Transcriptional coregulators: emerging roles of SRC family of coactivators in disease pathology

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Historical perspective

Transcriptional coactivators are defined, broadly, as the family of coregulator molecules that interact with nuclear receptors and other transcription factors to enhance the rate of gene transcription. The existence of coactivator-like proteins was predicted in early 1970s, as some nuclear, nonhistone receptor-associated proteins were found to bind nuclear receptors and increase their interaction with DNA to enhance their transcription potential (Spelsberg et al. 1971). This crude fraction was later shown to contain many diverse coactivators; a large number of such proteins were unpredicted at the time and prevented purification. Although it was clear that steroid hormones such as estrogen can rapidly induce the new synthesis of specific mRNA and proteins (Means et al. 1972), the importance of these nuclear acceptor molecules in ligand-dependent functions was postulated to enhance nuclear receptor (NR) transcription but the concept was not proven (Yamamoto & Alberts 1975). In the interim, a series of sophisticated molecular studies unfolded, which indicated that ligand binding activates conformational changes in the steroid receptor to promote DNA-binding and transcriptional activity; anti-hormones were shown to effectively oppose such structural alterations (Allan et al. 1992). In addition to ligand-dependent functions, the steroid receptors were also found to be activated in a ligand-independent manner (Denner et al. 1990, Power et al. 1991).

In the 1990s, studies designed to elucidate the functional roles of the corepressors and coactivators were commenced again, initially in yeast (McDonnell et al. 1991a,b, Baniahmad et al. 1993). An inherent negative regulatory function for the steroid receptors was identified in steroid receptors themselves and was analyzed first in yeasts by demonstrating binding of steroid receptors to repressors such as SSN6, which when mutated allowed receptor activation of gene expression (McDonnell et al. 1992, Vegeto et al. 1992). Similar yeast studies were carried out to demonstrate ligand-mediated coactivation. These proof-of-principle yeast studies led to the definition of two classes of coregulators – coactivators and corepressors – and were followed by the biochemical discovery of a corepressor activity for TR in mammalian cells and the
publications of other receptor-associated proteins in mammals (Cavailles et al. 1994, Halachmi et al. 1994, Baniahmad et al. 1995a,b). In aggregate, these studies set the stage for the first cloning of a cDNA encoding a mammalian nuclear receptor-interacting coactivator protein. This first authentic NR coactivator, termed steroid receptor coactivator 1 (SRC-1 (NCOA1)), was identified using yeast two-hybrid genetic screening employing the ligand-binding domain of the progesterone receptor (Onate et al. 1995, Xu et al. 1998). SRC-1 was the first member of the p160 family of coactivators cloned, following which two additional family members SRC-2 (NCOA2/GRIP1/TIF2; Voegel et al. 1996) and SRC-3 (NCOA3/ACTR/pCIP; Chen et al. 1997, Torchia et al. 1997) were identified. The p160 family members are closely related molecules with ~60% homology, but are functionally distinct. In addition to the full-length SRCs, some shorter forms of SRCs were identified as well. SRC-3A is a splice isoform of SRC-3 with a deletion of exon 4 (SRC-3A4) and the protein lacks the N-terminal helix–loop–helix (bHLH) domain that contains a nuclear localization signal (Reiter et al. 2001, Long et al. 2010). More recently, a shorter 70 kDa isoform of SRC-1 has been identified and found to be highly elevated in human and mouse endometriotic tissues (Han et al. 2012). This 70 kDa isoform of SRC-1 is the C-terminal fragment of the full-length SRC-1, which is proteolytically cleaved by MMP9. Over the last two decades, we gained considerable knowledge about the coactivators and their impact on human health and physiology. These findings together classified a novel family of nuclear receptor coactivators that became known as the master regulators of gene regulation.

**Coactivator complexome**

After the discovery of the first authentic coactivator SRC-1, it was predicted that cells may have approximately five to ten coactivators and few corepressors to regulate the gene transcription. Surprisingly, more than 400 coregulators have been reported so far, substantiating their prevalent and critical role in transcriptional regulation (Lonard & O’Malley 2007). Molecular analyses by mass spectrometry identified that SRCs work in tandem with other coregulators in close association by forming large multi-subunit stable complexes. This proteomics information concerning a coactivator–protein complex also known as ‘complexome’ – identified that the complexes are in a dynamic rearrangement in an ordered manner to facilitate various reactions and subreactions in transcription. These reactions include phosphorylation, ubiquitination, methylation, and acetylation of the associated molecules in the coactivator complex, which further defines the specific affinity of the coactivators for NRs, transcription factors, and other associated molecules (Han et al. 2009). This multifunctional component of the coactivator complexome allows them to integrate different upstream environmental stimuli and to transmit to a variety of enzymatic activities at the promoter for regulating transcription.

