Urocortin-dependent effects on adrenal morphology, growth, and expression of steroidogenic enzymes in vivo

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Abstract

Urocortin (UCN) 1, 2, and 3 are members of the corticotropin-releasing factor (CRF) family that display varying affinities to the CRF receptor 1 (CRFR1 (CRHR1)) and 2 (CRFR2 (CRHR2)). UCNs represent important modulators of stress responses and are involved in the control of anxiety and related disorders. In addition to the CNS, UCNs and CRFRs are highly expressed in several tissues including the adrenal gland, indicating the presence of UCN-dependent regulatory mechanisms in these peripheral organ systems. Using knockout (KO) mouse models lacking single or multiple Ucn genes, we examined the potential role of the three different Ucn genes on morphology and function of the adrenal gland. Adrenal morphology was investigated, organ size, cell size, and number were quantified, and growth kinetics were studied by proliferative cell nuclear antigen staining and Ccn1 expression analysis. Furthermore, mRNA expression of enzymes involved in steroidogenesis and catecholamine synthesis was quantified by real-time PCR. Following this approach, Ucn2, Ucn1/Ucn2 dKO and Ucn1/Ucn2/Ucn3 tKO animals showed a significant cellular hypertrophy of the adrenal cortex and an increase in Ccn1 expression, whereas in all other genotypes, no changes were observable in comparison to age-matched controls. For steroidogenesis, Ucn2/Ucn3 dKO animals displayed the most pronounced changes, with significant increases in all investigated enzymes, providing indirect evidence for increased stress behavior. Taken together, these data suggest that mainly Ucn2 and Ucn3 could be involved in adrenal stress response regulation while Ucn2 additionally appears to play a role in morphology and growth of the adrenal gland.

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Introduction

Corticotropin-releasing factor (CRF) was the first identified member of the CRF family and has been determined as the main mediator for stress response-dependent activation of the hypothalamus–pituitary–adrenal (HPA) axis (Vale et al. 1981, 1983). The neuropeptides urocoritin 1 (UCN1), UCN2 (stress-copin-related peptide), and UCN3 (stresscopin) have been described more recently as additional members of the CRF family. Ucn1 was first reported in 1995 in the rat midbrain region (Vaughan et al. 1995), while Ucn2 and Ucn3 were identified on the basis of sequence homology searching tools not until 2001 (Reyes et al. 2001). CRF, UCNs, and their receptors are involved in homeostatic mechanisms of the stress response and in the control of anxiety, cognitive functions, appetite, and related disorders (Fekete & Zorrilla 2007).

The effects of CRF and UCNs are mediated via two G-coupled receptors, CRFR1 (CRHR1) and CRFR2 (CRHR2), which share 71% amino acid sequence similarity (Grigoriadis et al. 1996). CRFR1 and CRFR2 signal largely by coupling to Gs, leading to the stimulation of adenyl cyclase and activation of protein kinase A. However, stimulation of specific CRFRs in distinct tissues or cell types by receptor-selective CRF peptides has been demonstrated to activate a variety of further signaling pathways (Brar et al. 2002, Bale 2005). UCN1 has similar binding affinity to CRFR1 and CRFR2 while UCN2 and UCN3 are considered as specific ligands for CRFR2 (Fukuda et al. 2005). These specific binding properties translate into distinct phenotypic changes induced by targeted deletions of the CRF receptors. Animals deficient for CRFR1 have an impaired stress response (Smith et al. 1998, Timpl et al. 1998) and decreased anxiety-like behavior (Smith et al. 1998) and fail to display the characteristic HPA axis response to restraint stress. In contrast, Csf2-mutant mice display increased anxiety-like behavior and are hypersensitive to stressors (Bale et al. 2000, Coste et al. 2000).

