Binding characteristics of antibodies to the TSH receptor

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ABSTRACT

We have used fragments of the TSH receptor (TSHR) expressed in E. coli as glutathione S-transferase fusion proteins to produce rabbit polyclonal antibodies and a panel (n=5) of monoclonal antibodies to the extracellular fragment of the TSHR. The binding characteristics of the antibodies to linear, conformational, glycosylated and unglycosylated forms of the receptor in different assay systems have been investigated.

The reactivity of these antibodies with the TSHR was assessed by Western blotting with both native and recombinant human TSHR expressed in CHO cells, immunoprecipitation of 35S-labelled full-length TSHR produced in an in vitro transcription/translation system, immunoprecipitation of 125I-TSH/TSHR complexes, inhibition of 125I-TSH binding to the TSHR and fluorescence activated cell sorter (FACS) analysis of binding to CHO-K1 cells expressing the TSHR on their cell surface. Fab fragments of monoclonal antibodies were isolated, labelled with 125I and used to determine the affinity constants of these antibodies with receptor, bound and free Fab being separated by polyethylene glycol (PEG) precipitation.

Rabbit polyclonal and mouse monoclonal antibodies reacted with the TSHR in Western blotting and one monoclonal antibody (3C7) was able to inhibit 125I-TSH binding to native human TSHR (74% inhibition), recombinant human TSHR (84% inhibition) and porcine TSHR (65% inhibition). Affinity constant values for TSHR monoclonal antibody Fab fragments calculated using Scatchard analysis were about 10⁷ M⁻¹. Four out of five monoclonal antibodies reacted in FACS analysis with TSHR expressed on the surface of CHO-K1 cells. The FACS unreactive monoclonal (3C7) bound well to detergent solubilised TSH receptors and this emphasised the importance of using a combination of FACS analysis and radioactively-labelled probes in analysis of the TSH receptor.

The monoclonal antibodies produced in this study were found to be of relatively low affinity but proved useful for detection of the receptor by Western blotting and by FACS analysis.

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INTRODUCTION

The thyrotrophin receptor (TSHR) is a major thyroid autoantigen and TSHR autoantibodies (TRAb) are responsible for the hyperthyroidism of Graves’ disease (Rees Smith et al. 1988, Furmaniak & Rees Smith 1993). Sequence analysis indicates that the TSHR gene codes for a single peptide chain with a computed molecular mass of 84 kDa (Libert et al. 1989, Nagayama et al. 1989, Frazier et al. 1990, Misrahi et al. 1990). However the region of the peptide chain linking the receptor’s large extracellular domain to its seven membrane spanning domain is readily cleaved giving rise to a A subunit (extracellular domain) linked by a disulphide bridge(s) to a B subunit (membrane spanning domain) (Buckland & Rees Smith 1984, Kajita et al. 1985a,b, Rees Smith et al. 1988, Loosfelt et al. 1992).

The binding sites for TSH and for TRAb are formed principally by the receptor’s A subunit but the A subunit alone does not appear sufficient for high affinity TSH binding (Davies Jones et al. 1984, 1985). Similar observations have been reported following studies using recombinant extracellular domain of the TSHR (Harfst et al. 1992, Nagayama & Rapoport 1992, Huang et al. 1993, Kosugi et al. 1997).

Experimentally-produced antibodies to different regions of the TSHR could be helpful in

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characterising the receptor and we now describe the preparation of polyclonal and monoclonal antibodies to the extracellular domain of the TSHR.

MATERIALS AND METHODS

Production of an eukaryotic cell line expressing full length TSHR

An ATG start codon in the 5’ untranslated region (5’UTR) of the full length hTSHR cDNA (Misrahi et al. 1990) was removed by AvaI digestion and the cDNA cloned into pRC/CMV (Invitrogen, Leek, The Netherlands). DNA encoding the full length TSHR (pRC/CMV/hTSHR) was transfected into CHO cells (CHO-K1 from ECACC) using the calcium phosphate precipitation method (Chen & Okayama 1987). Clones expressing TSHR were detected using 125I-TSH binding directly to cells growing on 24-well plates. The clones showing highest TSH binding were expanded and recloned twice by limiting dilution. One stable cell line expressing 4 × 10^5 TSHRs per cell, as assessed by Scatchard analysis of TSH binding (Scatchard 1949) was selected and grown in continuous culture in DMEM (Gibco-BRL, LifeTechnologies, Paisley, Strathclyde, UK) containing high glucose and 10% foetal calf serum (FCS), with 1000 µg/ml geneticin sulphate (G-418) (Life Technologies).