Proteomic investigations identified the dynamic nature of a SRC-3 complex assembled on estrogen response element (ERE) in a ligand-dependent manner (Fig. 1A). The SRC-3 complex consists of several interacting partners with enzymatic activities, which include kinases, ATPases, acetyl transferases, methyl transferases as well as ubiquitin ligases, all of which contribute to the dynamic functions of the coactivators (Malovannaya et al. 2010). Recent studies on coregulator dynamics have identified some novel mechanisms for ER-regulated gene transcription, and the findings postulated a ‘three-state model’ of coactivator-dependent complex formation (Foulds et al. 2013). In the first step, ligand-bound ER on canonical EREs forms a biochemically stable ‘poised’ complex by attracting a set of coactivators and certain corepressors. Addition of ATP rapidly converts these complexes into an ‘activated’ state by the kinetic activity of DNA-dependent protein kinase (DNA-PK), which mediates phosphorylation events on coactivators and ER. Finally, DNA-PK promotes ERα-mediated transcription by phosphorylating coactivators SRC-3 and MED1 as well as dismissing corepressors RIP140 from the complex (Foulds et al. 2013). These studies unravel the dynamic events mediated by kinases on a coactivator complexome to fine-tune transcription.

Integrated mass spectrometry-based analysis of affinity-purified endogenous coregulator complexes identified a hierarchical organization of protein complexes, which exists as three discrete layers in an intrinsically tiered organization of the complexome (Malovannaya et al. 2011). These include relatively stable minimal endogenous core modules; these combine to form the variable core complex isoforms; finally, coregulator complex–complex interactions form networks. Based on the type of protein complexes formed, the coregulators can be broadly classified into two major types: type 1 represents relatively stable multi-subunit complexes consisting of conserved coactivator molecules, whereas type 2 represents context-dependent coactivators that are recruited in response to various extra-cellular stimuli (Malovannaya et al. 2011). Type 1 coregulators include mediators, corepressor–repressor element 1-silencing transcription factor (CoREST) complexes, nuclear receptor...
corepressors (NCOR), nucleosome remodeling and deacetylation (NuRD) complexes, and the SWI/SNF (BAF/P-BAF), whereas SRCs are prime examples of type 2 complexes. This dynamic regulation of coactivator complex assembly by the SRCs is in turn regulated by various upstream signaling events that impart post-translational modifications (PTMs) onto the coactivators (Dasgupta et al. 2014).

**Signal-specific PTM codes on SRCs**

The molecular recognition of the activity of SRCs depends upon the PTM codes on them. Phosphorylation, acetylation, sumoylation, ubiquitination, and methylation of the SRCs (Fig. 1B) intricately coordinate and fine-tune their activity, localization, and protein stability and dictate the interacting partner molecules used to build up the activated transcription complex. Post-translational modifications (PTMs) on SRCs such as phosphorylation (P), acetylation (Ac), and methylation (Me) also regulate the coactivator complex association and modulate the assembly of general transcription factors such as TATA-binding protein (TBP) and TAF (TBP-associated general transcription factors) along with RNA polymerase II (PolII).

