The red wine phenolics piceatannol and myricetin act as agonists for estrogen receptor α in human breast cancer cells

M Maggiolini, A G Recchia, D Bonofiglio, S Catalano, A Vivacqua, A Carpino, V Rago, R Rossi and S Andò

Abstract

Previous epidemiological reports have suggested that red wine intake is associated with beneficial health effects due to the ability of certain phytochemical components to exert estrogen-like activity. It has been also documented that estrogens induce the proliferation of hormone-dependent breast cancer cells by binding to and transactivating estrogen receptor (ER) α, which in turn interacts with responsive DNA sequences located within the promoter region of target genes. In order to provide further insight into the positive association between wine consumption and the incidence of breast carcinoma in postmenopausal women, we have evaluated the estrogenic properties of two abundant wine-derived compounds, named piceatannol (PIC) and myricetin (MYR), using as model systems the estrogen-sensitive MCF7 and the endocrine-independent SKBR3 breast cancer cells. On the basis of our experimental evidence PIC and MYR may contribute to the estrogenicity of red wine since: (1) they transactivate endogenous ERα; (2) they activate the agonist-dependent activation function (AF) 2 of ERα and ERβ in the context of the Gal4 chimeric proteins; (3) they rapidly induce the nuclear immunodetection of ERα; (4) they regulate the expression of diverse estrogen target genes; (5) they compete with 17β-estradiol for binding to ERα and ERβ; and – as a biological counterpart of the aforementioned abilities – (6) they exert stimulatory effects on the proliferation of MCF7 cells. Hence, the estrogenic activity of PIC and MYR might be considered at least as a potential factor in the association of red wine intake and breast tumors, particularly in postmenopausal women.

Introduction

Piceatannol (PIC; 3,5,3′,4′-tetrahydroxy-trans-stilbene) and myricetin (MYR; 3,3′,4′,5′,7-hexahydroxyflavone) (Fig. 1) are phenolic compounds that occur naturally in grapes and red wine (MacDonald et al. 1998, Teugo et al. 1998). The total amount of PIC and MYR in red-grape wines has been reported to be up to 15 mg/l (MacDonald et al. 1998, Burns et al. 2000, Vuorinen et al. 2000, Cantos et al. 2003), however the biotransformation of the abundant red wine component named resveratrol contributes to increase PIC concentrations at tissue level (Piver et al. 2004). Previous studies have demonstrated that PIC is a natural tyrosine kinase inhibitor (Geahlen & McLaughlin 1989) exerting repressive effects in leukemia, melanomas and colorectal cancer cell lines (Wieder et al. 2001, Wolter et al. 2002, Larrosa et al. 2004), while MYR has been identified as a potent topoisomerase II repressor similar in activity to the epipodophyllotoxin widely used in cancer therapy (Markovits et al. 1989, Azuma et al. 1995, Chang et al. 1995). Several epidemiological investigations have suggested that the consumption of red wine is associated with a variety of health benefits due to the estrogenic activity of certain bioactive components (Stampfer et al. 1988, Frankel et al. 1993, Bertelli et al. 1995, Goldberg et al. 1995, Gronbaek et al. 1995, Pace-Aciak et al. 1995). It has been largely reported that 17β-estradiol (E2) mainly acts by binding to and transactivating estrogen receptor (ER) α and ERβ, however different transduction pathways involved in the estrogen signaling have recently received a great deal of attention (Katzenellenbogen et al. 2000, Safe 2001, Maggiolini et al. 2004, Simoncini et al. 2004). A large body of evidence has demonstrated that E2 and E2-like compounds elicit stimulatory effects on breast cancer, the leading cause of tumor deaths among women in Western countries (Hoskins & Weber 1994, Eisen & Weber 1998, Lopez-Otin & Diamandis 1998, Maggiolini et al. 2001). Therefore, much effort has been addressed to ascertain the ability of natural compounds to interact with ERα and to trigger breast tumor progression. In this regard, it should be taken into account that wine consumption even in moderate amounts is directly associated with the
occurrence of breast carcinoma particularly in post-menopausal women (van den Brandt et al. 1995, Feigelson et al. 2001). Notably, in a multicentre Italian case-control study an increased risk of breast cancer was attributed to wine ingestion compared with other types of alcoholic beverages, explaining up to 12% of breast tumor cases in Italy (Ferraroni et al. 1998). In the present study, we have used as model systems the hormone-sensitive MCF7 and endocrine-independent SKBR3 breast cancer cells in order to provide new insight into the molecular mechanisms by which the wine-derived PIC and MYR behave as estrogen-like compounds. In a concentration range physiologically achievable with a moderate wine consumption, both phytochemicals elicited growth stimulatory effects in MCF7 cells as a biological counterpart of their agonistic activity for ERα.