Production of GST–TSHR fusion proteins

Two fragments of cDNA coding for the extracellular domain of the TSHR, TSHR800 (amino acids 89–363) and TSHR400 (amino acids 245–380) were cloned into the bacterial expression vector pGEX-2T (Pharmacia, St Albans, UK) to produce in frame fusion proteins with GST (Powell et al. 1996). An overnight culture of E. coli (strain UT580) transformed with pGEX-2T/TSHR plasmids was diluted 1/5 into 2xYTG medium (16g tryptone, 10g yeast extract, 5g NaCl, 20g glucose per litre, pH 7.0), incubated for 3h at 30°C; isopropyl-β-D-thiogalactopyranoside (IPTG) added to a final concentration of 1mM to induce protein expression and incubated for a further 3h. The bacterial pellets were resuspended in PBS (8g NaCl, 0.2g KCl, 1.44g Na₂HPO₄ and 0.24g KH₂PO₄ per litre, pH 7.4) containing 1% Triton X-100 and sonicated three times for 1 min on ice. The inclusion bodies were pelleted, washed in 4M urea, solubilised in 8M urea and separated on 9% acrylamide gel slices in 0.1 M NaHCO₃ and 0.1% SDS pH 7.8 (Smith & Corcoran 1994) dialysed against 50 mM Tris–HCl pH 8·0 and stored in aliquots at −70°C.

Production of antibodies

Female New Zealand White rabbits were immunised with 50 µg GST–TSHR fusion proteins in complete Freund’s adjuvant and boosted at 4-week intervals with 50 µg GST–TSHR in incomplete Freund’s adjuvant until the antibody titre was high (see below for assay method).

Six- to eight-week-old BALB/c mice were immunised with 50 µg GST–TSHR fusion proteins and boosted at 3–4 week intervals until the antibody titre was high (see below for assay method). The fusion of mice spleen cells with X63–Ag8·653 myeloma cells (ECACC) was performed 4 days after the final intraperitoneal boost (De St Groth & Scheidegger 1980). The cells were cultured in 96-well plates in DMEM with high glucose, 20% FCS, 10% Doma Drive (Immune Systems, Paignton, Devon, UK) and selected with HAT (Life Technologies).

Antibody secretion was detected by immunoprecipitation assays (IPA) based on 35S-labelled full length TSHR produced in the in vitro transcription/translation system (TnT) (see below for assay method). Positive wells were recloned two times by limiting dilution (0.5 cells/well), grown up and used to produce ascites.

Immunoprecipitation assay (IPA)

The cDNA encoding full-length TSHR was placed downstream of the T7 promoter in pYES2 (Invitrogen) and used in an in vitro TnT system (Promega, Southampton, Hants, UK) to produce TSHR labelled with 35S-methionine as previously described (Colls et al. 1995, Prentice et al. 1997).

Briefly, 50 µl 35S-labelled TSHR (25 000–30 000 c.p.m.) diluted in HSB (150 mM Tris–HCl pH 8·3, 200 mM NaCl and 10 mg/ml bovine serum albumin containing 1% Tween 20) were added to duplicate 50 µl aliquots of diluted test sera or tissue culture supernatant and incubated for 2 h at room temperature. Immune complexes were then precipitated by addition of either protein A Sepharose (Sigma, Poole, Dorset, UK) or anti mouse IgG agarose (Sigma) and counted for 35S.

Isolation of Fab fragments

TSHR antibody IgGs were purified from ascites fluid using affinity chromatography on Prosep A (Bioprocessing, Consett, UK) according to the manufacturer’s instructions.

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Purified IgG (10 mg/ml) was dialysed against 70 mM sodium acetate, 50 mM NaCl pH 4-0 and digested for 18 h at 37 °C with 1 mg/ml pepsin. The reaction mixture was dialysed against 1 M glycine/NaOH pH 8-6, 150 mM NaCl and passed through a Prosep A column in the same buffer to remove undigested IgG from the (Fab)_2.

The (Fab)_2 fragments were reduced with 10 mM cysteine in 2 M Tris pH 8-3 for 1 h at 37 °C and the reaction terminated by the addition of 200 mM iodoacetamide in 2 M Tris pH 8-3 to a final concentration of 50 mM. Fab fragments were analysed by 9% SDS–PAGE and 50 µl (20 µg) aliquots of Fab were labelled with 125I using the chloramine T method to a specific activity of 500 KBq per µg followed by Sephadex G-25 chromatography (Greenwood et al. 1963).