In addition to the well-characterized effects of Ucns in the CNS, over the last few years, a number of peripheral sites of action have been identified.
Ucn1 expression has been observed in adipocytes (Seres et al. 2004), heart (Nishikimi et al. 2000), skin, and immunological tissue including the thymus and spleen, among others (Baigent 2001). Ucn2 is highly expressed in skin and skeletal muscle, and is differently regulated by circulating glucocorticoid levels, as manipulated by exogenous administration or adrenalectomy (Chen et al. 2004). Ucn3 gene expression can be detected in pancreatic β cells, where it stimulates insulin and glucagon secretion (Li et al. 2003), and in the adipose tissue, heart, and skin. CRFR2 is broadly distributed in peripheral tissues including the heart and skeletal muscle (Dautzenberg & Hauger 2002, Suda et al. 2004). In addition to the expression of CRFR1 in different sites of the brain (Justice et al. 2008), Csf1r expression could be documented in many peripheral tissues such as the testis, endocrine pancreas, or colon (Tao et al. 2007, Huising et al. 2010, O’Malley et al. 2010). Similar to other peripheral organs, members of the CRF/UCN system are also present in the adrenal gland, with Ucn2 and Ucn3 as well as CRFR2 being predominantly expressed in the adrenal cortex, while Ucn1 and CRFR1 are mainly detectable in the adrenal medulla, and CRF is distributed throughout the adrenal gland. A proposed intra-adrenal UCN-dependent regulatory system would thus be dependent on the balance between the local concentration of CRF and UCNs and the presence of their receptors. Following this notion, it has been suggested that a CRFR1 receptor/ligand system might be active in the medulla, while a parallel CRFR2 receptor/ligand system might modulate adrenocortical function (Tsatsanis et al. 2007). Some examples provide evidence for a functional significance of a local UCN system within the adrenal gland: human fetal adrenal cells are able to respond to CRF stimulation through activation of CRFR1 with an increase in steroidogenesis, which has been hypothesized to be required for the late gestational increase in fetal adrenal cortisol and DHEAS production (Sirianni et al. 2005). In addition, preliminary data suggest a stress-related upregulation of Ucn2 in the adrenal gland (Liu et al. 2008).

However, the physiological role of the influence of Ucns on adrenal function and structure has not been investigated in depth so far. Thus, our approach was to investigate the effects of the three Ucns on the adrenal gland using six different Ucn-knockout (KO) mouse models: individual KO for each of the three peptides genes (Ucn1 KO, Ucn2 KO, and Ucn3 KO) as well as combined deletions of two (Ucn1/Ucn2 KO and Ucn2/Ucn3 dKO) or all the three Ucns (Ucn1/Ucn2/Ucn3 KO). To achieve this goal, we investigated the mRNA expression of enzymes involved in steroidogenesis and catecholamine production as well as the histological appearance of the adrenal gland of those mouse models.

Materials and methods

Animals and housing conditions

For the animal experiments, six different genotypes of male Ucn KO mice were used. Mouse breeding took place at the Weizmann Institute of Science, Rehovot, Israel. Mice lacking two or all the three Ucn genes (Ucn1, Ucn2, and Ucn3) were generated by cross-breeding of Ucn1, Ucn2, and Ucn3 single KO mice, which has been reported earlier (Vetter et al. 2002, Chen et al. 2006, Neufeld-Cohen et al. 2010a,b). All mice were on a mixed C57BL/6×129 background. WT mice of the mixed C57BL/6×129 background were derived from the same breeding colony. Male mice that were used in this study were housed up to five mice per cage on a 12 h light:12 h darkness photoperiod (lights on at 1800 h) with food and water ad libitum. All experimental protocols were approved by the Institutional Animal Care and Use Committee of The Weizmann Institute of Science.

To exclude interferences due to the female estrous cycle, as documented earlier (Chen et al. 2006, Xu et al. 2006), only male mice were investigated. Ucn1 KO (n=10), Ucn2 KO (n=9), Ucn3 KO (n=9), Ucn1/Ucn2 dKO (n=8), Ucn2/Ucn3 dKO (n=11), Ucn1/Ucn2/Ucn3 tKO (n=8), and WT controls (n=11) were killed at the age of 3 months.