**Phosphorylation**

In response to multiple upstream signaling events such as growth factor, cytokine, hormone, and nutrient signaling, PKs phosphorylate SRCs either at a single site or at multiple sites. Depending on the pattern of the phosphorylation code(s) on SRCs, they attract selective binding...
partners, nuclear receptors or transcription factors along with other coregulator molecules to regulate the gene transcription. In addition to exerting effects on the nuclear genome by binding directly to the NRs, steroid hormones also activate several kinases such as MAPK, JNK, AKT, and ERK1/2, which then phosphorylate NRs and coactivators to stimulate gene transcription by non-genomic signaling (Lonard & O’Malley 2007). Steroid hormone signaling phosphorylates SRC-3 at multiple residues including N-terminal Thr24, several sites in a serine/threonine-rich region, and Ser857, Ser860, and Ser867 in the receptor-interacting domain (RID); Wu et al. 2004, Yi et al. 2005, 2008, Long et al. 2012). Similarly, SRC-1 is phosphorylated on Thr1179 and Ser1185, and SRC-2 on Ser736 by MAPK, thereby increasing coactivator’s affinity to NRs (Rowan et al. 2000, Gregory et al. 2004). SRC-2 has emerged as a major coactivator for glucocorticoid receptor (GR) and certain phosphorylation events on SRC-2 by casein kinase (CK) and cyclin-dependent kinase 9 dictate GR actions (Dobrovolna et al. 2012). Four major phosphorylation sites Ser469, Ser487, Ser493, and Ser499 in the N-terminal domain of SRC-2 protein promote GR-dependent transcription by facilitating recruitment of coactivator complex to native GR targets (Dobrovolna et al. 2012). SRC-3Δ4, the splicing variant of SRC-3, also is regulated by phosphorylation. But instead of a direct role in nuclear transcription, the SRC-3Δ4 is localized to the cytosol and is phosphorylated by PAK kinase, whereupon it then binds to epidermal growth factor receptor (EGFR) and transduces activity to focal adhesion kinase (FAK). Thus, phosphorylated SRC-3Δ4 acts as a critical signaling molecule to regulate the migratory potential of tumor cells by bridging the gap between EGFR and FAK (Long et al. 2010). In summary, coactivators are molecular integrators of upstream signaling events, and phospho-coded SRCs direct assembly of specific interacting partners for gene transcription.

**Acetylation and methylation**

Histone acetylases and deacetylases, along with methylases and demethylases, are essential components of coactivator complexes responsible for modifying chromatin. Based on their function of adding or removing histone marks, they are classified as epigenetic ‘writers’ or ‘erasers’. A number of co-activators including CREBBP (p300/CBP), GCNS (KAT2A), and PCAF (KAT2B) possess intrinsic histone acetyltransferase (HAT) activity (Couture & Trievel 2006). SRCs recruit the HATs and methyl transferases such as peptidylarginine methyltransferases to remodel chromatin and regulate gene transcription. Additionally, a coactivator such as SRC-3 is in turn acetylated by p300/CBP and methylated by coactivator-associated arginine methyltransferase 1 (CARM1) at Arg1171 (Feng et al. 2006). Acetylation of SRC-3 by CBP coincides with the attenuation of hormone-induced gene transcription by enforcing the complex disassembly (Chen et al. 1997, 1999). Mechanistically, acetylation neutralizes the positive charges of two lysine residues adjacent to the ‘LXLLL’ motif of SRC-3, thereby disrupting the association of HAT complexes with the NR coactivator complex and terminating the gene transcription (Chen et al. 1999). CARM1, which activates transcription by modifying core histone tails, also promotes dissociation of coactivator complex and terminates hormone-induced transcription by methylating SRC-3 (Feng et al. 2006). In addition to the acetylases, the family of lysine deacetylases, histone deacetylases (HDACs), and sirtuin proteins also regulate gene transcription as coregulators (Lahue & Frizzell 2012). HDACs are recruited to the coregulator complex to repress gene transcription, in particular by corepressors such as NCoR. There are two classes of HDACs, classes I and IIa, the latter being relatively weak in enzymatic activity. Additionally, sirtuins, the NAD-dependent deacetylases, are also recruited to the coregulator complex and are known to modulate gene transcription.