Preparation of detergent solubilised native and recombinant human TSHR

CHO-K1 cells were grown to confluence (4 days) in a 175 cm² flask, the cells were washed with Dulbecco’s PBS minus calcium and magnesium ions and scraped into 10 ml ice-cold 50 mM NaCl, 10 mM Tris–HCl pH 7-5, 1 mM phenylmethylsulphonyl fluoride (PMSF; buffer A). The cells were pelleted at 1000 g for 5 min at 4 °C, the pellet resuspended in 1 ml buffer A and homogenised with a glass homogeniser on ice. The cell membranes were pelleted at 12 000 g for 30 min at 4 °C, resuspended in 1 ml ice-cold buffer A with 1% Triton X-100 and homogenised. The solubilised TSHR preparations were centrifuged at 90 000 g for 2 h and the supernatant aliquoted and stored at −70 °C.

Native human TSHR was prepared from human thyroid tissue as previously described (Rickards et al. 1981) using 1% Triton X-100.

Precipitation of TSHR/125I-TSH complexes

Solubilised porcine TSHR (RSR Ltd, Cardiff, UK) or recombinant human TSHR were incubated for 2 h at room temperature with an equal volume of 125I-labelled bovine TSH (24 000 c.p.m.; RSR Ltd) to form TSHR/TSH complexes. Fifty microlitres TSHR/TSH complex were incubated with 50 µl antibody for 1 h at 37 °C. Fifty microlitres protein A (10% suspension; RSR Ltd) were then added and incubated for 1 h at room temperature. The mixture was then diluted with 1 ml assay buffer and centrifuged (1500 g for 30 min at 4 °C) and the pellet washed once with 1 ml assay buffer and counted for 125I.

Western blot analysis

Twenty five microlitres solubilised CHO-K1 cells, recombinant human TSHR or native human TSHR were mixed with an equal volume of sample buffer (4% SDS, 20% glycerol, 100 mM Tris–HCl pH 6·8, 0·002% bromphenol blue) plus dithiothreitol (10 mM) and heated to 100 °C for 3 min, electrophoresed on 9% SDS–PAGE gels and blotted onto nitrocellulose. Molecular mass markers (6H; Sigma) were run on each gel.

Western blotting analysis was carried out according to the method of Birk and Koepsell (1987). The membranes were blocked using 1 mg/ml polyvinylalcohol (PVA) in PBS and developed using anti-mouse or anti-rabbit horse-radish peroxidase conjugate followed by enhanced chemiluminescense (ECL) reagents (Amersham International plc, Amersham, Bucks, UK). Calculation of relative mobility was carried out using the centre of each protein band as a reference point.

TSH binding inhibition assays

TSH binding inhibition assays were carried out according to the method of Southgate et al. (1984) using native human TSHR or recombinant human TSHR (expressed in CHO cells). Briefly, 50 µl detergent solubilised TSHR were preincubated with 50 µl sera, purified IgG or Fab fragment for 15 min at room temperature before the addition of 100 µl 125I-TSH (30 000 c.p.m.) and incubation at 37 °C for 1 h. TSHR/125I-TSH complexes were precipitated by the addition of 2 ml 16·5% polyethylene glycol (PEG) precipitator (RSR Ltd) and 25 µl healthy blood donor serum, centrifuged at 1500 g for 30 min at 4 °C, aspirated and the radioactivity of the pellets counted in a gamma counter.

Scatchard analysis of 125I-labelled Fabs binding to detergent solubilised TSHR

The characteristics of 125I-labelled Fabs binding to detergent solubilised recombinant human TSHR were assessed by Scatchard analysis (Scatchard 1949). Fifty microlitres solubilised TSHR were incubated with 50 µl unlabelled Fab (concentrations ranging from 0·4 to 40 µg/ml) and 100 µl 125I-Fab (15 000 c.p.m.) for 1 h at 37 °C. Fifty microlitres healthy blood donor sera and 2 ml 16·5% PEG (RSR Ltd) were added, mixed, centrifuged at 1500 g for 30 min at 4 °C and the radioactivity remaining in the pellet after aspiration counted. The concentrations of bound and free Fab were calculated, and a plot of bound against bound/free was used to calculate the affinities of the Fabs.
Fluorescence activated cell sorter (FACS) analysis

CHO-K1 cells (untransfected or expressing the TSHR) were grown to confluence in 175 cm² flasks in DMEM, 10% FCS at 37 °C and 5% CO₂. Flasks were washed two times with Dulbecco’s PBS minus calcium and magnesium ions (PBS(−)) then removed using a cell scraper into 10 ml PBS(−), washed two more times and resuspended using a needle and syringe. A total of 2 × 10⁵ cells were blocked with 100 µl 10% non-immunised rabbit sera in PBS(−) for 20 min at 4 °C, centrifuged at 200 g for 5 min at 4 °C and incubated with 100 µl IgG (100 mg/ml) in PBS(−) with 1% BSA for 30 min at 4 °C.