Morphometry and immunohistochemistry

Adrenal glands from all investigated genotypes were rapidly dissected and placed in 4% paraformaldehyde overnight. Tissues were dehydrated, embedded in paraffin, sectioned, and stained with hematoxylin and eosin (H&E) following standard protocols. H&E-stained adrenal sections were examined using a light microscope with a magnification of 40× and 400×. Areas of medulla and cortex were quantified on four independent sections per mouse from the center of the adrenal gland using the SpotAdvanced Software (Spot, Burroughs, Sterling Heights, MI, USA). The medullary area was normalized for the cortical area and expressed as the medullary/cortical area ratio. Cell nuclei within the outer and inner adrenal cortex were counted on three independent sections per animal under standardized conditions. A decrease in cell size was defined as cellular hypotrophy, whereas a decrease in nuclear density was interpreted as evidence of cellular hypertrrophy, as the amount of extracellular matrix was comparable between the lines. In the adrenal cortex, less cytoplasm can be regarded as a rough estimate of lower functional activity of the cells. Cell counts were expressed as cell number/high power field (ncl/hpf). For proliferative cell nuclear antigen (PCNA) immunohistochemistry (IHC), paraffin-embedded sections
were rehydrated, blocked with 0.3% H2O2 in methanol for 10 min, and incubated with blocking buffer for 15 min. PCNA was immunolocalized overnight at 4°C by means of a rabbit polyclonal antibody (Santa Cruz Biotechnology, Santa Cruz, CA, USA) in a dilution of 1:100 in blocking buffer containing 3% BSA (Roche Applied Science), 5% goat serum (Jackson ImmunoResearch Laboratories, West Grove, PA, USA), and 0.5% Tween 20. After rinsing for 15 min in PBS, secondary antibody (goat-anti-rabbit-biotinylated IgG (Vector Laboratories, Burlingame, CA, USA)) in a dilution of 1:200 in blocking buffer was applied for 30 min at room temperature. For the visualization of the bound PCNA antibody, Vectastain Elite ABC system (Vector Laboratories) and Sigma Fast diaminobenzidine (Sigma) were used.

Quantitative real-time PCR

Adrenal glands from mice of all investigated genotypes were immediately cleansed of adjacent tissue under a stereo microscope, snap frozen in liquid nitrogen, and stored at −80°C. Both adrenals from each individual animal were combined and homogenized in extraction buffer while still frozen. RNA was extracted using the SV Total RNA Isolation System according to the manufacturer’s instructions (Promega). RNA quality was verified on a 1% agarose gel. For cDNA synthesis, 500 ng total RNA were reverse transcribed using the reverse transcription system (Promega).

Gene expression of selected genes involved in steroidogenesis, catecholamine production, and cell cycle regulation was investigated: steroidogenic acute regulatory protein (Star), cholesterol side-chain cleavage enzyme (Cyp11a1), 11-b-hydroxylase (Cyp11b1), aldosterone synthase (Cyp11b2), tyrosine hydroxylase (Th), phenylethanolamine N-methyltransferase (Pnmt), and cyclin D1 (CcnD1). β-Actin was used as housekeeping gene. For primer sequences, see Table 1.

Quantification of β-actin, Star, Cyp11a1, and Cyp11b2 was accomplished for all genotypes using the FastStart DNA MasterPlus SYBR Green I reaction mix in the LightCycler 1.5 (Roche). Real-time PCR conditions in the LightCycler were preincubation at 95°C for 10 min followed by amplification of 40 cycles at 95°C for 10 s, the annealing temperature (primer dependent, see Table 1) for 5 s, and extension at 72°C, at which the time is calculated by the product length in base pairs divided by 25. The melting curve analysis was performed between 65 and 95°C (0.1°C/s). For the quantification of Cyp11b1, Th, Pnmt, and CcnD1, the Sso Fast EVA Green Supermix (Bio-Rad Laboratories) in the Mx3000P QPCR System (Stratagene, La Jolla, CA, USA) was used. Real-time PCR conditions were preincubation at 95°C for 10 min followed by amplification of 40 cycles at 95°C for 10 s, the annealing temperature for 5 s (primer dependent, see Table 1), and extension at 72°C. The melting curve analysis was performed between 65 and 95°C (0.1°C/s).

