Vitamin A supplementation induces adipose tissue loss through apoptosis in lean but not in obese rats of the WNIN/Ob strain

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Abstract

Vitamin A is a known regulator of adipose tissue growth. In this paper, we report the possible role of dietary vitamin A supplementation in the regulation of adipose tissue mass, using a novel obese rat model of the WNIN/Ob strain developed at the National Centre for Laboratory Animal Sciences of the National Institute of Nutrition, India. Twenty-four male lean and obese rats of the WNIN/Ob strain were broadly divided into two groups at 7 months of age; each group was subdivided into two subgroups consisting of six lean and six obese rats and they were given diets containing either 2.6 mg or 129 mg vitamin A/kg diet for 2 months. Feeding a high but non-toxic dose of vitamin A (129 mg/kg diet) resulted in a significant reduction in the adiposity index and retroperitoneal white adipose tissue (RPWAT) weight in obese rats while a marginal reduction was observed in lean rats. Further, this treatment resulted in a significantly increased RPWAT apoptotic index and Bax protein expression and a decreased expression of Bcl2 in the lean rats. However, no such changes were observed in the RPWAT of the obese rats subjected to identical treatment. Thus, our data suggests that chronic dietary vitamin A supplementation at a high dose effectively regulates adipose tissue mass both in the lean and obese phenotypes of the WNIN/Ob rat strain, perhaps through different mechanisms.

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Introduction

Obesity is a condition where excess energy is stored as triglycerides in white adipose tissue (WAT) (Palou et al. 2000). WAT, the major component of the adipose organ, is widely distributed in the mammalian body in large quantities and serves as a reservoir of energy. On demand, it supplies energy in the form of free fatty acids (FFAs), which are released by a lipolytic process and utilized by various tissues (Frayn et al. 2003). Adipose tissue mass is determined by both size and number of adipocytes. The latter in turn is determined by adipocyte apoptosis and differentiation (Prins & O’Rahilly 1997). Apoptosis, an evolutionarily conserved intracellular pathway, allows the organism to control cell number and tissue size very tightly, and protects it from rogue cells that threaten tissue homeostasis (Reed 2002). Apoptosis has been described both in WAT and brown adipose tissue (BAT) (Prins et al. 1994). Several studies have demonstrated in vitro and in vivo adipocyte apoptosis under various conditions such as growth factor deprivation, dietary conjugated linoleic acid supplementation and streptozotocin administration (Prins et al. 1997, Loftus et al. 1998, Miner et al. 2001, Hargrave et al. 2002). Even in patients with malignancy-associated weight loss, apoptosis is detected in abdominal, subcutaneous and omental fat (Prins et al. 1994). Thus, emerging evidence suggests that adipose tissue is an active organ with continuous recruitment of newer cells and elimination of adipocytes by apoptosis under normal physiological conditions as well as disease conditions.

Vitamin A, an important micronutrient, has an unusually wide range of vital physiological actions in mammals (e.g. morphogenesis, vision, embryonic development, reproduction and immune function etc.) (Villarroya et al. 1999). Although liver is the major organ involved in the storage and homeostasis of retinoids, adipose tissues contain substantial amounts of retinol and retinyl esters, which account for 15–20% of the total body retinoid stores (Tsutsumi et al. 1992). Moreover, adipose tissue is considered to be a potential site for retinoic acid (RA) action by expressing retinoid receptor subfamilies, namely RA receptor (RAR) and retinoid X receptor (RXR) (Bonet et al. 2003). It is very well documented that vitamin A and its active metabolite RA are positive regulators of uncoupling protein-1 (UCP1) (Kumar & Scarpace 1998, Bonet et al. 2003). Studies from brown adipocyte cell cultures and an in vivo system have demonstrated the role of RA as a transcriptional activator of UCP1 gene expression and its implication in...
the control of body adiposity (Puigserver et al. 1996, Bonet et al. 2000). Furthermore, the role of vitamin A and its metabolite status on body fat regulation in a rodent model has also been well documented (Bonet et al. 2003, Felipe et al. 2003).