Materials and methods

Reagents

E2, PIC, MYR and cycloheximide (Cx) were purchased from Sigma, while the ER antagonist ICI 182,780 (ICI) was obtained from Tocris Chemicals (Bristol, UK). Hydroxyflutamide (OHF) and ZK 98299 were a gift from Schering (Berlin, Germany). All compounds were solubilized in dimethylsulfoxide, except E2 which was dissolved in ethanol.

Plasmids

Firefly luciferase reporter plasmids used were XETL (Bunone et al. 1996) and TFF1/pS2-ERE (Berry et al. 1989) for the ERs and GK1 (Webb et al. 1998) for the Gal4 fusion proteins. The reporter plasmid XETL carries firefly luciferase sequences under the control of an estrogen response element (ERE) upstream of the thymidine kinase promoter. TFF1/pS2-ERE (a gift from V. Giguère, McGill University, Quebec City, Canada) contains the ∼1050 bp TFF1/pS2 promoter preceding the luciferase reporter of pGL3 (Berry et al. 1989). As an internal transfection control, we co-transfected the plasmid pRL-CMV (Promega) that expresses renilla luciferase enzymatically distinguishable from firefly luciferase by the strong cytomegalovirus enhancer/promoter. Gal4 chimeras Gal-ERα, Gal-ER(V), Gal-ER(R), Gal-ER(L), Gal-ER(543/4A), Gal-ER(ΔF) and Gal-ERβ were expressed from plasmids GAL93.ER(G), GAL93.ER(V), GAL93.ER(R), GAL93.ER(L), GAL93.ER(ML543/4AA), GAL93.ER(ΔF) and GAL93.ERβ respectively. They were constructed by transferring the coding sequences for the hormone-binding domain (HBD) of ERα (amino acids 282–595) from HEG0 (Bunone et al. 1996), pCMVh ERG400 V (Aumais et al. 1997), pCMVhERG521R (Ekena et al. 1996), pCMVhERL525A (Ekena et al. 1996), a PCR-
mutagenized intermediate with the point mutations M543A-L544A, a PCR fragment lacking the coding sequences for the F domain, and for the ERβ HBD (C-terminal 287 amino acids) from plasmid pCMV5-hERbeta (a gift from J.-Å. Gustafsson, Karolinska Institute, Huddinge, Sweden) respectively, into the mammalian expression vector pSCTEVGal93 (Siepel et al. 1992).

Cell culture
Wild-type ERα-positive MCF7 human breast cancer cells were a gift from E. Surmacz (Sbarro Institute for Cancer Research and Molecular Medicine, Philadelphia, USA), the ER-negative SKBR3 human breast cancer cells and the ERα-containing human uterine Ishikawa cells were a gift from D. Picard (Department of cell biology, University of Genève, Switzerland). MCF7 and Ishikawa cells were maintained respectively in Dulbecco’s modified Eagle’s medium (DMEM) or DMEM/F12 both without phenol red supplemented with 10% fetal bovine serum (FBS) (Invitrogen); SKBR3 cells were maintained in RPMI 1640 without phenol red supplemented with 10% FBS (Invitrogen).

Transfections and luciferase assays
A total of 10⁵ MCF7, SKBR3 and Ishikawa cells were transferred into 24-well plates with 500 µl of regular growth medium/well the day before transfection. Medium lacking phenol red as well as serum was used on the day of transfection, which was performed by Fugene6 reagent (Roche) with a mixture containing 0.5 µg reporter plasmid, 5 ng pRL-CMV and 0.1 µg effector plasmid where applicable. After 6 h, treatments were added and cells were incubated for a further 18–20 h. Luciferase activity was then measured with the Dual Luciferase Reporter Assay System (Promega) according to the manufacturer’s recommendations. Firefly luciferase activity was normalized to the internal transfection control provided by renilla luciferase activity. Luciferase activity of cells receiving vehicle was set as 1-fold induction, upon which the results of treatments were calculated.