Furthermore, all products were run on a 1% agarose gel to verify the correct size of the amplified product. Quantification was adjusted using the housekeeping gene β-actin. To facilitate overall comparison of individual real-time experiments, expression levels of the particular genes were set as 100% for WT animals.

Statistical analysis

All results are expressed as mean ± s.e.m. Statistical significance was determined using the Mann–Whitney U test with the Prism 3.02 Software (GraphPad Software, La Jolla, CA, USA). Statistical significance was defined as P<0.05.

Table 1 Primer sequences

<table>
<thead>
<tr>
<th>Gene</th>
<th>Primer sequences</th>
<th>Annealing temperature (°C)</th>
<th>Nucleotides</th>
</tr>
</thead>
<tbody>
<tr>
<td>β-actin</td>
<td><strong>F</strong>: TCATGAAGTGTACGTGAGCATCC <strong>R</strong>: CCTAGAAGCATTGTCGGTGGACGATG</td>
<td>60</td>
<td>911–1195</td>
</tr>
<tr>
<td>Star</td>
<td><strong>F</strong>: GACCTCTGAAAGGCTACAGGGACAGACATCC <strong>R</strong>: TAGCTGAAAGATGGGGAAGACAGTTTC</td>
<td>60</td>
<td>7–987</td>
</tr>
<tr>
<td>Cyp11a1</td>
<td><strong>F</strong>: AGAAGCTTTCCCTGCGGT <strong>R</strong>: GCACCTGCAATATGTTGG</td>
<td>53</td>
<td>34–980</td>
</tr>
<tr>
<td>Cyp11b2</td>
<td><strong>F</strong>: CAATGTGCCAGCTCTACAGACGT <strong>R</strong>: ACAGAGACCTTCCAGAGACCT</td>
<td>63</td>
<td>268–636</td>
</tr>
<tr>
<td>Cyp11b1</td>
<td><strong>F</strong>: GTGCTGCGCCACGCTGCTG <strong>R</strong>: ACCAACAGATGGACTCTGCTG</td>
<td>63</td>
<td>878–1171</td>
</tr>
<tr>
<td>Th</td>
<td><strong>F</strong>: TGCAGGCGCTGTCTG TTTTCCCTTAT <strong>R</strong>: TTCAACAGTGGATTG</td>
<td>60</td>
<td>1175–1381</td>
</tr>
<tr>
<td>Pnmt</td>
<td><strong>F</strong>: GTGAAACGAGTATCTGTGCTG <strong>R</strong>: AAGATGGCATTCCAGACATAC</td>
<td>484–851</td>
<td></td>
</tr>
<tr>
<td>CcnD1</td>
<td><strong>F</strong>: AGCGAGACCATCCCGCAAGCAT <strong>R</strong>: AAGTGCGTGGTGGGCTGCA</td>
<td>60</td>
<td>756–927</td>
</tr>
</tbody>
</table>

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**Results**

**Ucn1 KO**

The area of the adrenal gland and the cortical/medul-
lar area ratio showed no differences between *Ucn1*
KO mice and WT controls. Morphometrical studies
of the outer adrenal cortex displayed a significant
increase in nuclei per hpf (ncl/hpf) in *Ucn1* KO mice in
comparison with age-matched WT controls, indicating
cellular hypotrophy. In the inner adrenal cortex, no
structural differences in comparison with WT animals
could be observed. On the molecular level, *Ucn1* KO
mice displayed a significant decrease in *Cyp11b2*
expression, but there was no significant differences in
the expression of *Star*, *Cyp11a1*, or *Cyp11b1*. Investig-
ation of the expression pattern of enzymes of the
adrenal medulla, involved in catecholamine synthesis,
showed a significantly lower *Th* and *Pnmt* expression in
*Ucn1* KO mice (Fig. 1). Cell proliferation, assessed by
IHC with PCNA, did not demonstrate any differences in
the staining intensity between *Ucn1* KO and WT mice.
Cell cycle activation was quantified with *Ccnd1*
expression, but again no changes could be documented
in the *Ucn1* KO animals (Fig. 2).