**Ubiquitination and sumoylation**

Activity and stability of coactivators are regulated by ubiquitination, an enzymatic process in which 8.5 kDa small molecules named ubiquitin are systematically added by E3 ubiquitin ligase. Ubiquitination is a highly regulated process, and phosphorylation on coactivators acts as a priming event for this modification by increasing their affinity toward ubiquitin E3 ubiquitin ligase. Phosphorylation by GSK3β on SRC-3-Ser505 increases coactivator’s affinity toward Fbw7α, a component of E3-ligase complex which then ubiquitinates SRC-3 on Lys723 and Lys786 (Lonard & O’Malley 2007, Wu et al. 2007). Mono-ubiquitinated SRC-3 has a higher affinity for ERα and stimulates ERα-dependent gene transcription, whereas poly-ubiquitinated SRC-3 is rapidly degraded, thereby decreasing SRC-3 protein stability. SRC-3 protein stability and activity are also regulated by specific phosphorylation codes that induce degradation of the protein known as ‘phospho-degron’ in the N-terminal domain of the protein; phosphorylation of Ser102 in the degron by CKI increases coactivator’s affinity for speckle-type POZ protein (SPOP)-E3 ligase (Li et al. 2008). On the contrary,
certain mutations in the SPOP protein alter the affinity of SPOP for SRC-3 imposing a SPOP-dependent regulation of SRC-3 activity and gene transcription (Geng et al. 2013). Similarly, CUL3, a member of the family of E3-ligase scaffolding proteins also modulates SRC-3 activity by binding to the Ser860-phosphorylated SRC-3 in response to retinoic acid induction (Ferry et al. 2011). Thus, PTMs on SRC-3 by phosphorylation-coupled ubiquitination modulate the activity and stability of the coactivator to control the dynamics of transcription.

In addition to ubiquitination, covalent modifications by addition of a small ubiquitin-like modifier (SUMO) to the lysine residues of the coactivators have been identified. SRCs are subjected to sumoylation at two conserved lysine residues in the RID motif, which functionally enhance their interaction and affinity for NRs (Wu et al. 2006). However, sumoylations of SRC-3 on Lys723 and Lys796 were found to have a negative impact on its activity, most probably due to the competitive inhibition of ubiquitination in these sites. Nevertheless, sumoylation of coactivators provides another degree of dynamic regulation to monitor and manipulate gene transcription.

Coactivators in disease pathophysiology

Coactivators have emerged as cellular integrators of various upstream signaling pathways that transduce these signals into transcriptional outputs to regulate expression of myriad gene targets (Fig. 2). Hence, dysfunctions in coregulators are principal drivers of numerous pathologies (Lonard & O’Malley 2012). Herein, we will highlight selected examples of the clinicopathological conditions affected by the transcriptional coactivators.

Neurological disorders

Mutations in certain coregulator genes alter the epigenetic marks on chromosomes, affecting brain development and promoting onset of certain neurodevelopmental disorders (Urdinguio et al. 2009). These epigenetic dysfunctions cause moderate to severe perturbations in the transcriptomics, disrupting the neuronal growth and differentiation. Mutations in the chromatin remodeling protein ATRX (ATP-dependent helicase ATRX, X-linked helicase II) confer aberrant DNA methylating patterns in the chromatin leading to a neurodegenerative disorder named ATRX syndrome (Gibbons et al. 2008). This syndrome is an X-linked disorder confined only to the males while the female carriers manifest limited symptoms. Symptoms include mental retardation often accompanied with α-thalassemia, unusual facial appearance and urogenital defects (Gibbons et al. 1995). ATRX is a member of the Snf2 family of enzymes, which maintains nucleosome stability and regulates gene transcription by modulating the functions of chromatin remodeling transcriptional regulators, such as the polycomb-group protein EZH2 (Eisen et al. 1995). Patients with ATRX syndrome have severely compromised genetic defects due to mutated ATRX gene.

Rubinstein–Taybi syndrome (RTS) is another example of a neurological disorder associated with the dysfunction of HAT. The majority of the Rubinstein–Taybi cases are associated with mutations in the CBP gene located at chromosome 16p13.3 and some in the E1A-binding protein p300 (EP300) gene at chromosome 22q13.2 (Lonard & O’Malley 2012). In 1963, Jack Herbert Rubinstein and Hooshang Taybi described a series of cases with this syndrome demonstrating some typical features that include mental disability, distinctive facial features, and broad thumbs and toes, and are often associated with cryptorchidism in males. This disease is rare and approximately one out of 100 000–125 000 children are born with this disorder. CBP is a transcriptional coactivator that has intrinsic HAT activity and binds to the transcription factor cAMP response element-binding protein (CREB) to regulate gene transcription (Park et al. 2014). Mutation or deletion in the CBP gene severely affects HAT activity of CBP and the ability of CBP to transactivate CREB, indicating that loss of the HAT activity of CBP may cause RTS.