The cells were then washed three times with PBS(−) and incubated with 100 µl second antibody; rabbit anti-mouse Ig F(ab')₂ (Dako), FITC conjugated, diluted 1/40 in PBS(−) with 1% BSA for 30 min at 4 °C. The cells were washed three times with PBS(−) and fixed with 1% paraformaldehyde in PBS(−).

A total of 5000 cells were analysed for each sample using a Becton Dickinson FACSscan System and experiments were repeated on three separate occasions.

RESULTS

Reactivity of rabbit and mouse monoclonal antibodies in the IPA

Two rabbits were immunised with each of the TSHR constructs (TSHR400 and TSHR800) and tested every month. Non-immunised rabbit sera and sera from rabbits immunised with a GST/steroid 21-hydroxylase fusion protein (GST/21OH) (Chen et al. 1996) used as negative controls bound about 1–2% of 3⁵S-TSHR (full length) at 1/100 dilution (Table 1). In contrast, sera from rabbits immunised with TSHR800 and TSHR400 diluted 1/100 bound 42·5 and 15·1% of 3⁵S-TSHR respectively after 4–12 months of immunisation (Table 1). Five monoclonal antibodies were identified from three fusions (3F3, 3E4, 3C6, 5D8 and 3C7). All five reacted in the IPA, showing binding to 3⁵S-TSHR of between 7 and 46% (Table 2).

Precipitation of TSHR/TSH complexes

Rabbit antisera were used to immunoprecipitate recombinant human and porcine TSHRs complexed with ¹²⁵I-TSH (Table 3). Sera from rabbits immunised with TSHR400 and TSHR800 precipitated both human and porcine TSHR/¹²⁵I-TSH complexes. Sera from rabbits immunised with TSHR400 cross reacted strongly with the porcine TSHR complexes, 30·4% being bound at 1/10 dilution, compared with sera from rabbits immunised with TSHR800 which precipitated 16·2% at the same dilution and non-immunised rabbit sera which bound 12·7%. Sera from the TSHR800 rabbits reacted more strongly with the human TSHR than sera from the TSHR400 rabbits, 64% of the recombinant human TSHR complex was precipitated at 1/10 dilution with TSHR800 rabbit antiserum compared with 47·5% at the same dilution for TSHR400 rabbit antiserum and 8·3% for non-immunised rabbit sera (Table 3).

Western blot analysis of solubilised human TSHR

The five monoclonal antibodies (3C6, 3C7, 3F3, 3E4 and 5D8) reacted with detergent solubilised recombinant human TSHR on Western blot at 1/100 and 1/1000 dilution. Figure 1 shows the reaction of 3C7 TSHR monoclonal antibody (panels A and B); a TSHR monoclonal antibody directed to the C terminus of the receptor (T3–495; TRANSBIO, Boulogne, France) (panel C) and a negative control monoclonal antibody to glutamic acid decarboxylase (GAD6; DSHB, Iowa City, IA, USA) (panel D) with solubilised CHO-K1 (untransfected) membranes (lane 1), recombinant human TSHR (lane 2) and native human TSHR (lane 3). Analysis of solubilised CHO cells expressing the full length TSHR with 3C7 shows two bands at 117 ± 4 kDa (mean ± s.d.; n=5) and 99 ± 4 kDa (mean ± s.d.; n=5) and a broad band with the centre at 58 ± 2 kDa (mean ± s.d.; n=5).

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**Table 1. Binding of rabbit antibodies to ³⁵S-TSHR (full length).** Sera from rabbits immunised with TSHR800 and TSHR400 were tested by IPA based on ³⁵S TSHR (full length) produced in the TnT reaction. Results from the rabbits showing the best responses are shown in the table and are representative of at least five separate experiments carried out in duplicate. GST/21OH= control rabbit sera from rabbits immunised with GST/human steroid 21-hydroxylase fusion protein.