**Ucn2 KO**

*Ucn2* KO mice had a significantly higher cell number
per hpf in the adrenal cortex in comparison with
WT animals. In contrast to the cellular hypotrophy,
no changes in the area of the adrenal gland or the ratio
between cortex and medulla could be found. On the
molecular level, a pronounced upregulation of *Cyp11a1*
expression in *Ucn2* KO animals was observed. In
contrast, expression levels of *Star*, *Cyp11b1*, and
*Cyp11b2* remained unchanged. As for the adrenal

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Figure 1 Gene expression levels of enzymes involved in steroidogenesis: *Star* (A), *Cyp11a1* (B), *Cyp11b1* (C), and *Cyp11b2* (D) and in catecholamine synthesis: *Th* (E) and *Pnmt* (F) in the adrenal gland of *Ucn1* KO, *Ucn2* KO, *Ucn3* KO, *Ucn1/Ucn2* dKO, *Ucn2/Ucn3* dKO, and *Ucn1/Ucn2/Ucn3* KO mice. Asterisks denote significant changes for each gene over WT expression levels that are set as 100% for each gene.
medulla, Pnmt expression was found to be significantly lower in Ucn2 KO mice, whereas Th expression did not differ significantly between the two investigated groups showing, though, the same tendency (Fig. 1). Cell proliferation assessed by Ccnd1 expression remained unchanged in this KO model (Fig. 2).

**Ucn3 KO**

Analysis of the structure of adrenal cortices of Ucn3 KO mice displayed a higher number of cells per hpf in the outer adrenal cortex in comparison with WT animals. In contrast, the cell number of the inner adrenal cortex was comparable between the two groups as well as the size of the adrenal and the cortical/medullary area ratio. Similarly, expression levels of Star, Cyp11b1, and Cyp11b2 did not differ between Ucn3 KO and WT mice. However, elevated levels of Cyp11a1 expression in Ucn3 KO animals could be observed. Th and Pnmt expression did not differ between Ucn3 KO and WT mice (Fig. 1). Ccnd1 levels showed a trend toward reduced expression, but PCNA staining did not reveal any changes in the proliferation rate in Ucn3 KO animals (Fig. 2).

**Ucn1/Ucn2 dKO**

Ucn1/Ucn2 dKO mice displayed no differences in the area of the adrenal gland and in cortical/medullary
Ucn1/Ucn2 dKO mice had a significantly higher cell number per hpf in the outer and inner adrenal cortex, indicating cellular hypotrophy. Significantly lower levels of expression for Star and Cyp11b2 were found in adrenals from Ucn1/Ucn2 dKO mice in comparison with WT controls. In contrast, Cyp11a1 and Cyp11b1 expressions were comparable between the two groups, with no significant difference between WT and Ucn1/Ucn2 dKO animals. Both Th and Pnmt expression levels, involved in catecholamine production, showed no differences in the Ucn1/Ucn2 dKO mice (Fig. 1). Cell proliferation proved to be significantly higher in Ucn1/Ucn2 dKO mice. Likewise, IHC with PCNA presented an increased staining in both the medulla and cortex of those mice (Fig. 2).

**Ucn2/Ucn3 dKO**

Mice lacking both Ucn2 and Ucn3 displayed no significant differences in morphological studies of the adrenal cortex by counting the cell number per hpf in comparison with WT controls. However, Ucn2/Ucn3 dKO mice displayed smaller adrenal glands with an unchanged ratio of the cortex to medulla area in comparison with WT animals. In contrast, significant upregulation of Star, Cyp11a1, and Cyp11b1 could be shown. Cyp11b2 levels did not differ between Ucn2/Ucn3 dKO mice and WT controls. Ucn2/Ucn3 dKO mice presented a significant upregulation of the expression levels of enzymes catalyzing catecholamine synthesis (Fig. 1). No changes in the Ccnd1 levels could be observed in this investigated group, similarly as after PCNA IHC staining (Fig. 2).