In Huntington’s disease, transcriptional coactivator PGC1α (PPARGC1A) expression is severely impaired, and mouse genetic studies revealed that loss of PGC1α severely impairs metabolism and accentuates neurodegeneration. Huntington’s disease is an autosomal dominant disorder characterized by impaired muscle coordination that leads to cognitive malfunctioning and psychiatric problems. PGC1α is a potent suppressor of reactive oxygen species (ROS) by activating the transcription of ROS defense enzymes such as superoxide dismutase 1 (SOD1), manganese SOD (SOD2), catalase, and glutathione peroxidase (Chaturvedi et al. 2009). In absence of PGC1α coactivator, the neuronal cells are extremely sensitive and vulnerable to neurotoxins leading to apoptotic death of neuronal cells and oxidative damage in the brain.

Studies using SRC knockout animals identified important roles for nuclear receptor coactivators in the coordination of neurobehavioral functions and brain development. SRC-1 is ubiquitously expressed in
the human brain with more prominent presence in hippocampus, olfactory bulbs, and cortex (Meijer et al. 2000). SRC-1 is a crucial regulator of sexually dimorphic regions in the brain and coactivates GR functions to coordinate the hypothalamic–pituitary–adrenal axis of the brain. Neurobehavioral tests on Src-1−/− animals compared with WT littermates discovered some novel roles of SRC-1 in anxiety response (Stashi et al. 2013). In comparison, Src-2−/− females displayed decreased anxiety responses under certain environmental stimuli, whereas...
males were found to have deficits in sensorimotor gating, a neurological process which is important to understand the functional significance of attentional abnormalities. By contrast, Src-3−/− males were devoid of any noticeable neurological abnormalities; however, the females exhibit reduced exploratory activities and increased anxiety behavior (Stashi et al. 2013). Collectively, these findings establish the role of SRCs in the regulation of the CNS and coordination of neurobehavioral phenotypes in a gender-specific manner.

Cardiac development and disease

Transcriptional coactivators can play an essential role in cardiac development by regulating the mitochondrial response of the heart by broadly regulating gene expression from both nuclear and mitochondrial genomes. PGC1 has been extensively studied with respect to cardiac development and bioenergetics of the heart, and its expression was found to be repressed in numerous models of heart failure with a maladaptive energetic profile (Rowe et al. 2010). PGC1α induces expression of numerous genes in cardiac cells regulating major metabolic pathways to maintain a steady supply of ATP production. Genes induced by PGC1α include the majority of mitochondrial respiratory subunits, ATPase complexes, enzymes of fatty acid biosynthesis and transport, and key enzymes of the glycolytic and tricarboxylic acid cycle (TCA; Banke et al. 2010). In addition to metabolic pathways, PGC1α induces angiogenesis in myocytes by directly activating a broad range of angiogenic factors including vascular endothelial growth factor (VEGF) independent of the hypoxia-inducible factor pathway (Arany et al. 2008). Overexpressing PGC1α in the heart identified univocal roles of the coactivator in mitochondrial biogenesis (Lehman et al. 2000). PGC1α activates both mitochondrial as well as nuclear genes by directly transactivating transcription factors such as nuclear respiratory factor (NRF) and estrogen-related receptor (ERR) (Hock & Kralli 2009). These findings have clearly placed PGC1 as a prime regulator of metabolism in the heart, in both cardiomyocytes and cardiac cells.

In addition to PGC1, expression of coactivator SRC-2 is found to be repressed in failing hearts. Genetic ablation of Src-2 identified activation of a ‘fetal gene program’ in adult mice by altering the expression of metabolic and sarcemeric genes (Reineke et al. 2012). Mechanistically, Src-2 depletion reduces the expression of several transcription factors such as GATA as well as coactivators such as PGC1α, indicating that SRC-2 is a prime regulator of the steady-state adult cardiac transcriptomic profile (Reineke et al. 2012). These studies have deciphered the importance of coactivators in cardiac functioning and how subtle changes in their expression can lead to catastrophic medical conditions.

Inflammatory diseases

The most common lung diseases including asthma, chronic obstructive pulmonary disease (COPD), cystic fibrosis, and acute respiratory distress involve inflammatory responses coordinated by expression of multiple proinflammatory genes. Several transcriptional coactivators have been linked as the molecular regulators of inflammatory responses, of which HDACs deserve special mention (Barnes et al. 2005). Patients with asthma exhibit increased expression of HAT with simultaneous reduction in HDAC1 level in the bronchial and alveolar macrophages compared with normal airways (Cosio et al. 2004). In patients with COPD, there is a significant decrease in HDAC2 expression with a concomitant increase in HAT activity facilitating activation of NFκB and transcription of proinflammatory cytokines (Qu et al. 2013). The alveolar macrophages in COPD patients display increased release of tumor necrosis factor alpha (TNFα) and interleukin 8 (IL8) in response to stimuli, thus contributing to the adversity of the pathology. Traditional therapy includes corticosteroids that effectively suppress the transcription of proinflammatory genes by inhibiting NFκB and AP1 transcription factors (Barnes 2013).