<table>
<thead>
<tr>
<th>Rabbit serum</th>
<th>Dilution</th>
<th>% ³⁵S-TSHR bound</th>
</tr>
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<tbody>
<tr>
<td>Non-immunised</td>
<td>1/10</td>
<td>2.6</td>
</tr>
<tr>
<td>1/100</td>
<td>1.2</td>
<td></td>
</tr>
<tr>
<td>GST/TSHR800</td>
<td>1/10</td>
<td>57.2</td>
</tr>
<tr>
<td>1/100</td>
<td>42.6</td>
<td></td>
</tr>
<tr>
<td>GST/TSHR400</td>
<td>1/10</td>
<td>41.2</td>
</tr>
<tr>
<td>1/100</td>
<td>15.1</td>
<td></td>
</tr>
<tr>
<td>GST/21OH</td>
<td>1/10</td>
<td>1.0</td>
</tr>
<tr>
<td>1/100</td>
<td>1.2</td>
<td></td>
</tr>
</tbody>
</table>
T3–495 antibody recognised bands of about 120 and 100 kDa in the CHO cells expressing the human TSHR. In the case of native TSHR from human tissue, the 3C7 monoclonal antibody reacted with proteins at 93 and 53 kDa whilst T3–495 antibody at a 1/1000 dilution did not show reactivity with native human TSHR. Neither 3C7 nor T3–495 reacted with the untransfected CHO-K1 cell preparations (Fig. 1, lanes 1 in panels A–C).

Figure 2 shows the reaction of the 3C7 TSHR monoclonal antibody (panel B) and 3C6 monoclonal antibody (panel A) with solubilised CHO-K1 (untransfected) membranes (lane 1), recombinant human TSHR (lane 2) and native human TSHR (lane 3). The 3C6 monoclonal showed a similar reactivity to 3C7 (Fig. 1, panels A, B; Fig. 2, panel B) with the two bands at 117 ± 4 kDa (mean ± s.d.; n=5) and 99 ± 1.5 kDa (mean ± s.d.; n=5) but differed from 3C7 in its reactivity to a broad band with the centre at 52 ± 1 kDa (mean ± s.d.; n=5). 3C6 showed no reaction with the A subunit of the native TSHR extracted from human thyroid tissue.

Antisera from both TSHR400 and TSHR800 immunised rabbits diluted 1/100 and analysed by Western blotting showed a similar pattern of reactivity to recombinant human TSHR as the monoclonal antibodies (3C6 and 3C7). In particular, bands at 119 ± 6 kDa (mean ± s.d.; n=5) and 102 ± 5 kDa (mean ± s.d.; n=5) and a broad band with the centre at 54 ± 1 kDa (mean ± s.d.; n=5) were recognised (data not shown).

### Inhibition of TSH binding to TSHR

Only one out of the 5 TSHR monoclonal antibodies (3C7) was able to inhibit 125I-TSH binding to native human TSHR (74% inhibition), recombinant
human TSHR (84% inhibition) and porcine TSHR
(65% inhibition) (Fig. 3).

This inhibiting effect was studied in more detail
using purified 3C7 IgG (Fig. 4). Concentrations of
3C7 IgG ranging from 0·03 to 2·5 mg/ml showed
125I-TSH binding inhibition ranging from 21 to
92% respectively. Furthermore, 3C7Fab was able to
inhibit TSH binding (41% inhibition at 0·15 mg/ml;
data not shown).

Scatchard analysis of anti TSHR Fab
fragments

The binding characteristics of two TSHR mono-
clonal antibody Fabs (3C7 and 3C6) to recombinant
human TSHR analysed by a Scatchard plot is
shown in Fig. 5. The affinity constant (Kₐ) was
8·9 × 10⁶ M⁻¹ for 3C7 and 1·3 × 10⁷ M⁻¹ for 3C6.

FACS analysis

Four of the monoclonal antibodies (3C6, 3F3, 3E4
and 5D8) bound to recombinant human TSHR

expressed in CHO-K1 cells in FACS analysis, whereas 3C7 did not appear to recognise the receptor on the cell surface (Fig. 6, Table 4). CHO-K1 cells were used as a negative control cell line and IgG purified from M1/21-OH ascites (RSR Ltd) was used as negative control IgG; Table 4 shows the percentage positive cells detected. The control IgG (M1/21-OH 10 µg per ml) showed similar low reactivity to CHO-K1 cells (untransfected) and CHO cells expressing TSHR (6·8 and 6·9% respectively). The TSHR monoclonal 3C7 IgG (10 µg per ml) also showed similar low reactivity to CHO-K1 cells (6·7%) and CHO cells expressing TSHR (8·6%). However, 3E4, 3C6, 5D8 and 3F3 did not bind to CHO-K1 cells (1·7–5·7% cells positive) but bound well (75·8–83·1% cells positive) to CHO cells expressing the TSHR (Fig. 6; Table 4).
DISCUSSION