**Ucn1/Ucn2/Ucn3 tKO**

Following morphometrical analyses, adrenals from tKO mice had a significantly higher cell number per hpf in the inner and outer adrenal cortex. Besides the cellular hypotrophy, a reduced surface of the adrenal gland could be found in comparison with WT controls. However, the medulla/cortex area ratio displayed no differences between triple KO and WT mice. Significantly lower expression levels of Star and Cyp11b1 in the adrenal were evident in tKO animals when compared with WT controls while differences in Cyp11a1 and Cyp11b2 displayed the same trend but failed to reach statistical significance. Adrenal medulla enzymes showed no changes in their expression levels between tKO mice and WT controls (Fig. 1). Adrenal glands of tKO animals displayed a significant increase in the cell cycle protein CCND1. Accordingly, an increased IHC staining with PCNA could be documented in the adrenals of those animals (Fig. 2).

**Discussion**

To evaluate the impact of the three different UCNs on the adrenal gland in an in vivo situation, we made use of six UCN KO models and investigated the effect of these targeted deletions on adrenal structure and function. Following this morphological and molecular approach, it became evident that absence of specific UCNs results in a complex pattern of functional changes in the adrenal gland, which would not have been predicted in all instances.

Ucn1 and Ucn2 have been implicated in fluid and sodium regulation and modulation of cardiac function during heart failure. Specifically, administration of Ucn1 in a sheep model with acute heart failure has been reported to result in decreasing levels of plasma vasopressin, renin activity, as well as aldosterone and natriuretic peptide levels (Rademaker et al. 2002), which was associated with delayed development of overt heart failure (Rademaker et al. 2007). Similarly, Ucn2 was shown to reverse furosemide-induced increases in plasma renin activity and induced greater decreases in plasma aldosterone and vasopressin (Rademaker et al. 2009). While these effects involve multiple systems including cardiac and vascular function, there is only limited information on UCN-dependent adrenocortical aldosterone secretion. Interestingly, in vitro treatment of the human adrenocortical cell line NCIh295 with Ucn1 has been demonstrated to induce and increase in Cyp11b2 expression and aldosterone secretion (Kageyama et al. 2010). In line with this finding in our KO model, lack of Ucn1 and Ucn2 alone or in combination was associated with cellular hypotrophy of the outer adrenal cortex and lower expression levels of Cyp11b2. Thus, while the interventional protocols using Ucn1 and Ucn2 in models of heart failure demonstrate an inhibition of several volume-retaining hormones including the renin–angiotensin aldosterone system, direct effects on aldosterone secretion from the adrenal cortex cannot be excluded and might impact on long-term effects of these peptides.

In previous studies, it has been described that Ucn1 KO mice display normal hormonal responses to acute stress (Vetter et al. 2002, Wang et al. 2002). In accordance with these findings, we demonstrated that the structure of the inner adrenal cortex as well as the expression levels of the enzymes involved in corticosterone and catecholamine synthesis remained unchanged. As Crfr1 KO mice have been reported to have an atrophic adrenal medulla and lower epinephrine plasma levels (Yoshida-Hiroi et al. 2002), it is prudent to assume that functional redundancy through retained CRF-dependent stimulation of the CRFR1 could be the reason for the observed minor effects on zona fasciculata function but not for the significant effects in adrenomedullary function in Ucn1 KO animals.
In previous experiments, we could demonstrate that female, but not male, Ucn2 KO mice exhibit an increase in the basal daily rhythm of ACTH and corticosterone (Chen et al. 2006). These findings are endorsed by our present studies in which the absence of Ucn2 caused cellular hypertrophy of the adrenal cortex in male animals with only minor differences in the expression of steroidogenic enzymes and enzymes involved in catecholamine synthesis. Almost identically, in Ucn3 KO mice, no or only small differences regarding adrenal structure, steroidogenesis, and epinephrine synthesis were detectable, a finding that also correlates with earlier data demonstrating normal corticosterone values in Ucn3 KO animals (Deussing et al. 2010). Similar to the situation in single Ucn1 KO animals, it is well possible that the isolated lack of one of both UCNs acting via the CRFR2 might be compensated by the other, resulting in a rather mild phenotype in single Ucn2 and Ucn3 KO mice.