The transcriptional coactivator SRC-3 acts as a protective factor against acute inflammatory response by repressing translation of inflammatory cytokines. Src-3−/− animals are more susceptible to endotoxic shock compared with their WT littermates with enhanced levels of proinflammatory cytokines including TNFα, IL6, and IL1β (Yu et al. 2007). Thus, it is sufficient to conclude that expression of coactivators delicately balances the inflammatory responses by modulating expression of ILs and cytokines.

Metabolic disorders and circadian biology

Coactivators are essential coordinators of whole-body energy homeostasis by modulating the expression of multiple metabolic enzymes. SRC family coactivators are prime regulators of metabolic pathways in different tissues, and genetic deletion of their expression corresponds to various physiological abnormalities and metabolic disorders (Dasgupta et al. 2014). Src-1−/− animals
display reduced energy expenditure with an increased risk of developing obesity as well as a defective gluconeogenic program (Picard et al. 2002, Louet et al. 2010). Moleculerly, SRC-1 coactivates CEBPα (CEBPα; CCAAT enhancer-binding proteins) to promote transcription of regulatory enzymes in the gluconeogenic pathways such as pyruvate carboxylase, phosphoenolpyruvate carboxykinase, and fructose-1,6-bisphosphatase (FBP1) (Picard et al. 2002). By contrast, Src-2<sup>-/-</sup> animals are protected from high-fat diet-induced obesity and exhibit increased insulin sensitivity, higher lipolysis, and reduced fat uptake (Picard et al. 2002). Loss of Src-2<sup>-/-</sup> also affects the hepatic glucose release due to decreased expression of glucose-6-phosphatase simulating the phenotypes observed in the genetic disorder Von Gierke’s disease (Chopra et al. 2008). SRC-2 also stimulates absorption of fatty acids from the gut by activating the expression of bile salt export pump (BSEP (ABCB11)) by coactivating farnesoid X receptor (FXR (NR1H4)) under conditions of reduced energy status, thereby coordinating whole-body energy homeostasis (Chopra et al. 2011). Even in tumor cells, SRC-2 was found to modulate fatty acid biosynthesis by distinct reprogramming of metabolic functions (Dasgupta et al. 2012a). By contrast, SRC-3 participates in white adipocyte development and supports fatty acid metabolism in skeletal muscle by regulating the expression of the long-chain fatty acid transporter carnitine/acylcarnitine translocase (CACT (SLC25A20); York et al. 2012). Thus, alterations in the expression of SRCs promote global changes in numerous metabolic pathways in different tissues (York et al. 2013) to maintain the energy demands of our body, and genetic loss of their expression can lead to severe metabolic disorders (York & O’Malley 2010).

In light of this knowledge, recent studies have indicated the importance of transcriptional coactivators in circadian biology. Our recent findings have indicated that SRC-2 is a prime coordinator of circadian activities by regulating the expression of genes that regulate hepatic metabolism and diurnal rhythmicity (Stashi et al. 2014). Moleculerly, SRC-2 coactivates transcription factors brain and muscle ARNT-like 1 (BMAL1/ARNTL) and circadian locomotor output cycles kaput (CLOCK), the two core components of the clock machinery (Asher & Schibler 2011). Cistromic analyses revealed that recruitment of SRC-2 to the genome overlaps with BMAL1 during the light phase targeting expression of core metabolic genes and circadian regulators. In addition, metabolomic profiling of liver metabolites from Src-2<sup>-/-</sup> and WT littermates identified severe alterations in core metabolic pathways including glycolysis, TCA, and fatty acid biosynthesis (Stashi et al. 2014). Collectively, these findings uncovered the key role of the transcriptional coactivator SRC-2 in circadian biology and its impact on various metabolic processes.