The TSHR is an important autoantigen in Graves’ disease and the experimental production of antibodies to the receptor is critical to our understanding of the receptor’s structure and function and how it interacts with TSH and TRAb. However, the receptor is present in small amounts on the cell surface and has proved difficult to purify in large enough quantities to immunise experimental animals. Therefore we used two GST–TSHR fusion proteins produced in E.coli for immunisation and have produced rabbit polyclonal antibodies and

![Figure 3](image1.png)  
**Figure 3.** Inhibitory effect of antibodies on $^{125}$I-TSH binding to native human TSHR (a), recombinant human TSHR (b) and native porcine TSHR (c). ■ = TRAb in patient serum; ● = 3C7 TSHR monoclonal antibody (ascites) and ○ = GAD$_{65}$ monoclonal antibody (ascites). Results shown in the figure are representative of at least two separate experiments carried out in duplicate.

![Figure 4](image2.png)  
**Figure 4.** Inhibition of $^{125}$I-TSH binding to recombinant human TSHR by purified 3C7 TSHR IgG (■), 3C6 TSHR IgG (○) and M1/21OH IgG (●). Results shown in the figure are representative of at least two separate experiments carried out in duplicate.

![Figure 5](image3.png)  
**Figure 5.** Binding of TSHR antibody Fab fragment to recombinant human TSHR expressed in CHO-K1 cells. 3C6 TSHR Fab (■), 3C7 TSHR Fab (●).
a panel of five monoclonal antibodies to the extracellular domain of the human TSHR.

The rabbit polyclonal antibodies raised to both TSHR constructs (TSHR400 and TSHR800) reacted in an immunoprecipitation assay with full length \(^{35}\)S-TSHR; highest titres were produced by sera from the rabbits immunised with the longer (TSHR800) fragment.

Rabbit polyclonal antibodies were able to immunoprecipitate \(^{125}\)I-TSH/TSHR complexes of both human and porcine TSHR. Sera from rabbits immunised with TSHR400 reacted strongly with the human and porcine TSHR/\(^{125}\)I-TSH complexes compared with sera from rabbits immunised with TSHR800 which reacted only with human TSHR/\(^{125}\)I-TSH complexes. These differences in crossreactivity with human and porcine receptors suggest that the major epitopes recognised by sera from TSHR800 rabbits are located in regions which are different between human and porcine TSHR sequences. In contrast, sera from rabbits immunised with TSHR400 recognised some epitopes that are common for both human and porcine TSHR sequences (there is an approximate 70% sequence identity at the amino acid level of the known TSHR extracellular domain sequences between different species) (Furmaniak & Rees Smith 1993).

We were not successful in precipitating either recombinant human or porcine TSHR complexes with any of the monoclonals we produced. The reason for this is unclear but may reflect the relatively low affinities of the monoclonal antibodies. In addition, polyclonal antibodies are generally considered to be more suitable for immunoprecipitation than low affinity monoclonal antibodies (Harlow & Lane 1988).

Western blot analysis of the TSHR with 3C7 showed the recombinant full length receptor to be present as two bands of 117 and 99 kDa and a broad 58 kDa protein band representing the receptor A subunit, whilst analysis with 3C6 showed the two full length bands at 117 and

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**Figure 6.** FACS analysis of monoclonal IgG binding to untransfected CHO-K1 cells (CHO-K1) or to CHO-K1 cells expressing the TSHR (CHO-TSHR). (The percentages of positive cells for this experiment are shown in Table 4.) 21-OH IgG: M1/21-OH monoclonal antibody to steroid 21-hydroxylase; 3C6, 3F3, 3E4, 5D8, 3C7: monoclonal antibodies to the TSHR.
99 kDa but the receptor’s A subunit was present as a smaller broad band of 52 kDa. Since both of the antibodies recognise the receptor’s extracellular domain these results suggest the presence of two possible cleavage sites in the receptor leading to the variations of the A subunit molecular mass. Similar observations regarding the presence of two cleavage sites in the TSHR have recently been reported by Chazenbalk et al. (1997).