Many in vitro studies have reported that RA inhibits adipocyte differentiation at a high concentration (Kuri-Harcuch 1982, Hernandez et al. 1993). In addition, vitamin A supplementation at high doses results in decreased adiposity in rats (Kumar et al. 1999). On the contrary, the feeding of a vitamin A-deficient diet to rats resulted in increased adiposity and body weight gain (Bonet et al. 2000, Ribot et al. 2001). Furthermore, all-trans-RA, a vitamin A metabolite, has been shown to be a potent inducer of apoptosis in rat stromal–vascular cells (Kim et al. 2000). All these studies indicate that vitamin A status can modulate the adipose tissue mass in rodents. The present study was therefore undertaken to gain insight into the in vivo modulation of adipose tissue mass by vitamin A, using a novel obese mutant rat model developed at our institute.

Materials and methods

Animals and experimental design

The WNIN/Ob mutant rat strain developed from an 80-year-old Wistar inbred rat stock colony at the National Centre for Laboratory Animal Sciences (NCLAS) of the National Institute of Nutrition (NIN), Hyderabad, India, has three phenotypes, namely lean (+/+), carrier (+/−) and obese (−/−). Rats of the obese phenotype are hyperphagic, hyperinsulinemic, hypertriglyceridemic and hypercholesteremic (Giridharan et al. 1996). In addition, these rats are also characterized by hyperleptinemia (Vajreswari A, Harishanker N and Giridharan NV, unpublished data).

Male, 7-month-old obese rats of the WNIN/Ob strain were obtained from NCLAS and broadly divided into two groups, A and B, each consisting of 12 lean and obese rats and each further divided into two subgroups (A I, A II and B I, B II) consisting of six rats in each subgroup. Subgroups A I and B I received the stock diet, which provided 2.6 mg vitamin A/kg diet, while subgroups A II and B II received a high vitamin A-containing diet (129 mg vitamin A/kg diet) (as retinyl palmitate). The stock diet and the high vitamin A-containing diets are identical with regard to the nature and concentrations of all ingredients except the concentrations of vitamin A. The study was approved by the Institutional Animal Ethical Committee. The animals were maintained on their respective diets for a period of 2 months. Food and water were provided ad libitum. Daily food intake and weekly body weights were recorded.

Rats were housed individually at an ambient temperature of 22·0 ± 1 °C with relative humidity of 50–60% in a 12 h light:12 h darkness cycle and animals were cared for in accordance with the principle of the Guide to the Care and Use of Experimental Animals formulated by the CPC SEA (Committee for the Purpose of Control and Supervision on Experiments on Animals), Government of India. At the end of 2 months, the rats were killed after 12 h fasting. Various adipose tissues were excised, weighed, rapidly frozen in liquid nitrogen and stored at −80 °C until analysis.

Determination of adiposity index

Adiposity index was determined by the sum of the weights of WATs (retroperitoneal, epidydimal, subcutaneous and omental) divided by body weight × 100 (Taylor & Phillips 1996).

Adipose tissue cell density and apoptotic index measurements

Retroperitoneal WATs (RPWATs) were collected from the various experimental groups and fixed in 10% formalin. Tissues were then processed and slides were prepared and stained by hematoxylin and eosin by employing routine histopathological procedures for microscopic examination. To evaluate the adipocyte cell density, fat cell density was measured in a calibrated microscope eyepiece graticule at a uniform magnification of × 250. The number of cells within the marked area was counted and expressed as cells/mm². To evaluate the apoptotic index, adipose tissue apoptotic cells were counted based on the following criteria: (1) fragmented nuclei, (2) 50% smaller in size than normal cells and (3) irregular outer membrane shape, and expressed as per cent (apoptotic index (%)=no. of apoptotic cells/total no. of cells × 100).