**Coactivators as targets for cancer therapy**

Several coactivators including PGC1, SRC family members, p300/CBP have been found to be either amplified or overexpressed in different types of cancer (Xu et al. 2009). SRCs play important roles in endocrine-related cancers such as breast, prostate, ovarian, and endometrial cancers (Lonard & O’Malley 2012) and their functions in other types of cancer are rapidly being decoded (Fig. 3). SRC-1 and SRC-3 promote ER-dependent breast cancer proliferation, as well as facilitate cancer metastasis by upregulating transcription of invasive gene signature coactivating polyoma enhancer activator 3 (PEA3 (ETV4); Qin et al. 2009, 2011). SRC-1 and SRC-3 are overexpressed in endocrine-resistant tumors such as aromatase inhibitor-resistant and tamoxifen-resistant tumors (McBryan et al. 2012). In prostate cancer, deep sequencing studies revealed SRC-2 amplification in 8% of primary tumors and 37% of metastatic tumors (Taylor et al. 2010). In addition, SRC-2 expression correlates positively with the poor survival of prostate cancer patients (Agoulnik et al. 2006, Agoulnik & Weigel 2008), and its expression is an important predictor of time-to-disease relapse (Dasgupta et al. 2012b). Recent studies have identified coactivators such as SRC-1, SRC-3, and PGC1α as regulators of bioenergetic pathways in cancer cells (Vazquez et al. 2013, Motamed et al. 2014, Zhao et al. 2014). PGC1α promotes mitochondrial oxidative phosphorylation to generate sufficient energy supporting the anabolic needs of tumor cells. In addition, recent findings have indicated that coactivators such as p300/CBP along with SRC-3 play critical roles to maintain pluripotency and an embryo stem cell state (Percharde et al. 2012, Wu et al. 2012, Chitilian et al. 2014). SRC-3 coactivates estrogen-related receptor beta (ESRRB) to enhance the expression of Oct4 (Pou5f1), Sox2, and Nanog, the master drivers of stem-cellness. Thus, it will be important to understand the role of these coactivators in ‘cancer stem cells’.

As SRCs have emerged as ‘master regulators’ of cancer progression and metastasis by integrating various upstream signaling pathways, therapeutic targeting of these molecules may be beneficial for treatment of cancers. High-throughput screening of a chemical library containing compounds from the NIH–Molecular Libraries Probe Production Centers Network (MLPCN) was used to identify the
Figure 3

Graphical representation of the percentage of copy number alteration (CNA) frequency of steroid receptor coactivators (SRC-1, SRC-2, and SRC-3) across different types of cancer. Data represent various types of alterations including gene amplification, mutation, and deletion. Data generated using TCGA datasets from cBioPortal (Cerami et al. 2012, Gao et al. 2013).
inhibitors blocking the intrinsic transcriptional activity of SRCs (Wang et al. 2014). The study identified a cardiac glycoside bufalin as a potent small-molecule inhibitor (SMI) for SRC-3 and SRC-1. Molecularly, bufalin and digoxin (a cardiac glycoside) blocked SRC-3 expression by directly binding to it and promoting its rapid degradation in a proteasome-dependent manner. Bufalin was extremely potent in the nanomolar scale to block the growth and proliferation of breast and lung cancer cells (Wang et al. 2014). In addition, Verrucarin A was also identified as a SMI that can selectively promote the degradation of SRC-3 protein, while affecting SRC-1 and SRC-2 to a lesser extent but having no impact on CARM1 and p300 protein levels. Verrucarin A belongs to a group of sesquiterpene found in toxins of pathogenic fungus and has potent anticancer effects by blocking tumor cell growth, proliferation, and migration–invasion (Yan et al. 2014). Thus, targeting the coactivators represents a novel way to block tumor cell growth, and future studies should identify effective small-molecule inhibitors to circumvent other pathologies as well.

Conclusion

Transcriptional coactivators have emerged as an important new class of functional proteins that participate with virtually all transcription factors and NRs to intricately regulate gene expression in response to a wide variety of environmental cues. Recent findings have highlighted that coactivators are important for almost all biological functions. Coactivators work in tandem with specific interacting partners to precisely regulate activation of genes, and loss or genetic defects lead to severe pathologies. Future studies will further broaden our understanding about these fascinating molecules in their various biological functions, and drug discovery efforts targeting coactivators may prove valuable for treatment of a variety of diseases.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the review.

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Author contribution statement

Both authors contributed equally to all aspects of the article.

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