RT-PCR
MCF7 cells were grown in 10 cm dishes to about 60% confluence in regular growth medium and then were switched to medium lacking phenol red as well as serum for 24 h. Thereafter, treatments were added for 24 h before lysis. The primers for ERα, estrogen-inducible trefoil factor TFF1/pS2 (Brown et al. 1984), cathepsin-D (Cavailles et al. 1991), estrogen-responsive RING finger protein (EFP) (Ikeda et al. 2000) as well as for the internal control acidic ribosomal phosphoprotein 36B4 (Laborda 1991) were: 5′AAATTCAGATAATCGAGCGGCAAG3′ (ERα forward) and 5′GTGTGTTCAACATTGTCCCCCTCTC3′ (ERα reverse); 5′TTCTATCTCATAATCCACGACG3′ (pS2 forward) and 5′TTGAGTTAGTCAAAGTCAGAGC3′ (pS2 reverse); 5′AACACAGGTCGGGCTCTC3′ (cathepsin-D forward) and 5′ATGACAGAACAAGCTGTGC3′ (cathepsin-D reverse); 5′CGTGTGATTGTGTTGAGACG3′ (EFP forward) and 5′CCCGAGGTGGGAACGTGAACC3′ (EFP reverse); 5′CTCAACATCTCCCCCTCTC3′ (36B4 forward) and 5′CAATCCCATACCTCGTGCC3′ (36B4 reverse); these primers yielded products of 345, 210, 303, 162 and 400 bp with 20, 15, 20, 20 and 15 PCR cycles respectively.

Western blotting
MCF7 cells were grown in 10 cm dishes to about 60% confluence in regular growth medium and then were switched to medium lacking phenol red as well as serum for 24 h. Thereafter, treatments were added for 24 h before lysis. In the case of Cx, cells were pretreated for 3 h prior to the addition of ligands. Cells were lysed with 300 µl of 50 mM HEPES (pH 7.5), 150 mM NaCl, 1·5 mM MgCl₂, 1 mM EGTA, 10% glycerol, 1% Triton X-100, 1% SDS, and a mixture of protease inhibitors containing 1 mM aprotinin, 20 mM phenylmethylsulfonyl fluoride and 0·2 M sodium orthovanadate. Equal amounts of whole protein extract were resolved on a 10% SDS-polyacrylamide gel, transferred to a nitrocellulose membrane (Amersham), probed overnight at 4 °C with the antibodies F-10 against ERα, and then revealed using the ECL System (Amersham).

Immunocytochemical staining
MCF7 cells were maintained in medium lacking phenol red as well as serum for 3 days, treated for 1 h and then fixed in fresh paraformaldehyde (PFA; 2% for 30 min). After PFA removal, hydrogen peroxide (3% in methanol for 30 min) was used to inhibit endogenous peroxidase activity. Cells were then incubated with normal horse serum (10% for 30 min) to block the non-specific binding sites. Immunocytochemical staining was performed using as the primary antibody a mouse monoclonal immunoglobulin (Ig) G generated against the human C-terminus of ERα (F-10, Santa Cruz Biotechnology; 1:50 overnight at 4 °C). A biotinylated horse-anti-mouse IgG (1:600 for 60 min at room temperature) and 5′-biotinylated anti-rabbit IgG (1:2000 for 30 min) were used as secondary antibodies. Biotinylated antigens were visualized with the Vectorstain ABC kit (Vector Laboratories) and 3-amino-9-ethylcarbazole (AEC) substrate (Sigma). Immunohistochemical staining was performed using as the primary antibodies a mouse monoclonal IgG generated against the human C-terminus of ERα (F-10, Santa Cruz Biotechnology; 1:50 overnight at 4 °C). A biotinylated horse-anti-mouse IgG (1:600 for 60 min at room temperature) and 5′-biotinylated anti-rabbit IgG (1:2000 for 30 min) were used as secondary antibodies. Biotinylated antigens were visualized with the Vectorstain ABC kit (Vector Laboratories) and 3-amino-9-ethylcarbazole (AEC) substrate (Sigma).
temperature) was applied as the secondary antibody (Vector Laboratories, Burlingame, CA, USA). Subsequently, the amplification of avidin-biotin-horseradish peroxidase complex (ABC complex/HRP; Vector Laboratories; 1:100 for 30 min at room temperature) was carried out and 3,3’-diaminobenzidine tetrachloride dihydrate (Vector Laboratories) was used as a detection system. Cells were rinsed after each step with Tris-buffered saline (0-05 M Tris–HCl plus 0-15 M NaCl, pH 7-6) containing 0-05% Triton-X100 (TBS-T). In control experiments cells were processed replacing the primary antibody with mouse serum (Dako S.p.A., Milan, Italy) or using a primary antibody pre-absorbed (48 h at 4 °C) with an excess of purified ERα protein (PanVera Corp., Madison, WI, USA).