DNA fragmentation analysis

RPWAT samples (100 mg) were homogenized in 10 mmol/l Tris–HCl (pH 7·5), 0·32 mol/l sucrose, 5 mmol/l MgCl₂ and 0·5% lauryl sarcosyl containing 200 mg/l proteinase K, and then incubated at 55 °C for 1 h. DNA was subsequently precipitated overnight with ethanol. DNA (20 µl) was loaded onto a 1·5% agarose gel, which was stained with ethidium bromide after gel, which was stained with ethidium bromide after

Western blot analysis of pro- and anti-apoptotic proteins

Retroperitoneal adipose tissue was homogenized with lysis buffer (50 mM Tris (pH 7·5), 150 mM NaCl, 0·02% sodium azide, 1% SDS and 5% protease inhibitor cocktail (0·5% deoxycholate; Sigma Chemical Co.),
and centrifuged at 23,000 g for 1 h. Equal amounts of protein were separated on an SDS-12% polyacrylamide gel and transferred to a nitrocellulose membrane (Hybond-C extra; Amersham Pharmacia Biotech, Amersham, Bucks, UK). Equal loading of protein and transfer were ensured by staining membranes with Ponseau S (Sigma Chemical Co.). Blots were then blocked for 2 h at room temperature with PBS–0·02% Tween-20 containing 5% non-fat dry milk powder prior to incubation with 1:1000 diluted rabbit polyclonal antibodies to Bcl2 and Bax (Santa Cruz Biotechnology Inc., Santa Cruz, CA, USA). Blots were washed and then incubated with goat anti-rabbit IgG antibodies (1:20,000) conjugated with alkaline phosphatase (Sigma). After extensive washing, BCIP/NBT substrate (Sigma) was added, and the intensity of the developed bands was read and quantified by using a scanning densitometer with automatic calibration (GS-710 Imaging Densitometer; Bio-Rad, Hercules, CA, USA).

**Statistical analysis**

Results are expressed as means ± s.e. Statistical significance was determined by two-way ANOVA and the F ratios were considered significant at the $P \leq 0·05$ level.

### Results

#### Effect of high vitamin A supplementation on body weight gain, adiposity index and RPWAT weight

A significant reduction in body weight gain of the high vitamin A-supplemented obese group (B II) compared with the stock diet-fed obese group (B I) was observed without any alteration in their food intake. On the other hand, no such effect was seen in the lean rats receiving high doses of dietary vitamin A (A II) when compared with their lean counterparts maintained on the stock diet with normal levels of vitamin A (2·6 mg vitamin A/kg diet) (A I). Interestingly, this treatment resulted in a significantly decreased adiposity index and RPWAT weight in obese rats supplemented with high doses of vitamin A (B II). However, such effects were not seen in lean rats fed on a high vitamin A-containing diet (A II) (Table 1).

**Table 1** Effect of high vitamin A supplementation on physical parameters. Data represent the means±s.e. of six rats from each group

<table>
<thead>
<tr>
<th>Pretreatment body weight (g)</th>
<th>Post-treatment body weight (g)</th>
<th>Body weight gain (g)</th>
<th>Adiposity index (%)</th>
<th>RPWAT weight (g/100 g body weight)</th>
<th>Daily food intake (g)</th>
<th>F ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>A I 374±13·3a</td>
<td>405±14·1a</td>
<td>31·3±7·1a</td>
<td>6·3±1·7a</td>
<td>2·58±0·64a</td>
<td>14·7±0·7a</td>
<td></td>
</tr>
<tr>
<td>A II 373±10·7a</td>
<td>402±13·9a</td>
<td>29·0±6·6a</td>
<td>3·4±0·51a</td>
<td>1·68±0·38a</td>
<td>15·8±0·6a</td>
<td></td>
</tr>
<tr>
<td>B I 613±35·3b</td>
<td>801±39·9b</td>
<td>187±10·6b</td>
<td>33·6±1·7b</td>
<td>10·3±1·0b</td>
<td>23·6±0·8b</td>
<td></td>
</tr>
<tr>
<td>B II 612±18·3b</td>
<td>723±21·7b</td>
<td>111±18·2c</td>
<td>27·6±1·6c</td>
<td>6·12±0·8c</td>
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F ratio

<table>
<thead>
<tr>
<th>Group</th>
<th>Treatment</th>
<th>Interaction</th>
</tr>
</thead>
<tbody>
<tr>
<td>Group</td>
<td>121·3*</td>
<td>241·7*</td>
</tr>
<tr>
<td>Treatment</td>
<td>0·001</td>
<td>3·09</td>
</tr>
<tr>
<td>Interaction</td>
<td>0·000</td>
<td>2·6</td>
</tr>
</tbody>
</table>

Mean values without a common superscript are significant at $P<0·05$. The F ratios are significant at the $P \leq 0·05$ level (*, † and ‡ denote group, treatment and interaction respectively) (by two-way ANOVA).

