BAT SVF were infected with adenoviral vectors encoding Sfrp5 (Ad-Sfrp5-Gfp) or empty vectors (Ad-Gfp). The total mRNA was isolated to determine the expression of adipocyte markers, and Oil Red O staining was performed to observe the morphology of lipid droplets in mature differentiated adipocytes. As shown in Fig. 3A and B, and Supplementary Fig. 1F, Sfrp5 overexpression did not result in increased expression of white-adipocyte-specific marker genes, including resistin (Retn), homeobox C9 (Hoxc9), Transcription factor 21 (Tcf21), Inhibin beta B (Inhibb) and Serine or cysteine proteinase inhibitor, clade A, member 3K (Serpin3K), or brown adipocyte-specific marker genes, including Prdm16, Pparg1a and Cidea as expected (Fig. 3A and B, Supplementary Fig. 1F). Oil Red O staining revealed that Sfrp5 overexpression did not change the number of lipid droplets in differentiated 3T3L1 (Fig. 3C), C3H10T1/2 cells (data not shown) or BAT SVF (Supplementary Fig. 1G). Thus, although Sfrp5 is increased in fat tissues with obesity and during adipogenesis, it seems that SFRP5 does not affect adipogenesis of white or brown adipose tissue.

SFRP5 does not affect WNT/β-catenin signalling in preadipocytes

SFRP5 has been identified as an inhibitor of WNT signalling; however, its role in canonical WNT signalling in preadipocytes remains undetermined (Ouchi et al. 2010, Mori et al. 2012). 3T3L1 preadipocytes were treated with rmSFRP5 and canonical WNT signalling ligand WNT3a, individually or in combination. As shown in Fig. 3D, WNT3a treatment resulted in an increased translocalisation of β-catenin to the nucleus, whereas rmSFRP5 treatment did not change basal or WNT3a-induced nuclear β-catenin levels. Consistent with the results of rmSFRP5 treatment, the Ad-Sfrp5-Gfp-infected 3T3L1 cells displayed an unaltered amount of β-catenin in either the cytoplasm or the nucleus compared with controls. Altogether, these findings indicate that SFRP5 does not appear to regulate WNT/β-catenin signalling in preadipocytes.
Figure 3
Sfrp5 overexpression does not affect 3T3L1 and C3H10T1/2 cell differentiation or the translocation of β-catenin to the nucleus in 3T3L1 preadipocytes. (A) Exogenous overexpression of Sfrp5 did not alter expressions of Retn, Hoxc9, Tcf21, Ihhbb and Serpina3K in 3T3L1 adipocytes after infection with Sfrp5 (day 8 after differentiation; n = 5). (B) Exogenous overexpression of Sfrp5 did not alter expressions of Prdm16, Pparc1a and Cidea in C3H10T1/2 stem cells after infection with Sfrp5 (day 8 after differentiation; n = 3). (C) Sfrp5 overexpression did not affect adipocyte differentiation. (Left panel) Pictures of Oil Red O (ORO) staining showing lipid accumulation in 3T3L1 adipocytes. Objective magnification, ×10. (Right panel) Triglyceride (TG) content of 3T3L1 adipocytes (day 8 after differentiation; n = 5). Ad-Gfp, 3T3L1 adipocytes infected with an empty adenoviral vector; Ad-Sfrp5, 3T3L1 adipocytes infected with an adenoviral vector encoding Sfrp5. For (A), (B) and (C), mean values are plotted with s.e.m.; *P<0.05 and **P<0.01 vs Ad-Gfp. (D) SFRP5 does not affect β-catenin translocation in 3T3L1 preadipocytes. (Upper panel) Quantification of β-catenin in cytoplasm and nucleus of 3T3L1 preadipocytes. (Lower panel) 3T3L1 preadipocytes were treated with PBS, WNT3a and recombinant mouse SFRP5 individually or in combination. Ad-Gfp, 3T3L1 preadipocytes infected with an empty adenoviral vector; Ad-Sfrp5, 3T3L1 preadipocytes infected with an adenoviral vector encoding Sfrp5. Mean values are plotted with s.e.m. (n = 5). Differences among groups were calculated by ANOVA followed by the Bonferroni's multiple comparison analysis. **P<0.001 vs preadipocytes treated with PBS; and ***P<0.001 vs preadipocytes treated with recombinant mouse SFRPS.