Ligand binding assay for ERs

The ability of PIC and MYR to compete with [3H]E2 for binding to ERα and ERβ was evaluated and compared with that of E2. Two picomoles of purified recombinant human ERα and ERβ proteins (PanVera Corp., Madison, WI, USA), each in 20 mM HEPEs, pH 7-4, 1-5 mM EDTA, 0-5 mM dithiothreitol and 10% (v/v) glycerol, were incubated with 1 nM [2,4,6,7-3H]E2 (72 Ci/mmol; Amersham) in the presence of serial dilutions of unlabeled E2, PIC or MYR for 20–22 h at 4 °C. Bound and free radioligands were separated on Sephadex G-25 PD-10 columns. The amount of receptor-bound [3H]E2 was determined by liquid scintillation counting (OptiPhase, HiSafe 3 and 1409; Wallac, Inc., Gaithersburg, MD, USA). Relative counts per minute were plotted against the concentrations of the ligand, and data were evaluated with the use of a non linear, four-parameter logistic model to estimate the median effective concentration (EC50) value.

Proliferation assays

For quantitative proliferation assays 1 × 10⁴ MCF7 cells were seeded in 24-well plates in regular growth medium. Cells were washed extensively once they had attached and were incubated in medium without serum for 24 h. On the second day, the medium was changed and supplemented with 5% charcoal-stripped (CS)-FBS. Ligands were added at this point, thereafter the 5% CS-FBS medium was renewed every day together with treatments. On day 6, cells were trypsinized and counted in a haemocytometer using the trypan blue exclusion method.

Statistical analysis

Statistical analysis was performed using ANOVA followed by Newman–Keuls testing to determine differences in means. Data are expressed as means ± S.D. P values <0-05 were considered significant.

Results

Transactivation of the endogenous ERα

We first aimed to evaluate whether a transiently transfected ER reporter gene is able to respond to PIC and MYR. The reporter plasmid XETL carries firefly luciferase sequences under the control of an ERE upstream of the thymidine kinase promoter. As an internal transfection control, we co-transfected a plasmid that expresses renilla luciferase, which is enzymatically distinguishable from firefly luciferase, from the strong cytomegalovirus enhancer/promoter. Using MCF7 cells, which express ERα but no ERβ as judged by RT-PCR (data not shown), PIC and MYR were able to induce a substantial XETL expression from the concentration of 10 nM (Fig. 2A). The ER antagonist ICI abolished the activation by E2 and both compounds (Fig. 3A), whereas the antiandrogen OHF, the antiprogestin and antigluocorticoid ZK 98299 had no effect (data not shown), suggesting that PIC and MYR transactivate ERα directly. Next, using the natural pS2 ERE-containing promoter (Berry et al. 1989) we confirmed that both compounds were able to induce the ERα-mediated transcriptional activity (Fig. 2B), which was no longer noticeable in the presence of ICI (Fig. 3B). For comparison, we performed the same transfection experiments in uterine Ishikawa cancer cells showing that PIC and MYR activated the endogenous ERα (Fig. 2C) in a direct manner since ICI inhibited the transcriptional response also in this cellular context (Fig. 3C).