Analysis of the sugar residues on the TSHR has suggested that the upper band (117 kDa) of the full length TSHR represents fully glycosylated ‘matured’ TSHR while the lower band (99 kDa) represents a high mannose precursor (Costagliola et al. 1994, Johnstone et al. 1994, Chazenbalk et al. 1996, Couet et al. 1996a,b, Sanders et al. 1997). In our studies of native human TSHR, a small amount of the native receptor was present as a single band at 93 kDa whereas most of the receptor was detected by 3C7 at a molecular mass of 53 kDa representing the A subunit (Kajita et al. 1985a,b, Rees Smith et al. 1988, Loosfelt et al. 1992). 3C6 did not react well with the native TSHR extracted from human thyroid tissue in Western blotting analysis and the reason for this is not clear at present. The differences in molecular mass between native and recombinant human TSHRs may be due in part at least to differences in the extent of glycosylation. The T3–495 monoclonal antibody reacted with the 117 and 99 kDa bands of recombinant human TSHR but did not react with bands with molecular weights characteristic of the receptor’s A or B subunits (at 1/1000 dilution). Neither 3C7 (diluted 1/1000) nor T3–495 (diluted 1/1000) antibodies reacted with the full length native receptor in Western blotting; this was most probably related to the lower concentration of this form of the receptor in native receptor preparations. 3C7 was produced in response to immunisation with the extracellular region of the TSHR while the antibody T3–495 was produced using the C-terminus of the receptor (Loosfelt et al 1992). Consequently, in our studies the reactivity of T3–495 with the full length receptor but not the A subunit would be expected. We were unable to detect any reactivity between T3–495 and the receptor B subunit in native or recombinant TSHR preparations but this could be due to a relatively low concentration of the B subunit.

Western blotting analysis showed that the recombinant receptor was present as both full length receptor and the A subunit whereas the native receptor extracted from thyroid tissue was present predominantly as the A subunit (Fig. 1). The differences in the amount of cleaved and uncleaved receptors present in recombinant and native TSHR preparations could be due to the difficulties in preventing rapid protease cleavage of the receptor in native thyroid tissue removed at operation whilst it is relatively simple to add protease inhibitors to cultured cells expressing the receptor as soon as they are harvested. Experimental evidence has suggested that the cleavage of the full length receptor in cultured L cells and thyroid cells may be dependant on a matrix metalloprotease (Couet et al. 1996a,b). In addition, recent studies have indicated that the extracellular domain may be cleaved in two positions giving rise to two different A subunit forms (Chazenbalk et al. 1997).

We were unable to detect inhibition of TSH binding to either human or porcine TSHRs with the rabbit polyclonal antibodies in contrast to other reports (Dallas et al. 1994, Harfst et al. 1994). Only one of our five monoclonal antibodies (3C7) was found to inhibit binding of 125I-TSH to recombinant human, native human and porcine TSHR preparations very effectively in the TRAb assay with a dilution profile similar to that seen for patient sera. The affinity of this antibody (3C7) was similar to that of 3C6 which did not inhibit TSH binding and this suggested that the differences in properties were due to differences in the epitopes recognised. Other laboratories have produced mouse monoclonals which appear to inhibit TSH binding but to a much lesser extent than 3C7 (Johnstone et al. 1994, Seetharamaiah et al. 1995, Nicholson et al. 1996). Consequently, this study and other studies show that to date, most mouse monoclonal antibodies raised to the receptor do not inhibit TSH binding (Johnstone et al. 1994, Seetharamaiah et al. 1995, Nicholson et al. 1996). Furthermore, none of the monoclonal antibodies we have produced had the ability to stimulate cAMP production in CHO–TSHR cells (Nagayama et al. 1989) (data not shown). This is consistent with other reports in the literature (Johnstone et al. 1994, Seetharamaiah et al. 1995, Nicholson et al. 1996)

**Table 4. FACS analysis of the binding of monoclonal antibody IgGs to intact cells.** The percentage positive cells for each IgG with both CHO-K1 cells and CHO-K1 cells expressing the TSHR are shown. Results are means ± s.d. of three separate experiments.