#### Effect of high vitamin A supplementation on RPWAT cell density and apoptotic index

The cell density of RPWAT of the stock diet-treated obese rats (B I) was significantly low compared with their lean counterparts (A I). However, chronic dietary challenging with high doses of vitamin A did not bring about any change in the cell density of adipose tissue of the lean and obese rats (A II and B II respectively) (data not shown).

Obese WNIN/Ob rats had an increased RPWAT apoptotic index compared with their lean counterparts (A I). However, chronic dietary challenging with high doses of vitamin A did not bring about any change in the cell density of adipose tissue of the lean and obese rats (A II and B II respectively) (data not shown).

In addition, interaction between phenotype and treatment significantly affected the apoptotic index, but not the cell density.

#### DNA fragmentation assay

DNA fragmentation was not observed in the RPWAT of the lean rats of group A I which received the stock diet. Interestingly, the same adipose tissue of the stock
diet-fed obese rats (B I) and also the lean and the obese rats maintained on a high vitamin A dietary regimen (A II and B II groups) exhibited nucleosomal DNA fragmentation (Fig. 2).

**Effect of high vitamin A supplementation on apoptosis-related proteins**

The obese rats of group B I and B II (irrespective of the vitamin A content in their diets) exhibited an under-expression of an important anti-apoptotic protein Bcl2 in RPWAT (Fig. 3A) compared with that observed in lean rats receiving stock diet (A I). Further, chronic challenging with the high vitamin A diet resulted in a significant reduction in RPWAT Bcl2 expression in the lean rats (A II), when compared with their stock diet-fed lean counterparts (A I). On the other hand, no such effect was seen in obese rats (B II), when compared with the expression observed in their respective lean and obese counterparts consuming stock diet (A I and B I respectively). In addition, this particular dietary regimen (129 mg vitamin A/kg diet) also resulted in over-expression of Bax, a pro-apoptotic protein, in lean rats (A II) compared with their respective control rats fed on a normal vitamin A diet (A I). However, no such over-expression was observed in obese rats (B II) compared with their obese counterparts fed on stock diet (normal dose of vitamin A) (B I) (Fig. 3A).

The ratio of Bcl2–Bax of RPWAT was significantly lower in the lean rats maintained on the high vitamin A diet (A II) as compared with lean rats receiving the stock diet (2·6 mg vitamin A/kg diet). However, the ratio was not altered in obese rats fed on an identical dietary regimen (129 mg vitamin A/kg diet) when compared with their respective control rats fed on the stock diet (having 2·6 mg vitamin A/kg diet) (B I) (Fig. 3B). Furthermore, the Bcl2–Bax ratio was significantly influenced by interaction between phenotype and treatment.

**Discussion**

Adipose tissue mass is tightly regulated by both the size and/or number of the adipocytes and the latter, in turn, is regulated by pre-adipocyte recruitment, differentiation and adipocyte apoptosis (Ailhaud 1990, Bjorntorp 1991). Further, recent studies substantiate the concept that adipocyte deletion by apoptosis is a significant contributor to the regulation of adipose tissue mass and its loss during weight reduction (Della-Fera et al. 2001, Hargrave et al. 2002, Fischer-Posovszky et al. 2004, Kim et al. 2004, Sun & Zemel 2004). Our retroperitoneal adipose tissue cell density data clearly showed that chronic vitamin A challenging through diet had no impact on cell size. This formed the basis for our hypothesis that vitamin A-mediated loss of adipose tissue
could be through altered (decreased) adipocyte number rather than size.