Activation of ERα and ERβ in a heterologous system

To provide mechanistic evidence on the ability of PIC and MYR to transactivate ERα and to examine the response of ERβ, we turned to a heterologous system using SKBR3 cells lacking ERs. Chimeric proteins consisting of the DNA-binding domain (DBD) of the yeast transcription factor Gal4 and the ERα or the ERβ HBDs containing the ligand-dependent activation function (AF) 2 were both activated by PIC and MYR although with a different potency compared with E2 (Fig. 4A). Hence, the HBD of each ER isoform was different potency compared with E2 (Fig. 4A). Hence, the HBD of each ER isoform was sufficient to allow the functional response, while the Gal4-DBD fused to the AF1 domain of ERα did not elicit transcriptional effects upon all treatments (Fig. 4B). Next, we examined the response of diverse ERs HBD mutants in the context of Gal4 proteins. The two-point mutations L525A and G521R that require a high E2 concentration for a substantial transactivation (Ekena et al. 1996) elicited very low activity upon PIC and MYR (Fig. 4B and C). The AF2 mutant M543/L544A failed to respond to all treatments (Fig. 4C), while the point mutation G400 V and the mutant lacking the F-domain at the C-terminal HBD core exhibited a strong response.
to E2, PIC and MYR (Fig. 4D). Taken together, our results argue that mutations of ERα in the hormone binding pocket are able to alter the transcriptional activity induced by E2 as well as that of the compounds tested.

Nuclear immunolocalization of ERα

Upon ligand binding, ERα undergoes conformational changes allowing the formation of receptor dimers and the interaction with specific EREs contained within the regulatory region of target genes (Tsai & O’Malley 1994, Beato et al. 1996, Pratt & Toft 1997). Performing an immunocytochemical assay we aimed to evaluate the ability of short PIC and MYR treatments (1 h) to elicit a nuclear localization of the endogenous ERα in MCF7 cells. No detectable ERα immunoreactivity was observed in basal experimental conditions obtained by maintaining cells in medium lacking phenol red as well as serum for 3 days (Fig. 5). Only in the nuclear compartment did E2, PIC and MYR produce strong staining signals and these were abrogated in the presence of the ER antagonist ICI 182 780 (Fig. 5). No signals were observed after replacing the anti-ERα antibody by irrelevant rabbit IgG or by using the primary antibody pre-absorbed with an excess of receptor protein (data not shown). These findings further supported the functional activity elicited by compounds tested in transfection experiments.

Down-regulation of ERα mRNA and protein levels

Levels of ERα in breast cancer cells are autoregulated by E2 through different mechanisms which include a reduced transcription rate of the ERα gene together with an increased turnover of the ligand-activated protein (Santagati et al. 1997). Hence, to evaluate whether PIC and MYR are also able to modulate the mRNA of ERα in MCF7 cells we performed a semiquantitative RT-PCR standardizing its expression by the housekeeping gene 36B4. Interestingly, both compounds induced a receptor down-regulation (Fig. 6A and B) which was observed even at the protein level depending on new synthesis (Fig. 6C and D) (El Khissiin & Leclercq 1999, Nonclercq et al. 2004).

Figure 2 PIC and MYR activate the endogenous ERα. MCF7 breast cancer cells were transfected with the luciferase reporter plasmid XETL(A) or TFF1/pS2 (B) and treated with increasing concentrations of E2, PIC and MYR. (C) Uterine Ishikawa cancer cells were transfected as described for panel A. Luciferase activities were standardized to the internal transfection control and expressed as the ratio of induced activity to the activity in the absence of ligand. Each data point represents the mean of triplicate samples of three separate experiments.
Up-regulation of estrogen target genes

As aforementioned, the E2/ERα-activated complex regulates the expression of genes that contain ERE sequences within the promoter regions (Truss & Beato 1993). Following the same experimental procedure described above, we documented the ability of PIC and MYR to up-regulate, like E2, the mRNA expression of target genes – such as pS2/TIFF1, cathepsin D and EFP E2, (Fig. 6A and B).

Competition for ER binding

The results obtained strongly suggested that PIC and MYR are ERs for ligands. This issue was examined directly by a competition binding experiment with purified recombinant human ERα and ERβ proteins. Our data demonstrated that both compounds compete with the radiolabeled E2 tracer for binding to the ERs in a concentration-dependent manner, however they are poor binders particularly for ERα in comparison to ERβ (Fig. 7). The latter phenomenon is in agreement with a previous report on ER isoform-specific preferences in binding to environmental estrogens (Kuiper et al. 1997).