<table>
<thead>
<tr>
<th>Monoclonal antibody</th>
<th>CHO-K1/TSHR positive cells (%)</th>
<th>CHO-K1 positive cells (%)</th>
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<tr>
<td>M1/21OH IgG</td>
<td>6.9 ± 3.2</td>
<td>6.8 ± 1.4</td>
</tr>
<tr>
<td>3C6 IgG</td>
<td>86.7 ± 1.5</td>
<td>1.7 ± 0.9</td>
</tr>
<tr>
<td>3F3 IgG</td>
<td>89.1 ± 1.4</td>
<td>5.4 ± 1.0</td>
</tr>
<tr>
<td>3E4 IgG</td>
<td>75.8 ± 13.8</td>
<td>2.8 ± 1.6</td>
</tr>
<tr>
<td>5D8 IgG</td>
<td>88.6 ± 2.0</td>
<td>5.7 ± 1.7</td>
</tr>
<tr>
<td>3C7 IgG</td>
<td>8.6 ± 1.5</td>
<td>6.7 ± 1.2</td>
</tr>
</tbody>
</table>

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and overall current studies indicate that it is difficult to produce TSHR mouse monoclonal antibodies with thyroid stimulating activity. However, newer approaches using DNA immunisation may be successful (Costagliola 1996, Shimojo et al. 1996).

Purification of 3C7 IgG showed that a relatively high concentration (2·5 mg/ml) was needed to obtain over 90% inhibition of TSH binding. These results implied that the affinity of this monoclonal antibody was considerably lower than that of TSH. Consequently, we purified Fab fragments from both 3C7 and 3C6 monoclonal antibodies to investigate their binding to TSHR. The results of Scatchard analysis showed that 3C7 and 3C6 have affinities of 8·9 × 10⁹ and 1·3 × 10⁷ M⁻¹ respectively, approximately 1000 times lower than that of TSH (1 × 10¹⁰ M⁻¹); (data not shown). These results are consistent with the high concentrations of 3C7 IgG needed to inhibit TSH binding. However, our data suggest that 3C7 binds at or close to part of the TSH binding site either inhibiting TSH binding directly by binding to the TSH binding site or indirectly by steric hindrance and/or altering the conformation of the receptor preventing TSH binding.

Four of the five antibodies tested in the FACS analysis (3C6, 3F3, 3E4 and 5D8) bound to CHO-K1 cells expressing the human TSHR on their cell surface. Surprisingly 3C7, the antibody which shows inhibition of TSH binding to solubilised TSHR, did not appear to react with receptors on intact cells as judged by FACS analysis. This suggests that the epitope for 3C7 only becomes available after the receptors have been extracted from the cells and/or solubilised with detergent. The reason for this discrepancy is not clear at present but studies with native TSH receptors have shown that TSH binding affinity is increased markedly when the receptor is solubilised in non-ionic detergents (Brennan et al. 1980). This effect on TSH binding may be the result of subtle changes in the three dimensional structure of the receptor induced by detergent solubilisation. A similar effect of detergent solubilisation on the region of the receptor involved in 3C7 binding might explain the differences in 3C7 reactivity between cell surface and solubilised TSHR. This observation emphasises the importance of using a combination of FACS analysis and radioactively labelled probes in analysis of the receptor using experimentally produced antibodies.

Overall our studies indicate that the ³⁵S-labelled TSH receptor produced in the TnT system is very useful in screening and characterising experimentally produced antibodies to the TSH receptor. Using this screening technique we have been able to select the first mouse monoclonal antibody which competes effectively with labelled TSH binding to detergent solubilised TSH receptors.

The monoclonal antibodies produced in this study using fusion proteins expressed in E. coli were of relatively low affinity. However, experimentally produced antibodies with the high affinities characteristic of TSH and TRAbs are potentially the most interesting but these have not been described as yet. Successful production of such antibodies is likely to be related to the availability of highly purified, correctly folded TSHR preparations for immunisation or use of techniques such as DNA immunisation (Costagliola 1996).

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REFERENCES


Chen C & Okayama H 1987 High efficiency transformation of mammalian cells by plasmid DNA. Molecular and Cellular Biology 7 2745–2752.


Dallas JS, Desai RK, Cunningham SJ, Morris JC, Gattadahalli SS, Wagle N, Goldblum RM & Prabhakar BS 1994 Thyrotropin (TSH) interacts with multiple discrete regions of the TSH receptor: polyclonal rabbit antibodies to one or more of these regions can inhibit TSH binding and function. Endocrinology 134 1437–1445.

Davies Jones E & Rees Smith B 1984 A water-soluble fragment of the thyroid-stimulating hormone receptor which binds both thyroid-stimulating hormone and thyroid-stimulating hormone receptor antibodies. Journal of Endocrinology 100 113–118.


immunisation with fibroblasts transfected with the thyrotropin receptor and a class II molecule. *Proceedings of the National Academy of Sciences of the USA* 93, 11074–11079.


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