In line with this notion and in contrast to the isolated KO models, mice lacking both Ucn2 and Ucn3 showed a robust upregulation of all investigated enzymes of steroidogenesis and catecholamine synthesis and displayed smaller adrenal glands, whereas no structural differences on cellular level or on proliferation could be observed. Accordingly, the increase in Star expression in Ucn2/Ucn3 dKO animals could be explained by the dominance of the stress phenotype of Ucn1 peptide in this model leading to increased steroidogenesis. Similarly, in each of the Ucn2 and Ucn3 KO and more pronounced in the Ucn2/Ucn3 dKO model, the dominance of the Ucn1 peptide was associated with an increase in Cyp11a1. For Cyp11b1, although the expression data were not significantly different in the single KO animals as expected, its expression was significantly upregulated in Ucn2/Ucn3 dKO mice and on the other hand significantly downregulated in Ucn1/Ucn2/Ucn3 tKO animals. Consequently, Ucn1 appears not to contribute to a rescue of the stress-copying phenotype, as described earlier, although it is known that it has a similar affinity to both CRF receptors. It has been previously shown that Crfr2 KO mice display normal baseline concentrations of ACTH and corticosterone but increased sensitivity to stress (Bale et al. 2000, Coste et al. 2000, 2006), supporting the hypothesis that stimulation of CRFR2 results in a decrease in anxiety and to a stress-copying behavior (Hsu & Hsueh 2001).

In Ucn1/Ucn2/Ucn3 tKO mice, a decrease in adrenal size, cellular hypertrophy of the adrenal cortex, reduced steroidogenic capacity, and increased proliferation rate were observable, indicating that absence of Ucn1 outweighs the lack of Ucn2 and Ucn3. However, Ucn1 is mainly expressed in the adrenal medulla and functions via binding to its receptors CRFR1 and CRFR2, which are mainly located in the adrenal cortex. A possible model of indirect or direct interaction between the adrenal cortex and the medulla has been proposed, for example, by Fukuda et al. (2005). Comparable to this phenotype is the Crfr1/2 KO model described previously, where an atrophy of the zona fasciculata and a phenotype of glucocorticoid insufficiency could be documented (Preil et al. 2001). Apparently, absence of either all three UCNs or both their receptors from the adrenal gland leads to a similar phenotypic development of the organ structure. Concordant with the reduced levels of steroidogenic enzymes in triple KO animals is the observation that Ucn1/Ucn2/Ucn3 tKO mice show a tendency toward reduced corticosterone levels under baseline conditions (Neufeld-Cohen et al. 2010b), although the adrenal insufficiency appears to be less pronounced in this model in comparison with CRF1/2 KO animals.

Finally, in Ucn1/Ucn2 dKO mice, significant downregulation of key enzymes of steroidogenesis was detectable, together with a significant cellular hypertrophy of the adrenal cortex and an increased proliferation rate. As this observation is not in absolute agreement with a recent study, in which older animals had been investigated (Neufeld-Cohen et al. 2010a), an explanation for this discrepancy could be that UCN-mediated effects display age dependency. While a number of parameters affecting adrenal formation and zonation have been found to be age dependent (Bielohuby et al. 2007, Hershkovitz et al. 2007), UCN-related effects on the adrenal glands will require further detailed experiments involving time-dependent functional studies. The fact that Ucn1/Ucn2-dependent phenotype is comparable with that of Ucn1/Ucn2/Ucn3 tKO mice, together with the very slight changes observed in Ucn3 single KO mice suggest that the role of Ucn3 on adrenal function and growth is relatively limited. Additionally, as only in Ucn2 KO, Ucn1/Ucn2 dKO, and Ucn1/Ucn2/Ucn3 tKO animals a cellular hypertrophy of the adrenal cortex could be documented, a potential role of Ucn2 in the development and the structure of the adrenal gland can be hypothesized.

Taken together, the absence of UCNs influences both the adrenal cortex and the medulla. As UCN single KO mice showed relatively mild adrenal phenotypes, these findings suggest that regulation of adrenal structure and function is dependent on the balance of all UCNs and their receptors.

Declaration of interest

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.
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