Proliferative effects on hormone-sensitive breast cancer cells

Having clearly demonstrated that PIC and MYR are functional activators of ERα, we evaluated in a time course study the proliferative response of MCF7 cells to increasing concentrations of both ligands. Cells were lysed on the indicated days, counted in a hemocytometer, and results obtained upon treatments were compared with those of untreated cells. Notably, both compounds were able to induce progressive proliferative effects (Fig. 8A) which were no longer noticeable in presence of ICI (Fig. 8B), suggesting that even the growth stimulation is mediated by an ERα-dependent mechanism.

Discussion

Our findings provide new insights into the activity exerted by the wine-derived phenolics PIC and MYR which can be considered ER agonists on the basis of their ability: (1) to activate endogenous ERα in hormone-dependent MCF7 and Ishikawa breast and uterine cancer cells respectively; (2) to activate the ligand-dependent AF2 domain of ERα and ERβ in the

Figure 3 The ER antagonist ICI inhibits the ERα transactivation by PIC and MYR. Cells were transfected with XETL (A and C) or with TFF1/pS2 ERE (B) and treated as indicated. Each data point represents the mean of triplicate samples of three separate experiments. ●, ■, ▲, P<0.05.
context of chimeric proteins with the Gal4 DBD in ER-negative SKBR3 breast cancer cells; (3) to elicit responses similar to E2 using different expression vectors for ERα mutated in the AF2 domain; (4) to rapidly induce a nuclear immunodetection of ERα; (5) to regulate ERα mRNA and protein levels as well as the expression of estrogen target genes such as TFF1/pS2, cathepsin D and EFP; (6) to compete with E2 for binding to purified ERα and ERβ proteins; (7) to stimulate the proliferation of MCF7 cells at concentrations that may be physiologically achievable through a moderate wine intake.

In recent years, the biological properties of natural estrogen-like compounds have attracted increasing interest for the potential beneficial effects on human health (Cotton 1994, Safe 1995, Lopez-Otin & Diamandis 1998). In this regard, a reduced incidence and mortality from a variety of diseases has been associated with moderate wine consumption (Fuhrman et al. 1995, Goldberg et al. 1995, Whitehead et al. 1995, Gronbaek et al. 2000, German & Walzem 2000), which is considered a relevant source of estrogenic compounds like the well-characterized resveratrol (Kopp 1998). In order to ascertain the mechanisms involved in the
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potential protective role exerted by resveratrol, many
studies have been performed demonstrating its ability to
regulate lipid metabolism and platelet function as well
as to trigger apoptotic events in different cancer cells
(Bagchi et al. 2001, Bianchini & Vainio 2003, Pervaiz
2003). However, other investigations evaluating the
potential estrogen-like properties of resveratrol have
documented an agonistic activity for ERα including the
ability to stimulate the proliferation of breast tumor cells
(Gehm et al. 1997, Basly et al. 2000, Nakagawa et al.
2001, Levenson et al. 2003, Gehm et al. 2004). These
findings are reminiscent of those reported for other
abundant dietary flavonoids (Hsieh et al. 1998, Maggiolini et al. 2001 and references therein), and may
suggest that the biological actions of resveratrol are
either mediated by ERα or involve ER-independent
mechanisms mainly in relationship to the cellular
environment and levels of exposure (Gehm et al. 1997,
Mgbonyebi et al. 1998, Ashby et al. 1999, Lu & Serrero
et al. 2001). Hence, phytoestrogens and other natural
compounds are potentially responsible for diverse
pharmacological effects, which depend on bioavailability
as well as metabolic clearance rates influencing
consequently ER agonism/antagonism and even anti-
cancer activity. In this respect, it is worth noting that
resveratrol can undergo hydroxylation to yield PIC via
different cytochrome P450 enzymes like CYP1B1 which
has been found overexpressed in a wide variety of
Taking into account that PIC also derives directly from
wine intake, we performed a panel of in vitro assays
designed to better define its estrogen-like properties to-
gether with those of MYR, an additional phytochemical