The observed differences between the lean and the obese phenotype with regard to various obesity (body weight gain and adiposity index) and apoptosis-related parameters (apoptotic index, Bcl2 and Bax expression) could be explained by the differences in their genetic composition. Despite identical treatments (vitamin A supplementation), the two phenotypes responded differently (with regard to the above-mentioned parameters) and this cannot be explained solely by genetic differences. Hence, the role of complex interactions between the phenotype (genetic make-up) and nutrient involved should be considered. In view of this, the data were subjected to two-way ANOVA and the results are presented.
tissue. However, the mechanism underlying these paradoxical findings is unclear.

Although, in general, decreased apoptosis contributes to obesity, this may not be true for this particular obese rat model (WNIN/Ob rat strain), thereby implicating the role of non-apoptotic pathways, namely preadipocyte recruitment, differentiation and the thermogenic pathway, in manifesting obesity. This derives further support from the concept that adipocyte number is not predetermined at the time of birth, but increases even in adulthood (Kawada et al. 2001). Interestingly, vitamin A metabolites, besides their effects on apoptosis,

Figure 3 Effect of high vitamin A on RPWAT apoptosis-related proteins expression. (A) The upper and middle panels are representative western blots of RPWAT Bcl-2 and Bax protein expression respectively and the lower panel shows the Ponceau-S-stained blot for the equal loading control. (B) RPWAT Bcl2–Bax ratio quantified densitometric values are expressed relative to a value of 1 for A I (lean) as control. Values are means±S.E. for three to four rats. Mean values without common superscripts are significant at P≤0.05. The F ratios are significant at the P≤0.05 level (*, † and ‡ for group, treatment and interaction respectively) (by two-way ANOVA).
impact two important non-apoptotic pathways, namely preadipocyte differentiation and the thermogenic pathway. Although the former aspect was not addressed in this particular study, the latter has been studied in detail and reported elsewhere (Jeyakumar et al. 2005).

UCP1 expression in the BAT of these rats indicated that obese rats receiving normal levels of vitamin A had lower UCP1 mRNA levels compared with their lean counterparts maintained on identical diets. Further, vitamin A supplementation resulted in significant over-expression in the obese but not in the lean phenotype (Jeyakumar et al. 2005). This particular observation and several other in vitro studies and those employing experimental models of obesity highlight the role of the non-apoptotic (thermogenic) pathway in the regulation of adiposity/body weight gain (Alvarez et al. 1995, Puigserver et al. 1996, Kumar & Scarpace 1998, Kumar et al. 1999, Bonet et al. 2000, Villarroya et al. 2004).

RA induces thermogenic activity by enhancing the expression of BAT UCP1 through its nuclear receptors RXR and RAR (Villarroya et al. 1999, Felipe et al., and Bonet et al. 2003). The RAR receptors homodimerize or heterodimerize with RXR and ligand dependently bring about specific gene activation (BAT UCP1). On the other hand, RXR homodimerize or heterodimerize with various members of the nuclear hormone superfamily receptors (Peroxosome proliferator-activated receptor/Thyroid hormone receptor/Vitamin D receptor) RAR and ligand dependently activate several other genes and elicit various physiological responses (Bonet et al. 2003).

It has recently been shown that the administration of a synthetic RXR agonist (LG268) through oral and intracerebral routes to Zucker fa/fa rats induces anorexia, decreases food intake, body weight gain and adiposity and activates apoptotic pathway without affecting lean body mass, possibly by activating RXR receptors in the central nervous system (Ogilvie et al. 2004). However, in the present study, vitamin A feeding, although resulting in reduced adiposity and body weight gain in obese rats, had no effect on food intake and retroperitoneal adipose tissue apoptosis. These effects in obese rats of the WNNIN/Ob strain could obviously be due to the activation of non-apoptotic pathways.

Taken together, a balance between apoptotic and non-apoptotic pathways determines adipose tissue weight/homeostasis. Notably, nutrients like vitamin A have a modulatory role on these two opposite events which, in turn, are determined by the genetic make-up of the species. This is evident from the differential expression of some apoptosis-related proteins (Bcl2 and Bax) in the lean and obese phenotypes and their divergent response to the identical dose of vitamin A, in terms of the expression of these proteins and UCP1 expression in these two phenotypes.

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