Serum SFRP5 levels cannot be used to distinguish obese patients from non-obese subjects

On the basis of our data, SFRP5 is highly enriched in mature adipocytes and adipose tissues of obese humans and mice. Thus, SFRP5 might be a circulating indicator of obese patients. To address this issue, we measured serum SFRP5 concentrations of 80 non-diabetic obese and 80 age- and sex-matched non-obese subjects (Supplementary Table 3, see section on supplementary data given at the end of this article). The specificity of the antibodies used in ELISA was tested by western blot experiments (Fig. 4A and B). The standard curve confirmed a wide detection range between 0 and 800 ng/ml (Fig. 4C). SFRP5 was detectable in non-obese (41.47 ± 2.26 ng/ml) and obese subjects (44.84 ± 2.36 ng/ml); however, there was no statistically significant difference between the two groups (Fig. 4D). Additionally, no significant associations were found between the circulating SFRP5 levels and anthropometric measurements, including BMI, waist circumference, percentage body fat, the biochemical characteristics and HOMA-IR (Supplementary Table 4, see section on supplementary data given at the end of this article).

SFRP5 is incorporated into the ECM

SFRP1 and SFRP2 are thought to act locally by remaining tightly associated with the ECM. To determine whether SFRP5 exhibits similar extracellular localisation, HEK293T cells were infected with Ad-Gfp or Ad-Sfrp5-Gfp. Cell lysates,
Figure 4
SFRP5 is tightly associated with the extracellular matrix (ECM), and serum SFRP5 concentrations cannot be used to distinguish obese patients from non-obese subjects. (A and B) Expression and localisation of SFRP5 in vitro. SFRP5 protein is secreted but remains associated with the ECM. Western blot confirming the specificity of goat (A) and mouse (B) anti-human SFRP5 antibodies used in ELISA. Ad-Gfp, HEK293T cells infected with an empty vector; Ad-Sfrp5, HEK293T cells infected with an adenoviral vector encoding Sfrp5; CL, cell lysates; CM, conditioned media. (C) The standard curve confirmed a wide detection range of the ELISA Kit. (D) Serum samples were obtained after an overnight fast and SFRP5 levels were measured using ELISA. SFRP5 concentrations are similar in obese (n = 80) and non-obese (n = 80) subjects. Mean values are plotted with S.E.M.

Discussion
In the last decade, emerging evidence has indicated an important role for the WNT signalling pathway in adipogenesis (Ross et al. 2000, Kennell & MacDougald 2005). Research regarding the regulation of Sfrp5 and obesity demonstrates conflicting results. A signal-net array examined in our previous study indicated a notable decline in Sfrp5 mRNA expression in lean Lgr4m/m mice compared with their normal-weight littermates (J Wang, R Liu, F Wang, M Chen, J Hong and G Ning, unpublished observations). Consistently, results obtained from compelling experiments indicate that Sfrp5 is highly induced in mouse models of genetic and diet-induced obesity, such as Lepdb/db, leptin receptor-deficient (Lepdb/db) and C57BL/6J mice exposed to a high-fat diet (Koza et al. 2006, Lagathu et al. 2009, Okada et al. 2009, Mori et al. 2012), and support a model of adipogenesis in which SFRP5 inhibits WNT/β-catenin signalling to stimulate adipocyte growth (Lagathu et al. 2009, Lv et al. 2012, Mori et al. 2012). In contrast, Ouchi et al. (2010) reported downregulation of SFRP5 under these conditions and proposed that SFRP5 neutralised non-canonical JNK activation by WNT5a in murine adipocytes. In this study, we observed that exogenous SFRP5 treatment did not change basal or WNT3a-induced nuclear β-catenin levels, which indicates that SFRP5 does not work through canonical WNT/β-catenin signalling in preadipocytes and that other signalling, such as WNT5a, might be involved (Kawano & Kypta 2003, Ouchi et al. 2010, Cruciat & Niehrs 2013). Further mechanism studies should also investigate the non-classical roles of SFRPs, including the interaction of SFRP5 with the cell-surface receptors and many other signalling networks, such as transforming growth factor beta, BMPs and FGFs (Lee et al. 2006, Placencio et al. 2008, Cruciat & Niehrs 2013).

In this study, we clarified that SFRP5 was increased overall in fat tissues under an obese state, which indicates that SFRP5 may play an important role in the development of obesity. Nevertheless, our results indicate a different expression pattern of SFRP5 between humans and mice. This might be due, in part, to the different features of fat depots between humans and mice. For example, a recent publication reports that the functional profile of human sWAT is similar to that of mouse mesenteric and perigonadal depots (Cypess et al. 2013).