Figure 6 PIC and MYR regulate the expression of estrogen target genes. (A) Semiquantitative
RT-PCR of ERα, TFF1/pS2, cathepsin D and EFP mRNA. MCF7 cells were stimulated for 24 h with
1 µM E2, PIC and MYR; 36B4 mRNA levels served as a control. (B) Quantitative representation of
data of two independent experiments including that of panel A after densitometry and correction for
36B4 expression. (C) Immunoblot of ERα from MCF7 cells treated for 24 h with 1 µM E2, PIC and
MYR and 50 µM Cx where applicable. (D) Quantitative representation of data of two independent
experiments including that of panel C after densitometry. β-Actin served as a loading control. ○, □, ●,
■, P < 0.05.
occurring abundantly in red wine. At a low concentration range physiologically achievable through wine consumption, both compounds stimulated the growth of hormone-sensitive breast tumor cells as ERα activators. Moreover, the results obtained in transfection experiments with Gal4 chimeras suggested that PIC and MYR

Figure 7 PIC and MYR compete with [3H]E2 for ERα and ERβ binding. Competitive binding of increasing concentrations of PIC and MYR to purified recombinant human ERα (A) and ERβ (B) protein. Each data point represents the mean of triplicate samples of three separate experiments.

Figure 8 PIC and MYR stimulate the proliferation of breast cancer cells. (A) Time- and dose-response study to PIC and MYR in MCF7 cells. Data obtained in cells after treatments were compared with those of vehicle-treated cells set as 100%. (B) MCF7 cells were treated as indicated, lysed and counted on day 6. Each data point represents the mean of triplicate samples of three separate experiments. □, P<0.05.
are AF2 agonists alongside other selective estrogen receptor modulators (SERMs) grouped as class I estrogens, according to the molecular classification previously proposed (Jordan et al. 2001). The ability of ligands to modulate intracellular levels of ERα is important for ERα pharmacology. The human ERα is a substrate for ubiquitination in the presence of E2 and the exposure to different ligands has been shown to lower ERα levels by a proteasome-mediated proteolysis (Nawaz et al. 1999). The down-regulation of ERα following treatments with PIC and MYR in MCF7 cells further supported their agonistic activity observed also through a ligand-dependent receptor degradation process. Moreover, despite a low binding affinity for ERα both compounds up-regulated the expression profiles of diverse estrogen target genes providing a reliable assessment of the transcriptional efficacy exhibited in the context of natural DNA response elements.

A large body of evidence has suggested that ERα-mediated estrogen signaling plays a major role in the etiology and progression of breast carcinoma (Ciocca & Fanelli 1997, Lopez-Otin & Diamandis 1998). Consequently, ERα antagonists have been extensively utilized as adjuvant endocrine treatment to prevent such malignancy and/or its recurrence (Early Breast Cancer Trialists Collaborative Group 1998, Cosman & Lindsay 1999). However, about half of ERα-positive tumors fail to respond to antiestrogen therapy from the beginning and most of the remaining ones eventually acquire pharmacological resistance. In this context, it should be noted that E2 and diverse phytoestrogens irrespective of ER expression trigger rapid genomic and non-genomic events (Brownson et al. 2002, Maggiolini et al. 2004) which, at least in part, contribute to the failure of antiestrogen treatment as well as to breast tumor progression. On the basis of the aforementioned observations and the present findings, the estrogenic properties of phytochemicals studied might be considered as a risk factor in postmenopausal women who habitually consume red wine (van den Brandt et al. 1995, Ferraroni et al. 1998, Feigelson et al. 2001, Poschal & Seitz 2004). However, taking into account that PIC and MYR exert inhibitory effects in different tumor cells (Markovits et al. 1989, Azuma et al. 1995, Chang et al. 1995, Wieder et al. 2001, Wolter et al. 2002, Larrosa et al. 2004), their concentrations as well as the cellular context may direct various biological responses as has been previously reported for resveratrol and other phytoestrogens (Gehm et al. 1997, Hsieh et al. 1998, Basly et al. 2000, Maggiolini et al. 2001, Nakagawa et al. 2001, Bianchini et al. 2003, Peraza 2003). Herein, we have provided mechanistic insight into the estrogenic potential of PIC and MYR that may be involved in the beneficial as well as the adverse effects associated with wine consumption. In this vein, further investigations are needed to elucidate the level of absorption and exposure to these phytochemicals, and the dose-dependent properties as well as the activity exerted in vivo in estrogen target tissues such as breast, uterus and bone.

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