To date, studies of the action of Sfrp5 in differentiation of brown adipocytes are limited. Our present data first revealed that Sfrp5 mRNA displayed a tendency towards higher levels during differentiation of brown adipocytes in vitro, parallelizing the increased expression of brown adipocyte differentiation-associated genes. Consistent with our results, previous in vitro evidence has indicated a gradual induction of SFRP5 expression during differentiation of white adipose tissue (Lv et al. 2012, Mori et al. 2012). However, we found that the exogenous expression of Sfrp5 leads to unaltered expression levels of Cebpa, Pparg (data not shown) and white- or brown-adipocyte-specific marker genes. The MacDougald Lab also failed to verify a regulatory role for Sfrp5 in ear MSC differentiation, with no marked influence of Sfrp5 ablation in the morphology of lipid droplets or expression of common adipocyte marker genes such as Pparg and Fabp4 observed (Mori et al. 2012). It seems that these results cannot explain the decreased fat weight in the
Sfrp5-deficient mouse model (Mori et al. 2012), which might be caused by the fact that, although in vitro adipogenesis can be synchronously induced by exposure to a defined adipogenic cocktail, in vivo adipose tissue expansion is regulated by a combination of local and endocrine factors.

Our previous results have indicated that Lgr4<sup>m/m</sup> mice displayed reduced body weight when compared with WT mice. Lgr4<sup>m/m</sup> mice showed browning of white fat, which was more obvious when being activated by cold stress or isoprenaline treatment (Wang et al. 2013). In this study, we detected SFRP5 expression in WT and Lgr4<sup>m/m</sup> mice under different conditions to demonstrate a reduction in SFRP5 in a lean mouse model regardless of experimental conditions. Notably, we observed that isoprenaline treatment but not cold-room stimulation induced expression of Sfrp5. To our knowledge, both cold-room stimulation and isoprenaline treatment activate β-adrenergic signaling. The detailed molecular mechanism as to how isoprenaline regulates SFRP5 expression is not clear and future studies are needed to address this issue.

WAT appeared to be one of the major sources of SFRP5 in obese mice (Ouchi et al. 2010, Mori et al. 2012). It is important to establish whether SFRP5 influences metabolism in distal tissues and organs through an endocrine mechanism, similar to leptin or adiponectin. In line with observations in adipocytes (Mori et al. 2012), we found that, when overexpressed, SFRP5 was incorporated into the ECM, and its effects may be more relevant locally. As SFRP5 has been reported to be highly expressed in retinal pigment epithelium and be moderately expressed in the pancreas (Chang et al. 1999), the circulating SFRP5 concentrations might not reflect their expression levels in WAT. This notion is corroborated by our findings that the serum SFRP5 levels were clearly measurable but could not be used to distinguish obese patients from non-obese subjects, which is consistent with the findings of two other studies on Caucasians (Schulte et al. 2012, Carstensen et al. 2013).

In contrast with ours and others’ data (Schulte et al. 2012, Hu et al. 2013a,b), Carstensen et al. (2013) have reported a much wider range of circulating SFRP5 concentrations in mostly overweight and obese Caucasians. It is likely that this disparity is partly related to the different clinical characteristics of these studies, including glycaemia levels, blood pressure, certain medications, other diseases and the variance in the definition of obesity. Although both positive (Carstensen et al. 2013) and negative (Hu et al. 2013a,b) associations between circulating SFRP5 concentrations and HOMA-IR have been reported, we did not observe a statistically significant correlation between serum SFRP5 levels and HOMA-IR in our cohort consisting of a total of 160 young adults with a broad range of BMI. The reasons for this discrepancy might be a comparatively healthier glucose metabolic state (only 21 obese patients had impaired glucose regulation), a much younger age and a higher degree of obesity in our study population. This study focused on obesity in young adults, so that our results may not be fully generalisable to older populations. Moreover, another study on Chinese subjects including 82 subjects with T2DM and 42 controls also detected no statistically significant correlation between plasma SFRP5 concentrations and HOMA-IR ($r=0.148$, $P=0.101$). Unfortunately, this study did not mention whether this association was different between non-obese and obese subjects (Lu et al. 2013).

In summary, our findings demonstrate that the change in circulating SFRP5 levels may differ from the elevated Sfrp5 expression in fat tissue in an obese state. Moreover, Sfrp5 is enriched in both white and brown mature adipocytes, but the exogenous overexpression of Sfrp5 does not affect adipogenesis, which indicated that, as an adipokine, SFRP5 gene upregulation appears to be caused by obesity and that it might thus be a novel mature adipocyte marker gene.

Supplementary data
This is linked to the online version of the paper at http://dx.doi.org/10.1530/JME-14-0037.

Declaration of interest
The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

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